Ethanol production using vegetable peels medium and the effective role of cellulolytic bacterial (Bacillus subtilis) pre-treatment [version 1; referees: 2 approved with reservations]

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Abstract

Background: The requirement of an alternative clean energy source is increasing with the elevating energy demand of modern age. Bioethanol is considered as an excellent candidate to satiate this demand.

Methods: Yeast isolates were used for the production of bioethanol using cellulosic vegetable wastes as substrate. Efficient bioconversion of lignocellulosic biomass into ethanol was achieved by the action of cellulolytic bacteria (Bacillus subtilis). After proper isolation, identification and characterization of stress tolerances (thermo-, ethanol-, pH-, osmo- & sugar tolerance), optimization of physiochemical parameters for ethanol production by the yeast isolates was assessed. Very inexpensive and easily available raw materials (vegetable peels) were used as fermentation media. Fermentation was optimized with respect to temperature, reducing sugar concentration and pH.

Results: It was observed that temperatures of 30°C and pH 6.0 were optimum for fermentation with a maximum yield of ethanol. The results indicated an overall increase in yields upon the pretreatment of Bacillus subtilis; maximum ethanol percentages for isolate SC1 obtained after 48-hour incubation under pretreated substrate was 14.17% in contrast to untreated media which yielded 6.21% after the same period. Isolate with the highest ethanol production capability was identified as members of the ethanol-producing Saccharomyces species after stress tolerance studies and biochemical characterization using Analytical Profile Index (API) ® 20C AUX and nitrate broth test. Introduction of Bacillus subtilis increased the alcohol production rate from the fermentation of cellulosic materials.

Conclusions: The study suggested that the kitchen waste can serve as an excellent raw material in ethanol fermentation.

Keywords

Bioethanol, yeast, cellulolytic bacteria
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Abbreviations
SC1 - yeast isolates from sugarcane juice; DJ1 - yeast isolates from date juice; pH - Negative logarithm of hydrogen ion concentration; °C - Degree Celsius; % - Percentage; CH₂CH₂OH - ethanol or ethyl alcohol; YEPD - Yeast Extract Peptone Dextrose; nm - Nanometer; gm - Gram; ml - Milliliter; rpm - Round per minute; v/v - volume per volume; w/v - weight per volume; spp. - Species; et al. - And others.

Introduction
Fermentation-derived ethanol (CH₂CH₂OH) or ethyl alcohol is commonly known as bioethanol. Ethanol can be produced chemically from petroleum, and from biomass or sugar substrates fermentation. This organic chemical is a flammable, clear and colorless liquid which can be used as fuel. Other functions of ethanol include its use as a solvent, antifreeze and germicide. With the aims of protecting the environment and reducing dependence on petroleum and nonrenewable energy sources, the development of renewable energy sources has become increasingly important. Several processes of bioethanol production currently exist, such as microbiological production from fermentable organic substrates or carbohydrates by yeast. Fermentation of cellulosic biomass, molasses, vegetable peels or food wastes can be considered as an economical process of bioethanol production. Bioethanol produced from cellulosic materials by direct conversion is utilized in countries such as Brazil, Canada and, USA. The economical production of bioethanol requires an easily available supply of inexpensive raw materials. Organic food waste is one of the topmost suitable materials for that process. Solid food wastes from household, restaurants or food processing industries can be obtained as a substrate to be used as fermentation medium for bioethanol production. Food wastes can also be recycled as animal feed and fertilizer after specific treatment.

The foremost focus of this ethanol production technology is the optimized utilization of biomass resources and microbial action on fermentation. One promising technique is the fermentation of lignocellulosic biomass where hydrolysis by specific microbial cellulase enzymes is involved. Ethanol can be derived from the fermentation of sugar-containing materials. Different yeast varieties are reported for the fermentation of lignocellulosic substrates to produce ethanol.

The objective of the project is to establish a highly efficient microbial fermentation process by natural yeast isolates to produce ethanol. It is to be mentioned that ethanol production rate from insoluble lignocellulosic biomass is currently not economical. Therefore, commonly available cellulosic kitchen wastes were used as raw material. Proper treatment of the substrate was done to optimize the fermentation condition which has resulted in a highly efficient and economical production rate. Potential wild-type yeast strains were isolated from date juice, sugarcane juice, grapes, and pineapples. Wild-type yeasts were identified by the biochemical and physiological characterization and taken under comparative studies and experiments to obtain a strain with high productivity. Cellulose degrading bacteria (Bacillus subtilis) was used for pre-treatment of the fermentation media. Cellulolytic microorganism debased celluloses present in the lignocellulosic fermentation media and the degraded materials were easier and more readily available to be fermented by yeast.

Methods
Sample collection and isolation
Wild-type yeasts were isolated from sugarcane juice and date juice. Aforementioned sources were collected from the local market and kept for 1 week at room temperature for yeast growth. The samples were inoculated into YEPD (Yeast Extract Peptone Dextrose) broth which is composed of 1% yeast extract (Y1625), 2% peptone (P7750), 2% glucose or dextrose (G8270) (Sigma-Aldrich, St Louis, MO, USA) and the desired volume of distilled water. Cultures from the broth were plated on YEPD agar media and incubated at 37°C for 48 hours. The grown colonies were cultured again on YEPD agar medium under the same growth condition to obtain isolated colonies. After the incubation, the isolated colonies (slant) were preserved at 4°C refrigeration. The culture was maintained by periodic sub-culturing.

Identification
A compound microscope (Model-CX-21, Olympus, Japan) was used to observe the cell morphology and the presence of yeasts were confirmed which were isolated from sugarcane juice (Figure 1A) and date juice (Figure 1B) (named SC1 and DJ1 respectively). Identification of each isolate of yeast up to species level was carried by the methods demonstrated by Kreger-Van Rij (1984) based on the morphology, sporulation and fermentation characteristics, as well as the assimilation of nitrogen and a range of carbon sources. Yeast specific API® identification kit (bioMérieux, Marcy-l’Étoile, France) was also used by inoculating 48-hrs culture broths into the chambers according to the protocol.

Stress tolerance tests
Ethanol tolerance of yeast isolates was tested by inoculating isolates in YEPD broth supplemented with varying concentrations (5%, 10%, 15% and 20%) of absolute ethanol and incubated at 30°C for 48 hours. To observe the thermostolerance, the isolates were inoculated in YEPD broth and incubated at different temperatures (25°C, 30°C, 37°C and 44°C) for 48 hours. The growth of the yeast isolates at different pH was observed by inoculating isolates in YEPD broth with different pH (2–10; adjusted by adding drops of basic NaOH or acidic diluted HCl in the solution while reading a pH meter (E-201-C Shanghai Ruouhua Technology company, China)) and incubated at 30°C for 48 hours. Initial optical densities of each tube during inoculation and optical densities after incubation were measured using the spectrophotometer (UVmini-1240 spectrophotometer, Shimadzu, Kyoto, Japan) at 600 nm against the medium as the blank.
by inoculating 24 hours’ old culture into 10 ml NaCl (0.85%) saline. The suspensions were made homogenous using vortex machine after inoculation.

**Fermentation of cellulosic media**

150 ml fermentation media was added to 500 ml Erlenmeyer conical flasks. Cellulosic media were aseptically inoculated with *Bacillus subtilis* suspension and incubated for 24 hours at 37°C in shaking condition (80 rpm). After the incubation, yeast cell suspension was inoculated and the flasks were cotton plugged and incubated in a rotary incubator (WIS-20R, Wonju-si, Daihan Scientific, Korea) at 30°C for 48 hours in shaking condition (120 rpm). Yeast isolates were inoculated into another set of similar cellulosic media which were not treated with the cellulolytic organism and incubated under the aforementioned fermentation condition.

**Estimation of ethanol**

Initial assay of ethanol production rates was performed by volumetric analysis in Conway units. A fractional distillation set was used to separate ethanol from fermented broths. Samples yielding feasible results were distilled and the ethanol percentages of the distillates were determined by specific gravity using an alcohol meter (5453 Vinometer, LD Carlson, Kent, OH, United States).

**Results**

Morphology was visually observed as white and creamy texture, ovoid shape, multipolar budding pattern, under microscope (Figure 1A and B). Sporulation was confirmed due to the presence of ascospore. Nitrate reduction was not exhibited in the nitrate assimilation test (Figure 2A). Carbohydrate assimilation tests were conducted using API® 20C test strips (bioMérieux, Marcy-l’Étoile, France) (Figure 2B). In all cases, positive results were obtained for glucose, galactose, maltose, starch, and fructose (Table 1). Therefore, carbohydrates and nitrate assimilation test results signified the strong probability of isolates being the species *Saccharomyces cerevisiae*. Yeast isolates from sugarcane juice (SJ1) had a good growth at 25°C, 30°C, and 37°C, but showed poor growth at 40°C and 44°C. Yeast isolates from date juice (DJ1) had a good growth at 30°C, and 37°C, moderate growth at 40°C, but propagated poorly at 25°C and 44°C. Yeast isolate SC1 and DJ1 showed a variable growth result at pH 2–10. Overall, pH 5 and 6 was optimum growth conditions where the isolate SC1 exhibited the highest growth at pH 6 and DJ1 had its best growth at pH 5. All isolates showed excellent growth at 5% and 10% ethanol concentrations throughout the entire 48-hour incubation period (Table 2).

With fermentation conditions of 30°C incubation temperature with a pH of 6, the highest rate of alcohol production from a cellulosic medium (a mixture of papaya and potato peels pretreated with *Bacillus subtilis*) was 14.17% v/v or 141.7 gm/L (w/v).
Table 1. Fermentation result of different carbohydrates for sugarcane juice (SC1) and date juice (DJ1) isolate. (Legends: ++ Positive, + Variable, -- Negative).

<table>
<thead>
<tr>
<th>Carbohydrate</th>
<th>SC1</th>
<th>DJ1</th>
</tr>
</thead>
<tbody>
<tr>
<td>Glucose</td>
<td>++ (gas)</td>
<td>++ (gas)</td>
</tr>
<tr>
<td>Maltose</td>
<td>++ (gas)</td>
<td>++ (gas)</td>
</tr>
<tr>
<td>Galactose</td>
<td>++ (gas)</td>
<td>++ (gas)</td>
</tr>
<tr>
<td>Starch</td>
<td>+ +</td>
<td>+ +</td>
</tr>
<tr>
<td>Sucrose</td>
<td>+ -</td>
<td>- -</td>
</tr>
<tr>
<td>Fructose</td>
<td>+ + (gas)</td>
<td>+ + (gas)</td>
</tr>
<tr>
<td>Trehalose</td>
<td>- -</td>
<td>- -</td>
</tr>
<tr>
<td>Lactose</td>
<td>- -</td>
<td>- -</td>
</tr>
<tr>
<td>Xylose</td>
<td>- -</td>
<td>- -</td>
</tr>
</tbody>
</table>

Figure 2. Biochemical tests to identify yeast species. (A) Negative nitrate reduction test indicated by the yellow color (positive test would result in red color change) (B) API 20 C X kit results for different carbohydrates fermentation using API kit after 48 hours. Color in the chamber indicates a positive result, negative results retain the yellow color of the broth. Active ingredients contained in each chamber from left to right: 0 – none, 1 – D-glucose, 2 – Glycerol, 3 – Calcium 2-keto-D-gluconate, 4 – L-arabinose, 5 – D-xylose, 6 – Adonitol, 7 – Xylitol, 8 – D-galactose, 9 – Inositol, 10 – D-sorbitol, 11 – Methyl α-D-glucopyranoside, 12 – N-acetylglucosamine, 13 – D-cellobiose, 14 – D-lactose, 15 – D-maltose, 16 – D-saccharose/Sucrose, 17 – D-trehalose, 18 – D-melezitose, 19 – D-raffinose.

by yeast isolate SC1 (Figure 3A; Dataset 1). On the other hand, under the same fermentation conditions, the highest rate of alcohol production using the same cellulosic medium not treated with cellulolytic bacteria was 6.21% v/v or 62.1 gm/L (w/v) by isolate DJ1 (Figure 3A; Dataset 1) under a 48-hour fermentation condition at 30°C incubation temperature. The lowest alcohol production rate recorded under the same conditions using the untreated potato and cucumber media (2.15% v/v) by the isolate DJ1 (Figure 3B; Dataset 2). Fermented media with the highest percentage of alcohol (14.17% v/v) was distilled by a fractional distillation set. This highest percentage was achieved by the yeast isolate SC1 in fermentations condition of 30°C, pH 6 at 120 rpm. The cellulosic media pretreated with Bacillus subtilis was distilled after fermentation and the distilled product (one-time distillation) had an ethanol percentage of 52% v/v. In contrast, cellulosic media which was not treated with Bacillus subtilis had an ethanol percentage of 12% v/v after the first distillation. Fermentation in other media was recorded highest at 6.23% or 62.3 gm/L (w/v) alcohol production at pH 6 by SC1 isolate after 24-hrs fermentation (Table 3).

Dataset 1. Alcohol production from vegetable peels by yeast isolates SC1 and DJ1 at pH 6. Media pre-treated with Bacillus subtilis
http://dx.doi.org/10.5256/f1000research.13952.d195639

Dataset 2. Alcohol production from vegetable peels by yeast isolates SC1 and DJ1 at pH 6. Media without Bacillus subtilis pre-treatment
http://dx.doi.org/10.5256/f1000research.13952.d195640
Table 2. Alcohol production in different physical/chemical condition after 48 hours of fermentation. (Legends: + + Positive, + - Variable, -- Negative).

<table>
<thead>
<tr>
<th>Isolate</th>
<th>Temperature</th>
<th>48 hours growth</th>
<th>Initial O.D.</th>
<th>Final O.D.</th>
<th>O.D. change</th>
<th>Growth</th>
</tr>
</thead>
<tbody>
<tr>
<td>SC1</td>
<td>25°C</td>
<td></td>
<td>0.244</td>
<td>1.184</td>
<td>0.94</td>
<td>+ -</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.529</td>
<td>1.401</td>
<td>0.872</td>
<td>+ -</td>
</tr>
<tr>
<td>SC1</td>
<td>30°C</td>
<td></td>
<td>0.112</td>
<td>1.540</td>
<td>1.428</td>
<td>+ +</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.441</td>
<td>1.103</td>
<td>1.544</td>
<td>+ +</td>
</tr>
<tr>
<td>SC1</td>
<td>37°C</td>
<td></td>
<td>0.311</td>
<td>1.42</td>
<td>1.109</td>
<td>+ +</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.525</td>
<td>1.436</td>
<td>0.911</td>
<td>+ -</td>
</tr>
<tr>
<td>SC1</td>
<td>40°C</td>
<td></td>
<td>0.368</td>
<td>0.524</td>
<td>0.156</td>
<td>+ -</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.463</td>
<td>0.754</td>
<td>0.291</td>
<td>+ -</td>
</tr>
<tr>
<td>SC1</td>
<td>44°C</td>
<td></td>
<td>0.513</td>
<td>0.493</td>
<td>-0.02</td>
<td>- -</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.685</td>
<td>0.532</td>
<td>-0.153</td>
<td>- -</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Isolate</th>
<th>Ethanol percentage</th>
<th>48 hours growth</th>
<th>Initial O.D.</th>
<th>Final O.D.</th>
<th>O.D. change</th>
<th>Growth</th>
</tr>
</thead>
<tbody>
<tr>
<td>SC1</td>
<td>5%</td>
<td></td>
<td>0.103</td>
<td>1.476</td>
<td>1.371</td>
<td>+ +</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.172</td>
<td>1.555</td>
<td>1.385</td>
<td>+ +</td>
</tr>
<tr>
<td>SC1</td>
<td>10%</td>
<td></td>
<td>0.261</td>
<td>0.488</td>
<td>0.227</td>
<td>+ +</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.338</td>
<td>0.592</td>
<td>0.254</td>
<td>+ +</td>
</tr>
<tr>
<td>SC1</td>
<td>15%</td>
<td></td>
<td>0.201</td>
<td>0.314</td>
<td>0.113</td>
<td>+ +</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.097</td>
<td>0.151</td>
<td>0.054</td>
<td>+ -</td>
</tr>
<tr>
<td>SC1</td>
<td>20%</td>
<td></td>
<td>0.075</td>
<td>0.109</td>
<td>0.034</td>
<td>- -</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.191</td>
<td>0.254</td>
<td>0.063</td>
<td>- -</td>
</tr>
<tr>
<td>SC1</td>
<td>25%</td>
<td></td>
<td>0.218</td>
<td>0.167</td>
<td>-0.051</td>
<td>- -</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.257</td>
<td>0.11</td>
<td>-0.147</td>
<td>- -</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Isolate</th>
<th>pH</th>
<th>48 hours growth</th>
<th>Initial O.D.</th>
<th>Final O.D.</th>
<th>O.D. change</th>
<th>Growth</th>
</tr>
</thead>
<tbody>
<tr>
<td>SC1</td>
<td>2</td>
<td></td>
<td>0.186</td>
<td>0.066</td>
<td>-0.12</td>
<td>- -</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.201</td>
<td>0.009</td>
<td>-0.192</td>
<td>- -</td>
</tr>
<tr>
<td>SC1</td>
<td>3</td>
<td></td>
<td>0.303</td>
<td>1.13</td>
<td>0.827</td>
<td>- -</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.335</td>
<td>1.276</td>
<td>0.941</td>
<td>+ -</td>
</tr>
<tr>
<td>SC1</td>
<td>4</td>
<td></td>
<td>0.390</td>
<td>1.249</td>
<td>0.859</td>
<td>- -</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.409</td>
<td>1.44</td>
<td>1.031</td>
<td>+ -</td>
</tr>
<tr>
<td>SC1</td>
<td>5</td>
<td></td>
<td>0.179</td>
<td>1.412</td>
<td>1.233</td>
<td>+ +</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.164</td>
<td>1.472</td>
<td>1.308</td>
<td>+ +</td>
</tr>
<tr>
<td>SC1</td>
<td>6</td>
<td></td>
<td>0.108</td>
<td>1.536</td>
<td>1.428</td>
<td>+ +</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.377</td>
<td>1.846</td>
<td>1.469</td>
<td>+ +</td>
</tr>
<tr>
<td>SC1</td>
<td>7</td>
<td></td>
<td>0.145</td>
<td>1.337</td>
<td>1.192</td>
<td>+ -</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.452</td>
<td>1.441</td>
<td>0.989</td>
<td>- -</td>
</tr>
<tr>
<td>SC1</td>
<td>8</td>
<td></td>
<td>0.356</td>
<td>1.53</td>
<td>1.174</td>
<td>+ -</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.424</td>
<td>1.572</td>
<td>1.148</td>
<td>+ -</td>
</tr>
<tr>
<td>SC1</td>
<td>9</td>
<td></td>
<td>0.243</td>
<td>1.391</td>
<td>0.737</td>
<td>- -</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.384</td>
<td>1.264</td>
<td>0.880</td>
<td>- -</td>
</tr>
<tr>
<td>SC1</td>
<td>10</td>
<td></td>
<td>0.433</td>
<td>0.398</td>
<td>-0.035</td>
<td>- -</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td></td>
<td>0.528</td>
<td>0.522</td>
<td>-0.006</td>
<td>- -</td>
</tr>
</tbody>
</table>
Figure 3. Alcohol production from vegetable peels by yeast isolates SC1 and DJ1 at pH 6. Media treated with Bacillus subtilis (A) and No pre-treatment (B).

Table 3. Alcohol production from defined sugars by yeast isolate SC1 and DJ1.

<table>
<thead>
<tr>
<th>Isolate</th>
<th>Defined sugar medium</th>
<th>pH</th>
<th>Percentage of ethanol after 24 hours</th>
<th>Avg. gm/L (w/v)</th>
<th>Percentage of ethanol after 48 hours</th>
<th>Avg. gm/L (w/v)</th>
</tr>
</thead>
<tbody>
<tr>
<td>SC1</td>
<td>Glucose</td>
<td>6</td>
<td>2.28 2.11 2.24</td>
<td>22.4</td>
<td>2.37 2.31 2.34</td>
<td>23.4</td>
</tr>
<tr>
<td></td>
<td></td>
<td>5</td>
<td>3.35 3.33 3.34</td>
<td>23.4</td>
<td>2.99 2.93 2.96</td>
<td>29.6</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td>6</td>
<td>1.91 1.87 1.89</td>
<td>18.9</td>
<td>1.98 1.9 1.94</td>
<td>19.4</td>
</tr>
<tr>
<td></td>
<td></td>
<td>5</td>
<td>3.91 3.67 3.79</td>
<td>37.9</td>
<td>3.87 3.81 3.84</td>
<td>38.4</td>
</tr>
<tr>
<td>SC1</td>
<td>Sucrose</td>
<td>6</td>
<td>2.22 2.12 2.17</td>
<td>21.7</td>
<td>2.23 2.15 2.19</td>
<td>21.9</td>
</tr>
<tr>
<td></td>
<td></td>
<td>5</td>
<td>2.71 2.67 2.69</td>
<td>26.9</td>
<td>2.81 2.73 2.77</td>
<td>27.7</td>
</tr>
<tr>
<td>DJ1</td>
<td></td>
<td>6</td>
<td>1.74 1.68 1.71</td>
<td>17.1</td>
<td>1.79 1.81 1.80</td>
<td>18</td>
</tr>
<tr>
<td></td>
<td></td>
<td>5</td>
<td>2.21 2.13 2.17</td>
<td>21.7</td>
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<td>4.95 4.91 4.93</td>
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Discussion
Despite the availability of several industrial strains of yeasts, local isolates are usually more adapted to their own climatic condition. In this study, yeasts were isolated from local resources. Utilization of isolated yeasts is an important strategy for the production of bioethanol. On the basis of the white and creamy appearance of selected isolates on solid media with butyrous colony texture, polar budding and oval cellular shape it can be assumed that isolates are members of Saccharomyces spp. from the method described by Boekhout and Kurtzman (1996). Fermentation of different sugars by the selected yeast isolates was observed. Yeast isolates from sugarcane (SC1) utilized glucose, maltose, fructose, galactose, starch, sucrose, and arabinose but failed to grow on sorbitol, melibiose, mannitol, trehalose, inositol, xylose and lactose. Yeast isolates from date juice (DJ1) utilized glucose, maltose, fructose, galactose and starch, but failed to grow in trehalose, xylose, sucrose and lactose.

The conclusion was further reinforced by biochemical tests performed using bioMérieux’ API® 20C kits. Kit results for SC1 and DJ1 indicated that all of the isolates are Saccharomyces cerevisiae. As previous studies by Ramani et al. (1998) indicate that API® 20C kits have a statistical accuracy of 97% for common yeasts, the conclusions were assumed to be correct. Furthermore, nitrate assimilation tests for all isolates yielded negative results which confirming our hypothesis. Thermotolerance tests also indicated that all isolates (SC1 and DJ1) grew best at 30°C within a 48-hour incubation period; this is also the optimum growth temperature of Saccharomyces cerevisiae described by Alexopoulos (1962). As for ethanol tolerance, the general trend observed was a decrease in terms of tolerance of all isolates above 10% ethanol concentration signified by a slowdown in growth rate with a near growth stunt at 20%. Teramoto et al. (2005) demonstrated that members of Saccharomyces spp. can tolerate ethanol concentrations of up to 16.5%. However, since the isolates are
wild-type Saccharomyces yeasts, an average maximum tolerance of 10% ethanol does not mean that they cannot be members of Saccharomyces spp. Different growth factors affect the pH tolerance of yeast. It was reported by Ivorra et al. (1999)\(^\text{9}\) that the optimum pH range for ideal growth varies from 4–6 depending on the strain. The cellular structure of yeast has a diverse mechanism to endure pH. In this experiment, yeast isolates SC1 and DJ1 had a variable growth result from pH 2–10. Both of the isolates had excellent growth from pH 4 to 6. However, those isolates were able to grow at all the pH condition, but pH lower than 3 and higher than 7 was not suitable for a good growth. Overall, pH 5 and 6 were optimum growth conditions where isolate SC1 had its best growth at pH 6 and isolate DJ1 had its best growth at pH 5. The ethanol production rate was recorded from the fermentation of different cellulosic media after 24 and 48-hours fermentation. The production rate ranged from 2.15% or 21.5 gm/L to 14.17% v/v or 141.7 gm/L (w/v). Isolate SC1 had the highest rate of ethanol production (14.17% v/v), and isolate DJ1 had the lowest rate of ethanol production (2.15% v/v) in shaking condition at 30°C with a media pH of 6. Ethanol production rate was also observed in shaking condition at 30°C with a media pH of 5. In this condition, isolate SC1 had the highest rate of ethanol production (9.42% v/v) and isolate DJ1 had the lowest rate of ethanol production (2.17% v/v), which surpassed the previous reports\(^\text{23,24}\). Ethanol production using kitchen waste media has exceeded the earlier works\(^\text{25}\). In a study, Nofemele et al. (2012)\(^\text{26}\) demonstrated 7.8% percent ethanol production from sugarcane molasses using Saccharomyces cerevisiae. In Bangladesh, five yeast isolates were reported\(^\text{27}\) to be used for the similar experiments where those isolates (TY, BY, GY-1, RY and SY) had alcohol production rate of 12.0%, 5.90%, 5.80%, 6.70% and 5.80%, respectively at 30°C after 48 hours of incubation. Significant elevation of ethanol production rate was observed in the co-fermentation process where cellulosic media were inoculated with cellulolytic bacteria previously. Overall, the method proves the efficiency of the co-fermentation\(^\text{25,26}\).

Conclusions
The present study allowed the isolation and characterization of two Saccharomyces cerevisiae isolates (SC1 and DJ1) with potential for ethanol production. Yeast isolated from sugarcane juice (named SC1 for this study) showed the highest percentage of alcohol production from cellulosic substrates. Vegetable peels pretreated with cellulolytic bacteria are detected as a suitable fermentation substrate. If the fermentation conditions are optimized, this procedure may be used for large-scale bioethanol production from cellulosic wastes. Scaling up of the experiment can be beneficial for power generation, bioethanol can be used as an alternative to fossil fuels. The raw materials required for the production of bioethanol are cheap and available. It will decrease environmental pollution, pave the pathway towards a proper waste management system and also fertilizers can be produced from the wasted substrate.

Data availability
Dataset 1: Alcohol production from vegetable peels by yeast isolates SC1 and DJ1 at pH 6. Media pre-treated with Bacillus subtilis. 10.5256/f1000research.13952.d19563\(^\text{26}\)

Dataset 2: Alcohol production from vegetable peels by yeast isolates SC1 and DJ1 at pH 6. Media without Bacillus subtilis pre-treatment. 10.5256/f1000research.13952.d19564\(^\text{27}\)

Competing interests
No competing interests were disclosed.

Grant information
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The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

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Author information
When the research was carried out, SSR was a MSc student at Biotechnology Program, Department of Mathematics and Natural Sciences, BRAC University, and NC was the Coordinator of Biotechnology and Microbiology programmes at Department of Mathematics and Natural Sciences, BRAC University.

References


Open Peer Review

Current Referee Status:  ❓  ❓

Version 1

Referee Report 20 April 2018

doi:10.5256/f1000research.15166.r32524

Md Fakruddin
Department of Molecular Physiology, Faculty of Life Sciences, Kumamoto University, Kumamoto, Japan

1. Reference should be added in method sections.

2. Ethanol production was represented in both % and gm/L. Please use only one unit uniformly.

3. Long time fermentation (96 hr) should be done to ensure the efficiency of the isolate.

4. Introduction should be improved to address the objective of the study.

5. Compare your results with related published papers.

6. Please indicate the advantage of kitchen waste over more economical source such as molasses.

7. It is not clear what changes occur due to bacillus pretreatment.

8. Without large scale fermentation, the conclusion may be misleading.

Is the work clearly and accurately presented and does it cite the current literature?
Partly

Is the study design appropriate and is the work technically sound?
Partly

Are sufficient details of methods and analysis provided to allow replication by others?
Yes

If applicable, is the statistical analysis and its interpretation appropriate?
Not applicable

Are all the source data underlying the results available to ensure full reproducibility?
Partly

Are the conclusions drawn adequately supported by the results?
Partly
**Competing Interests:** No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Referee Report 16 March 2018

doi:10.5256/f1000research.15166.r31498

Nazia Hossain
Faculty of Integrated Technologies (FIT), University of Brunei Darussalam, Gadong, Brunei

1. Abstract
   - Kitchen waste might be a possible raw material for bioethanol production for clean technology according to the research result but it cannot be concluded as excellent source. Authors need to upgrade this view for the abstract (conclusion section).

2. Introduction
   - Please provide a brief description regarding the necessity of biofuel and bioethanol application in the beginning.
   - You may describe the common reactions what may take place for ethanol production from biomass and the role of microbes.
   - I suggest splitting first paragraph into 2 paragraphs and describing specific issues for each instead of mixing up.

3. Methods
   - Ok.

4. Result and Discussion
   - Did you isolate yeast from local resources due to economic purpose? If so, please mention it.
   - Sometimes fermentation might be effective after 48 hours (e.g. 72hr or 96hr). Please explain why this study have not tried upto 96 hr or above. Do your have any recommendation to do so in future?
   - Usually supplement addition (e.g. MgSO₄) enhances fermentation process and increases overall bioethanol production. May you please elaborate why this study has not applied any nutritional component?
   - In discussion section, authors cited ‘Teramoto et al. (2005) demonstrated that members of *Saccharomyces* spp. can tolerate ethanol concentrations of up to 16.5%’ which is from 2005. Many studies have been performed with *Saccharomyces spp.* after 2005 what proved more than 50% bioethanol production. For instance, I suggest authors to check (Sugar and Bioethanol Production from Oil Palm Trunk (OPT) by Nazia Hossain & Rafidah Jalil).
- Please provide few examples of comparative studies and compare your results with others since vegetable and fruits peels are very common sources of bioethanol generation worldwide.

5. Conclusion

- The study result showed little amount of bioethanol production rate what is not favourable for economical view but environmental. So authors simply cannot assume that this procedure may be used for large-scale application without any specific modification or upgrade. In that case, author might recommend some factors to accelerate the production rate before scale-up.
- I answered ‘Partly’ to the question ‘Are the conclusions drawn adequately supported by the results?’ because limitations are not mentioned in the conclusions.

6. Overall

- Bioethanol is more appropriate than ethanol (In Title).
- Grammar needs to be re-checked.
- Many poorly structured sentences were being visible in the whole manuscript. Therefore, overall english should be improved.
- I answered ‘Partly’ to the question ‘Is the work clearly and accurately presented and does it cite the current literature?’ as authors cited lot of old references while many upgraded experiments have been performed later on. I suggest authors to use references after 2000 only (especially for discussion part).

**Is the work clearly and accurately presented and does it cite the current literature?**
Partly

**Is the study design appropriate and is the work technically sound?**
Yes

**Are sufficient details of methods and analysis provided to allow replication by others?**
Yes

**If applicable, is the statistical analysis and its interpretation appropriate?**
Not applicable

**Are all the source data underlying the results available to ensure full reproducibility?**
Yes

**Are the conclusions drawn adequately supported by the results?**
Partly

**Competing Interests:** No competing interests were disclosed.

**Referee Expertise:** Biofuel and bioenergy

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