RESEARCH NOTE

Blood-derived non-extracellular vesicle proteins as potential biomarkers for the diagnosis of early ER+ breast cancer and detection of lymph node involvement [version 3; peer review: 2 approved]

Previously named: Blood-derived extracellular proteins as potential biomarkers for the diagnosis of early ER+ breast cancer and detection of lymph node involvement

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Abstract
Extracellular vesicles (EV’s) are membrane surrounded structures released by different cell types and are emerging as potential therapeutic and diagnostic targets in cancer. In the present study, plasma samples derived from 7 patients with metastatic and non-metastatic ER+ (estrogen receptor positive) breast cancer (BC) were collected and their respective (EVs) isolated and the protein content analyzed by mass spectrometry and FunRich analysis. Two putative plasma biomarkers (absent in healthy controls samples) were identified which could be used to detect early ER+ breast cancer and for those with lymph node (LN) involvement. However, given the current limitations of the EV isolation method used, it is possible that these biomarkers did not originate from EVs and may represent blood-derived extracellular proteins. Further work in a larger patient cohort is warranted to confirm these findings and examine the diagnostic potential of these biomarkers.

Keywords
ER+ breast cancer, extracellular vesicles, plasma, biomarkers, diagnostic, lymph node involvement, metastases
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Author roles: Tucker R: Conceptualization, Data Curation, Formal Analysis, Funding Acquisition, Methodology, Supervision, Validation, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing; Pedro A: Conceptualization, Data Curation, Formal Analysis, Funding Acquisition, Investigation, Methodology, Validation, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing

Competing interests: No competing interests were disclosed.

Grant information: This work is supported by the Foundation of Science and Technology of Portugal [RECI/BiM-ONC/0201/2012], Lyden lab (Weill Cornell Medical College, USA), Champalimaud Foundation Portugal, and Romã Laboratories Ltd
The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

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Extracellular vesicles (EVs) are membrane surrounded structures released by different cell types that are involved in cellular communications and are emerging as potential therapeutic and diagnostic targets in cancer1 as in the case of early pancreatic cancer2.

EVs can be classified in several subtypes based on their size, shape, and supposed origin. Exosomes are defined as ∼30–100 nm vesicles which originate from multivesicular bodies (MVB) and contain late endosomal markers3,4, although biochemically indistinguishable vesicles can bud directly from the plasma membrane5,6. Microvesicles or shedding vesicles are generally larger (>200 nm), are more variable in shape and density, and likely originate from the plasma membrane7,8. EVs may contain proteins, lipids, and RNAs, however how these components are sorted into EVs remains unclear.

Tumor-derived EVs are also critical components for preparing the tumor microenvironment because they enable tumor cells to escape from the immunological surveillance4 and help in the setting of a pre-metastatic niche for the engraftment of detached cancer cells8. Both exosomes and MVs have been extensively studied and attributed various important physiological roles in cancer9,10,11. For instance, EVs have been found to play an important role in every phase of cancer development from cancer initiation, invasion and metastasis12. For these reasons, EVs are potential therapeutic and diagnostic targets in cancer and EV-derived biomarkers maybe useful for predicting future metastatic development and identify metastasis sites13.

ER+ (estrogen receptor positive) breast cancer (BC) represents 60–80% of all BC cases14,15. Here we describe our preliminary findings exploring the role of tumour derived EVs biomarkers that could ultimately be used as part of a test kit for the detection of early ER+ BC and lymph node involvement.

Methods
Samples
Plasma samples from 4 control patients (2 adult women and 2 men) which were confirmed as not having any form of BC, ER+ BC metastases, BC1 and BC2 explants EVs, SKBC and parental BC (Lyden lab, WCM, USA). Samples CF37, CF5, CF1, CF25, CF33, CF27 and CF110 and C7 (female control plasma sample) were collected at Champalimaud Clinical Centre, Portugal, as part of a study on the role of tumor-derived microvesicles and bone marrow progenitor cells as diagnostic and prognostic biomarkers in advanced BC and inflammatory BC Patients (RECI/BIM-ONC/0201/2012, FCT, Portugal). ER+ BC patient samples were selected based on their stage of disease progression – confirmed by CT-scan and surgery. EVs derived from conditioned media of cells lines SKBr3, MCF7, MDA468, MDA231 and MCF10A were also used in this study (details about these samples can be found in Table 1).

Ethics approval and informed consent
This study was approved by an Ethics Review Board at Champalimaud Foundation, Portugal. All study patients provided their written, informed consent.

EV purification and analysis
EV purification and analysis were performed at the Lyden lab (WCM) according to Andre et al., 201616. Briefly, plasma was pelleted at 500 × g for 10 min, then the supernatant was centrifuged at 20,000 × g for 20 min. Exosomes were then harvested by centrifugation at 100,000 × g for 70 min. The exosome pellet is resuspended in PBS and collected by ultracentrifugation at 100,000 × g for 70 min. The exosome pellet is resuspended in PBS and then stored at −80°C. The LM10 nanoparticle characterization system (NanoSight) equipped with a blue laser (405 nm) equipment.

Proteomics and proteomic analysis
Proteomic analysis was performed at the Rockefeller University, Proteomics Center as described in Hamidi et al., 201717. Proteomic analysis was performed with the help of FunRich Program version 3. Only proteins with Mascot scores of approximately 90 or >90 were considered18.

Results and discussion
Clinical data on the EVs isolated from BC patient’s plasma samples and cell lines can be found in Table 1. The method used for EV isolation also precipitates lipoproteins and immunocomplexes (IC) which are known possible contaminants19. However, samples submitted for mass spectrometry analysis showed none of the recognised contaminants of high speed centrifugation. In the two patients with early BC (Table 2a), we detected HCG1745306 isofrom CRA-a, a protein from the family of alpha type haemoglobins and for the patient with lymph node involvement, we detected histone H1.2 (Table 2a-b). HCG1745306 isofrom CRA-a was only present in the two patients with early BC with Mascot scores of 3208.8 and 3966.5.
### Table 1. Clinical data for different patient samples and cell lines.

<table>
<thead>
<tr>
<th>Sample ID</th>
<th>Menopausal status</th>
<th>ER/PR/Her2 status (%)</th>
<th>Metastases pattern</th>
<th>Sample type</th>
</tr>
</thead>
<tbody>
<tr>
<td>CF5</td>
<td>pre</td>
<td>100/95/-</td>
<td>LN+</td>
<td>Plasma</td>
</tr>
<tr>
<td>CF37</td>
<td>pos</td>
<td>100/-/-</td>
<td>LN-</td>
<td>Plasma</td>
</tr>
<tr>
<td>CF110</td>
<td>pos</td>
<td>100/100/-</td>
<td>Locally advanced</td>
<td>Plasma</td>
</tr>
<tr>
<td>CF1</td>
<td>pre</td>
<td>100/100/-</td>
<td>LN, liver</td>
<td>Plasma</td>
</tr>
<tr>
<td>CF25</td>
<td>pos</td>
<td>75/25/-</td>
<td>LN, liver, cartilage, skin</td>
<td>Plasma</td>
</tr>
<tr>
<td>CF33</td>
<td>pos</td>
<td>100/?/-</td>
<td>LN, liver, bone, skin, lung, brain</td>
<td>Plasma</td>
</tr>
<tr>
<td>CF27</td>
<td>pos</td>
<td>100/1/-</td>
<td>LN, lung, bone</td>
<td>Plasma</td>
</tr>
<tr>
<td>SKBC</td>
<td>?</td>
<td>?</td>
<td>Multiple metastasis</td>
<td>Plasma</td>
</tr>
<tr>
<td>BC1</td>
<td>?</td>
<td>ER+</td>
<td>Bone</td>
<td>Bone metastasis explant conditioned media</td>
</tr>
<tr>
<td>BC2</td>
<td>?</td>
<td>ER+</td>
<td>Bone</td>
<td>Bone metastasis explant conditioned media</td>
</tr>
<tr>
<td>Parental breast cancer</td>
<td>?</td>
<td>?</td>
<td>Primary tumor</td>
<td>Primary breast cancer conditioned media</td>
</tr>
<tr>
<td>SKBr3 (metastatic in mice)</td>
<td>? (43y)</td>
<td>HER2+</td>
<td>Metastasis</td>
<td>Pleural effusion (ATCC) Conditioned media from cell line culture</td>
</tr>
<tr>
<td>MDA468 (metastatic in mice)</td>
<td>? (51y)</td>
<td>TN (triple-negative)</td>
<td>Metastasis</td>
<td>Pleural effusion (ATCC) Conditioned media from cell line culture</td>
</tr>
<tr>
<td>MDA231 (highly metastatic in mice)</td>
<td>? (51y)</td>
<td>TN</td>
<td>Metastasis</td>
<td>Pleural effusion (ATCC) Conditioned media from cell line culture</td>
</tr>
<tr>
<td>MCF7 (poorly metastatic in mice)</td>
<td>pos</td>
<td>ER+</td>
<td>Metastasis</td>
<td>Pleural effusion (ATCC) Conditioned media from cell line culture</td>
</tr>
<tr>
<td>MCF10A</td>
<td>pre</td>
<td>Benign -fibrocystic disease</td>
<td>-----</td>
<td>Mammary gland; breast (ATCC) Conditioned media from cell line culture</td>
</tr>
</tbody>
</table>

### Table 2. a–b, Plasma EV-derived candidate biomarkers for early ER+ breast cancer and LN involvement. Also, represented the Mascot scores for each protein in each sample.

#### a

<table>
<thead>
<tr>
<th>Sample ID</th>
<th>Female1</th>
<th>Female2</th>
<th>Male1</th>
<th>Male2</th>
<th>CF37 (LN-)</th>
<th>CF5 (LN+)</th>
<th>CF110</th>
<th>CF1</th>
<th>CF25</th>
<th>CF27</th>
<th>CF33</th>
</tr>
</thead>
<tbody>
<tr>
<td>G3V1N2 (HCG1745306, isoform CRA_a) Early breast cancer</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>3208.75461</td>
<td>3966.542</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>P16403 (Histone H1.2) Early breast cancer, LN involvement</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>325.1718</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>

#### b

<table>
<thead>
<tr>
<th>Sample ID</th>
<th>SKBC</th>
<th>BC1</th>
<th>BC2</th>
<th>Parental BC</th>
<th>SKBr3</th>
<th>MDA468</th>
<th>MDA231</th>
<th>MCF7</th>
<th>MCF10A</th>
</tr>
</thead>
<tbody>
<tr>
<td>P16403 Histone H1.2 Early breast cancer, LN involvement</td>
<td>0</td>
<td>638.4</td>
<td>117.38</td>
<td>102.9</td>
<td>154.45</td>
<td>427.1</td>
<td>90.35</td>
<td>NS</td>
<td>0</td>
</tr>
</tbody>
</table>
respectively and absent in all controls and other patient samples.

Histone H1.2 was also detected in samples from the two patients with bone metastases, a parental primary BC sample and metastatic SKBr-3, MDA468, MDA231 cell lines. However, histone H1.2 was absent from the plasma sample of a patient with multiple metastases, from the non-metastatic MCF7 cell line (a non significant mascot score) and from MCF10A cells EVs (Table 2b). These observation suggests that histone H1.2 might represent a potential marker for LN involvement and metastatic potential. Recent studies suggest histone H1.2 phosphorylation may be useful as a clinical biomarker of breast and other cancers because of its ability to recognize proliferative cell populations. Both MCF7 (expressing an allelic variant A142T) and MDA231, have a greater number of histone H1.2 phosphorylations when compared to MCF10A cell line22. Curiously, phosphorylation of histone H1.2 at S173 increases during the M phase relative to the S phase, suggesting that this event is cell cycle-dependent and may serve as a marker for proliferation of cancer cells during BC invasion22,24. Also, histone H1.2 is a novel component of the nucleolar organizer regions during mitosis25 and H1.2 depletion was observed in a human BC cell line caused cell cycle G1-phase arrest26. Indeed, a higher mitotic index (≥ 7) in primary tumors is significantly associated with LN involvement27 and higher mitotic indices accurately predict axillary LN involvement at operation28.

Although we have identified two potential biomarkers possibly derived from EVs, our study does suffer from a number of recognised limitations. Firstly, ultracentrifugation is insufficient to purify EVs from other contaminants29. For example, co-isolation of high-density lipoprotein and other particles with EVs isolated from blood by density gradient centrifugation has been reported29,30 suggesting that the biomarkers we identified might not be associated with EVs but with a constituent of another particle type such as a lipoprotein. Secondly, as mentioned above, exosomes are defined as ~30–100 nm vesicles that originate from MVB. In contrast, microvesicles or shedding vesicles are generally larger (>200 nm), more variable in shape and density and arise from the plasma membrane. The size of the particles we isolated ranged from 76.7-213.4 and 73.8-192.3 nm, for samples CF5 and CF37, respectively and for all the samples between 12.3-298.4nm (Figure 1 and original NTA files) possibly correspond to low-density lipoproteins which have the same size as EVs31. Moreover, it is unlikely that EVs would contain a histone (which are normally confined to DNA in the nucleus). However, Thakur et al., claim to have identified genomic DNA in EVs by electron microscopy (EM) though the EM image is not of sufficient magnification to allow for an accurate morphologic analysis and may simply represent cellular debris or apoptotic bodies or even unspecific staining32,33. Additionally, it is also unlikely HCG1745306 isoform CRA-a, would be present in EVs and it may simply be a precipitant similar to the α-globin seen in β-thalassemia34. Therefore our current data does not support

![Figure 1. Nanosight (NTA) analysis for samples C7, CF37, CF5, CF110, CF1, CF25, CF27 and CF33.](image-url)
the idea that these biomarkers derived from EVs and could in fact be blood-derived extracellular proteins.

Nevertheless, a strength of our study is that samples were drawn from those with confirmed non-metastatic and metastatic disease at different sites and so are likely to be representative patients.

Conclusion
In conclusion, our observations suggest the possibility that HCG1745306 isoform CRA-a, and histone H1.2, irrespective of their origin, could represent potential biomarkers for the detection of early ER+ BC. Further work in a larger cohort of patients is clearly needed to confirm these initial findings.

References


Data availability
Dataset 1: The mass spectrometry analysis results from all patient samples 10.5256/f1000research.14129.d203204

Competing interests
No competing interests were disclosed.

Grant information
This work is supported by the Foundation of Science and Technology of Portugal [RECI/BIM-ONC/0201/2012], Lyden lab (Weill Cornell Medical College, USA), Champalimaud Foundation Portugal, and Romã Laboratories Ltd.

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.


33. https://pubpeer.com/publications/713241A9DBF653E9E2E1D10B1EC847

34. http://www.pathophys.org/thalassemia/

Open Peer Review

Current Peer Review Status:  ✔  ✔

Version 3

Reviewer Report 18 June 2018
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Matthew J. Shurtleff
University of California, San Francisco (UCSF), San Francisco, CA, USA

The results and conclusions are improved from previous revisions. However, as written, the paper is non-sensical. The title now states that the biomarkers are non-EV proteins, and the results and conclusions are appropriately skeptical about the EV origin of these proteins. However, the introduction only discusses the promise of EVs as biomarkers (and seems substantially unchanged from previous versions). I think any reader would find the introduction to be quite disconnected from the revised title, results and conclusions. For clarity, and to appropriately contextualize the study, the introduction should be revised to focus on the utility of circulating biomarkers, irregardless of their association with EVs.

Competing Interests: No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Version 2

Reviewer Report 01 May 2018
https://doi.org/10.5256/f1000research.16084.r33412

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Matthew J. Shurtleff
University of California, San Francisco (UCSF), San Francisco, CA, USA
The authors have not made a true attempt to satisfy my reservations. They have only parroted my reservations in their response and discussion, and removed the word "vesicle" from their title. They have not attempted to revise the content of their article at all. Indeed, the first two words of the text remain "extracellular vesicles". I do not consider this work satisfactory for indexing.

**Competing Interests:** No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to state that I do not consider it to be of an acceptable scientific standard, for reasons outlined above.

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**Version 1**

Reviewer Report 16 April 2018

https://doi.org/10.5256/f1000research.15368.r32581

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Matthew J. Shurtleff

University of California, San Francisco (UCSF), San Francisco, CA, USA

Tucker and Pedro present results from proteomics studies of material precipitated from the plasma of breast cancer patients and healthy controls, and from the media of explant cultures and cell lines. They observe CRA-a (HBA2, alpha hemoglobin) as being present in precipitated material from the plasma of patients with early breast cancer and Histone H1.2 as being present in samples with lymph node involvement (plasma), and the media of explant cultures of bone metastases and metastatic cell lines.

While the results are clearly presented, the interpretation that these are EV-based biomarkers is not supported. The EV isolation method used is insufficient to purify EVs from other contaminants. Furthermore, it is not intuitive that EVs would be likely to contain a histone (normally confined to the nucleus) or hemoglobin. The authors' discussion of this major concern is insufficient ("However, samples submitted for mass spectrometry analysis showed none of the recognised contaminants and it was therefore concluded that the main EV type present in these samples is the MV.").

The EV literature is quite confusing due, in part, to the over-interpretation of observations from samples prepared using inadequate isolation methods. Therefore, in it's current version, I do not recommend indexing of this report. Ideally, the authors should repeat the work with approaches that better enrich for EVs over non-EV protein contaminants (e.g. density-gradient based ultracentrifugation or immuno-affinity purification) and protease protection assays in the presence/absence of detergent should be used evaluate if the proteins are indeed associated with EVs and if they are on the surface or interior of vesicles. If further experiments are not possible, I recommend revising the paper to remove the unsupported conclusion that the proteins identified are EV-associated. (For example the title could be: Blood-derived extracellular proteins as potential biomarkers for the diagnosis of early ER+ breast cancer and detection of lymph node involvement).
For reference, I direct the authors to an updated publication prepared by members of the EV community on the minimal information for studies of EVs in the Journal of Extracellular Vesicles.  

References

Is the work clearly and accurately presented and does it cite the current literature?
Partly

Is the study design appropriate and is the work technically sound?
No

Are sufficient details of methods and analysis provided to allow replication by others?
Yes

If applicable, is the statistical analysis and its interpretation appropriate?
Not applicable

Are all the source data underlying the results available to ensure full reproducibility?
Yes

Are the conclusions drawn adequately supported by the results?
No

Competing Interests: No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Reviewer Report 26 March 2018
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Shweta Aras
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In this research note by Tucker et al., authors have identified 2 novel biomarkers potentially involved in early breast cancer development as well as as lymph node metastasis using extracellular vesicles from patient samples along with adequate controls of metastatic and non-metastatic cell lines. Several reports
in the literature have already suggested the importance of EVs in cancer initiation and progression in various types of cancers. Although the novelty of this publication is limited, identification of 2 new molecules differentially expressed in primary versus metastatic breast cancer patients opens up the possibility of them being used in a simple prognostic blood test for detecting early BC development as opposed to mammography and other invasive techniques. A probable limitation of the study would be lack of identification of molecular mechanisms through which this upregulation of CRA-a and histone H1.2 phosphorylation plays a role in metastasis, specifically in which step of metastatic cascade e.g. intravasation, EMT, extravasation etc? But overall, the authors have done a great job and the article is a good addition to the field.

Is the work clearly and accurately presented and does it cite the current literature?
Yes

Is the study design appropriate and is the work technically sound?
Partly

Are sufficient details of methods and analysis provided to allow replication by others?
Yes

If applicable, is the statistical analysis and its interpretation appropriate?
I cannot comment. A qualified statistician is required.

Are all the source data underlying the results available to ensure full reproducibility?
Yes

Are the conclusions drawn adequately supported by the results?
Yes

**Competing Interests:** No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.