shiny-pred: a server for the prediction of protein disordered regions [version 1; peer review: 1 approved with reservations, 1 not approved]

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Abstract
Intrinsically disordered proteins or intrinsically disordered regions (IDR) are segments within a protein chain lacking a stable three-dimensional structure under normal physiological conditions. Accurate prediction of IDRs is challenging due to their genome wide occurrence and low ratio of disordered residues, making them a difficult target for traditional classification techniques. Existing computational methods mostly rely on sequence profiles to improve accuracy, which is time consuming and computationally expensive. The shiny-pred application is an \textit{ab initio} sequence-only disorder predictor implemented in R/Shiny language. In order to make predictions, it uses convolutional neural network models, trained using PDB sequence data. It can be installed on any operating system on which R can be installed and run locally. A public version of the web application can be accessed at \url{https://gmu-binf.shinyapps.io/shiny-pred}

Keywords
Disordered proteins, machine learning, convolutional neural networks, R, Shiny

This article is included in the RPackage gateway.

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2. Appadurai Rajeswari, Indian Institute of Science, Bangalore, India
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Any reports and responses or comments on the article can be found at the end of the article.
Introduction
Experimental structure resolution of intrinsically disordered proteins/intrinsically disordered regions (IDP/IDRs) is complex, lengthy and expensive, leading to a variety of computational approaches being developed (He et al., 2009). Over 60 computational protein disorder prediction servers are currently available, although not all publicly. Methods can be classified in one of the following categories (Atkins et al., 2015): (i) Ab initio or sequence-based, (ii) clustering, (iii) template based, and (iv) meta or consensus.

shiny-pred is an ab initio predictor, which means it relies exclusively on amino acid sequence information to make disordered predictions. It uses prediction models based on convolutional neural networks and reduced protein alphabets. Currently there are three available models, each one built using the same training protein data from PDB (Berman et al., 2000) but differing on the convolutional neural network architecture. Since it doesn’t rely in sequence profiles to make predictions, it is fast to be used in proteome-wide disorder scenarios. It performs at the same level or outperforms other state of the art sequence-only methods, achieving accuracy levels of 0.76 and AUC of 0.85 on the publicly available CASP10 dataset (Monastyrskyy et al., 2014), at faster speeds.

Methods
Implementation
shiny-pred is written in the R programming language (R Core Team, 2017) and the shiny web application framework is implemented using the Shiny R package v1.1.0 (Chang, 2018).

Currently, three convolutional neural network models are made available by our application:

(i) cnn-64-ker-local, is a one layer convolutional network (step size 1 and window size of 32) with 64 kernels and local max pooling model; (ii) cnn-128-ker-local, implements one convolutional layer (step size 1 and window size of 32) with 128 kernels and local max pooling model; and (iii) cnn-2-conv-local implements two convolutional layers (64 and 32 kernels) with local max pooling.

The models were created, trained and accessed using the keras R package v2.1.6 (Allaire & Chollet, 2018).

Operation
Our tool has two operation modes; predicting disordered residues in protein sequences (prediction) and benchmarking the predictor performance against sequences with known disorder information (benchmark). The mode is selected automatically based on the format of the input sequences. Users can either upload a sequence file, type/paste a sequence into the text area or select pre-loaded examples from a list.

When in prediction mode, the amino acid sequences are expected to be in FASTA format (Figure 1). In benchmark mode, input sequences in FASTA format are expected to have an additional line containing the disorder information (D=disorder, O=ordered). Multiple sequences can be submitted at once; several examples for different types of submissions (prediction and benchmark modes) are made available as examples. In both modes, the application will show a result panel, where for each input sequence a graph with the probability of disorder per residue is plotted (Figure 2).

(1) Prediction mode
The workflow for protein disorder prediction is:

(i) Input the target sequences (in FASTA format) in the text area;
(ii) Select the model to use for the prediction (default is cnn-128-ker-local) and submit the sequence for prediction;
(iii) Visualize and download results.

(2) Benchmark mode
In benchmark mode, input sequences are expected to have an extra line with the actual disorder information to be used as benchmark. Result tables will populate two extra columns (actual class and match) with the actual disorder information and if the prediction was accurate for the current residue. An extra panel (Benchmark) shows the ROC curve along with other common binary metrics (sensitivity, specificity, balance accuracy and Matthews correlation coefficient).

Use cases
We use shiny-pred to predict disordered regions within the publicly available CASP10 benchmark dataset. The dataset contains 94 target sequences, each one annotated with the disorder/order information at the residue level. The annotated dataset is provided as an example (‘CASP_all’) and it can be selected form the example selection list on the ‘Sequence Input’ tab. Figure 3 shows the input panel after the dataset is selected and loaded. Predictions per sequence can be viewed and downloaded from the ‘Results’ tab while the ‘Benchmark’ tab provides a summary of the performance using binary and statistical metrics. Figure 4 shows the server performance for the input dataset, achieving an AUC value of 0.85 and balance accuracy of 0.75.

Summary
This article presents shiny-pred, a sequence-only ab initio web application for predicting protein disorder. It’s based on reduced amino acid alphabets and convolutional neural networks, being fast and accurate, it is suitable for large proteome-wide experiments.

Software availability
Software available from: https://gmu-binf.shinyapps.io/shiny-pred

Source code available from: https://github.com/mauricioob/shiny-pred

 Archived source code as at time of publication: https://doi.org/10.5281/zenodo.256725 (Mauricio, 2019).

License: GNU public license (GPL-3)
Figure 1. Input sequence format (prediction mode).

Figure 2. Prediction results.
Figure 3. Input sequence format (benchmark mode).

Figure 4. Predictor benchmarking.
Grant information
The author(s) declared that no grants were involved in supporting this work.

Acknowledgments
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References


Reference Source

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The authors presented yet another neural network-based disorder prediction tool written in R, trained on PDB data and benchmarked on CASP10 dataset and they claim that the tool outperforms other existing tools in terms of both calculation speed and performance.

We tried using the tool for predicting the known disordered sequences and found that the predictions are accurate and similar to other tools such as IUPRED, DISOPRED3 servers for the well-known disordered sequences such as p53 and Histatin5.

In terms of concerns, I have following comments to make:

1. In general, I find the paper does not describe the motivation, methods and the results in a self-sufficient manner and these could be elaborated further.

2. As the authors state in the paper, there are over 60 tools already existing for disorder prediction. The justification for requiring another tool is not clearly stated.

3. The authors mention they have used PDB data for training the neural network. Do they take all the currently available PDB datasets for training? Does any overlap exist between the datasets trained and benchmarked? The reason why I am asking this is the CASP10 dataset that the authors used for benchmarking has been released in 2012, which would be a subset of the training PDB dataset if they have taken all the PDB data published till date.

4. The authors claim that their method is faster than the existing methods. It would be nice to provide evidence towards that and provide some benchmarking data.

5. AUC and balance accuracy are the two metrics used for evaluating the performance of the
tool. However, a clear definition of these terms are not described in the method section.

6. The tool should be tested and benchmarked against a larger data set such as Disorder-723, which contains 723 disorder sequences.

Is the rationale for developing the new software tool clearly explained?  
No

Is the description of the software tool technically sound?  
Partly

Are sufficient details of the code, methods and analysis (if applicable) provided to allow replication of the software development and its use by others?  
No

Is sufficient information provided to allow interpretation of the expected output datasets and any results generated using the tool?  
No

Are the conclusions about the tool and its performance adequately supported by the findings presented in the article?  
No

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** Membrane Biophysics, Protein Structures and Folding, Mechanotransduction, Statistical mechanics of Biological Systems, Integrative Modeling, Multiscale Biomolecular Simulations

We confirm that we have read this submission and believe that we have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however we have significant reservations, as outlined above.
There are already many disorder predictors, some of which are based upon deep convolutional neural network and can do prediction directly on amino acid sequence instead of sequence profile. This manuscript does not have sufficient results to justify why one more web server for disorder prediction is needed. Here are some concerns:

1. Please compare with existing, similar methods.
2. It is better to test the method on more recent CASP datasets and make sure that there is no redundancy between training and test data. Ideally, a much larger test set shall be used to evaluate the method.
3. AUC may not be a good metric for disorder prediction since the ratio of disordered residues is quite small.
   Precision and Recall may be better.
4. Existing work shall be cited.

Is the rationale for developing the new software tool clearly explained?
Partly

Is the description of the software tool technically sound?
Yes

Are sufficient details of the code, methods and analysis (if applicable) provided to allow replication of the software development and its use by others?
Yes

Is sufficient information provided to allow interpretation of the expected output datasets and any results generated using the tool?
Yes

Are the conclusions about the tool and its performance adequately supported by the findings presented in the article?
Partly

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Computational biology, machine learning, optimization.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to state that I do not consider it to be of an acceptable scientific standard, for reasons outlined above.
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