Molecular detection of Epstein-Barr virus in breast cancer among Sudanese female population: a case-control study

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Abstract

Background: Breast cancer is the most common cancer in women worldwide and in Sudan. Breast cancer occurs due to a multifactorial process and infection with an oncogenic virus has been recently investigated as a possible risk factor for breast cancer. For nearly two decades, studies have incriminated Epstein-Barr virus (EBV) in the etiology of breast cancer. However, the results are unconvincing, and their interpretation has remained a matter of debate. The aim of this study was to detect EBV in breast cancer biopsies obtained from Sudanese female patients.

Methods: A descriptive, hospital-based, case-control study, conducted at Faculty of Medical Laboratory Science, University of Khartoum, Khartoum, Sudan. Archival blocks were obtained from 115 patients with breast cancer and 115 controls during the period between November 2016 till March 2017.

Results: Among 115 breast cancer tissue specimens, EBV DNA was identified in 42/115 (36.5 %) samples and was not identified in 73/115 (63.5 %) tissue samples. The highest frequency of EBV detection was among 41–60 year-olds (23/42, 54.7 %), followed by 21–40 year-olds (12/42, 28.6 %) and 61–80 year-olds (5/42, 11.9 %). In the control group, the majority were diagnosed with fibroadenoma (70.4%), followed by fibrocystic changes (10.4%) and lactating changes (0.9%).

Conclusion: The data obtained in this study demonstrated that EBV was present in a high percentage of our study population; however, the exact role of EBV in Sudanese breast cancer needs to be studied more in depth.

Keywords
Breast cancer, Invasive Ductal carcinoma, Epstein-Barr virus
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Introduction

Epstein-Barr virus (EBV) was discovered in the early 1960s by direct electron microscopy and it infects up to 95% of the adult human population, primarily in their early life. It usually remains silent and doesn’t cause any symptoms as the immune system doesn’t treat the virus like an invader following the initial infection. However, infection can lead to severe manifestations in later life\textsuperscript{1-3}. This virus has been found to be the cause of many types of cancers with Burkitt’s lymphoma, non-Hodgkin’s lymphoma, nasopharyngeal carcinoma, as well as Leiomyosarcoma arising in immunocompromised individuals\textsuperscript{4}. The main mechanism by which the virus can induce cell transformation from normal to malignant cells is not yet understood and is still under investigation, although some researchers have related this to EBV viral protein effects\textsuperscript{5-6}.

Breast cancer is one of the most frequent types of cancers and is the leading cause of death in females globally\textsuperscript{6}. Several factors can promote the development of breast cancer, including infections, especially oncogenic viruses that have been proven to be possible risk factors for the development of the cancer. These viruses include mouse mammary tumour virus (MMTV, causes breast cancer in mice), EBV and human papilloma virus (HPV)\textsuperscript{7-10}. EBV DNA was identified previously in several studies using different molecular based techniques, including sequencing, polymerase chain reaction (PCR), in situ hybridization, immunohistochemistry and real-time PCR\textsuperscript{11}.

In Sudan, breast cancer is a leading type of cancer in adults, along with leukemia, prostatic carcinoma, lymphoma and colorectal carcinoma\textsuperscript{12}. Therefore, we aim in this study to detect EBV in breast cancer biopsies obtained from Sudanese female patients.

Methods

Study design

This is a descriptive, hospital-based, case-control study, conducted at Faculty of Medical Laboratory Science, University of Khartoum, Khartoum City, Sudan, during the period of November 2016 until March 2017.

Sample collection

Samples were collected from Soba Teaching Hospital and Military Hospital in Khartoum state, Sudan, during the period of November 2016 to January 2017. Archival blocks of 115 female patients diagnosed with breast cancer and 115 normal breast lesions (controls) were retrieved for the detection of EBV. All archival blocks from female patients diagnosed with breast cancer were included in the study, with no further inclusion or exclusion criteria. For the controls, archival blocks from women with a breast lesion, not diagnosed as breast cancer, taken during the same period as the breast cancer patient samples were included. The control samples were not matched to the breast cancer patient samples. Analysis of these blocks were carried out during the period of February 2017 to March 2017.

Preparation of formalin-fixed paraffin-embedded tissue

Formalin-fixed paraffin-embedded blocks (FFPB) were cut at a thickness of 10 μm using a rotary microtome and the ribbons of the cut tissues transferred into 1.5 ml eppendorf tube using sterile forceps. The samples were deparaffinized by adding 1 ml of xylene to each sample and incubating for 30 minutes to allow the removal of the wax from the tissue sections. This step was repeated twice. The tissue was rehydrated using a series of ethanol washes, using 95%, 80% and then 70% ethanol for 15 min for each.

DNA extraction and assessment of DNA quality

The DNA was extracted using the phenol-chloroform method with the aid of lysis buffer that composed from Sodium dodecyl sulfate (SDS) and proteinase K (40 μl) to get rid of proteins, as previously described\textsuperscript{13-15}. To purify the DNA from the lystate, buffer saturated phenol was used. Then, in order to precipitate the DNA, 3M sodium acetate and isopropanol were added and this was spun in a centrifuge at 16000 rpm for seven mins to pellet the nucleic acid. This step was followed by washing in 70% ethanol to get rid from the salts and centrifugation at high speed to reconstruct the pellet again.

DNA was resuspended in distilled water, quantified and stored at −20 °C. The final extracted DNA was checked for purity and quantity using a Nanodrop 1000 spectrophotometer. If the 260/280 nm absorption ratio was between 1.8-2, the DNA was considered to be pure. Purified DNA was used for PCRs.

Molecular identification of EBV

To determine if the EBV DNA was present in our samples, DNA was amplified for detection of the EBV LMP-1 gene using forward 5- CCG AAG AGG TTG AAA ACA AA-3 and reverse 5- GTG GGG GTC GTC ATC ATC TC-3 primers. PCR was performed in a final reaction volume of 25μl, with 1 μl of template DNA, using iNtRON’s Maxime PCR PreMix Kit (i-Taq, Korea; Catalog No. 25025) according to manufacturer instructions. The following PCR conditions were used: 4 min at 95°C; then 35 cycles consisting of a denaturing step at 94°C for 1 min, an annealing step at 60°C for 1 min and a polymerization step at 72°C for 1 min; followed by a single incubation step for 10 min at 72°C. PCR was performed on an Aeris Thermocycler (AERIS-BG096, Esco Micro Pte.Ltd, Singapore).

Analysis of PCR results

The amplification product of 131 bp was separated on a 2% agarose gel stained with ethidium bromide and visualized under UV light using the UVCI-1100 gel imaging system (Major Science, UVCI-1100, Saratoga, CA, USA). A 100 bp DNA ladder was used and a band of 131 bp was considered as a positive (Figure 1).

Statistical analysis

The statistical analysis was performed using SPSS (version 16.0) statistical software. Descriptive statistics were conducted to generate graphical and numerical summaries. The Chi-squared test was used to compare the frequencies of the categorical variables. A value of p < 0.05 was considered statistically significant.
Ethical statement
The study was approved by the faculty research board of the faculty of Medical Laboratory Sciences, University of Khartoum, Sudan. Written informed consent was obtained from patients for the use of their samples and publication of results.

Results
The age of the breast cancer patients ranged between 23 and 80 years and the mean age was 50.49 ± 9.50 years, while the control group age ranged from 25 to 72 years and the mean age was 37.65 ± 5.89 years. Regarding histological type, 96 (83.5%) of the patients diagnosed with breast cancer had invasive ductal carcinoma, 18 (15.7%) had invasive lobular carcinoma and one (0.9%) had mucinous carcinoma. Regarding the clinical staging (according to the TNM staging of the American Joint Committee on Cancer), 10 (8.7%) were stage I, 13 (11.3%) were stage II A, 15 (13%) were stage IIB, 17 (14.8) were stage IIIA, 24 (20.9%) were stage IIIB and 36 (31.1) were stage IV (Table 1). In the control group, the majority were diagnosed with fibroadenoma (70.4%), followed by fibrocystic changes (10.4%) and lactating changes (0.9%).

Among the 115 breast cancers tissue specimens, EBV DNA was identified in 42/115 (36.5 %) samples and was not identified in 73/115 (63.5 %) tissue samples (Table 2). In the control group, EBV was detected in 101/115 (87.8%) and only 14/115 (12.2%) tested negative for the presence of the virus. The Chi-square test showed that EBV has a significant effect on breast cancer with a p-value of 0.001.

The highest frequency of infection was seen in the age group 41–60, with infection found in 23/42 (54.7 %), followed by age ranges 21–40 (12/42, 28.5 %) and 61–80 (5/42, 11.9 %).

Notably, with regard to EBV infection and staging of breast cancer, by far the majority of infections were detected among stage IV (11/42, 26.1%), followed by stage IIIA (10/42, 23.8%), and stage IIIB (8/42, 19.0%), although these results were shown to be statistically insignificant (Table 3).

Discussion
Breast cancer has been proven to be a leading cause of death amongst women in the western world after lung cancer. Its incidence has been confirmed to be higher in females than males, and the number of cases increase with age. Early case detection has improved due to the promotion of screening procedures and diagnosis. In Sudan, breast cancer still accounts for many cases of cancer among women. In 1959, breast cancer was found to account 22.9% of all cancer cases. The bulk of those patients were females within the menopausal age, with disease clinically progressing in a similar pattern as in European ladies. In 2010, an updated study of breast cancer prevalence in Sudan by Intisar and colleagues found a prevalence of 25.1 per 100,000. For approximately twenty years, studies have been conducted that have aimed to identify the association between EBV and breast cancer. The first report of an association between EBV and breast cancer was from a study that carried out screening of blood
and DNA of 91 patients with breast cancer by PCR in order to detect the EBV genome in 1995. Since that, other studies have confirmed the finding that EBV is associated with breast cancer. However, these results have been controversial since negative results were obtained in some studies and explanations of the results have been under discussion for several years.

Moreover, in 2016 Hu and colleagues queried whether the EBV can be associated with breast cancer development. NOD/SCID mice were used and their results suggested that EBV can infect primary human mammary epithelial cells (MECs) through CD21, leading to phenotypic variations consistent with transformation. These immortalized MECs infected with EBV cooperatively (with activated Ras) increase tumor formation in vivo, recapitulating a multi-step tumorigenesis in an established animal model.

A study was conducted in 2014 by Yahia and colleagues to study the relationship between breast cancer and the presence of EBV genome. Interestingly, this was the first study in Sudan to assess the role of EBV infection in the etiology of breast cancer. They used PCR and in situ hybridization to screen 92 breast cancer samples, aiming to measure the presence of Epstein-Barr nuclear antigen 4 (EBNA-4) and LMP-1. Their final outcome was statistically significant (p = 0.0001), demonstrating the presence of EBV genome in 49 (53.3%) and 10 (11%) patients by LMP-1 and EBNA-4 PCR, respectively. This result is in agreement with our results, which showed that EBV DNA was identified in 42/115 (36.5 %) samples and was not identified in 73/115 (63.5 %) tissue samples.

Notably, the incidence of EBV among Iranian females with breast cancer has been found to be low compared to our findings. In 2016, Mohammad Izadeh and colleagues tested LMP-1 antigen expression in breast carcinoma using immunohistochemistry in 80 Iranian patients. LMP-1 expression was detected in 6 cases (7.5%) of breast carcinoma cases and these results were found to be statistically significant. LMP-1 expression was not detected in the normal breast tissue, taken from a site adjacent to the carcinoma, in all cases. Furthermore, another study from Iran, conducted by Kadivar and colleagues, to evaluate the presence of EBV in benign and malignant breast lesions using PCR and immunohistochemistry is also in disagreement with our results, as all breast cancer samples were negative for EBV, as were the control group samples.

The present study verified the presence of EBV in 36.5% of the total breast cancer samples, in agreement with the study conducted by Bonet and colleagues, in which 51 (51%) cases were found to be positive for EBV by PCR out of a total of 100 breast cancer cases.

However, this study has some limitations. The study did not use cohort techniques to assess whether EBV viral load had an effect on the prognosis of breast cancer and we could not use advanced techniques for visualization of the virus in the malignant cells due to time and budget constraints.

### Table 1. The distribution of study population by age, histological type and staging.

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Percentages</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age range</td>
<td></td>
</tr>
<tr>
<td>21–40</td>
<td>22 (10.1%)</td>
</tr>
<tr>
<td>41–60</td>
<td>77 (67.0%)</td>
</tr>
<tr>
<td>61–80</td>
<td>16 (13.9%)</td>
</tr>
<tr>
<td>Histological subtypes</td>
<td></td>
</tr>
<tr>
<td>Invasive ductal carcinoma</td>
<td>96 (83.5%)</td>
</tr>
<tr>
<td>Invasive lobular carcinoma</td>
<td>18 (15.7%)</td>
</tr>
<tr>
<td>Mucinous carcinoma</td>
<td>1 (0.9%)</td>
</tr>
<tr>
<td>Stage</td>
<td></td>
</tr>
<tr>
<td>Stage I</td>
<td>10 (8.7%)</td>
</tr>
<tr>
<td>Stage II A</td>
<td>13 (11.3%)</td>
</tr>
<tr>
<td>Stage II B</td>
<td>15 (13%)</td>
</tr>
<tr>
<td>Stage III A</td>
<td>17 (14.8)</td>
</tr>
<tr>
<td>Stage III B</td>
<td>24 (20.9%)</td>
</tr>
<tr>
<td>Stage IV</td>
<td>36 (31.3%)</td>
</tr>
</tbody>
</table>

### Table 2. The distribution of Epstein-Barr virus (EBV) among patient and control groups.

<table>
<thead>
<tr>
<th>Patients</th>
<th>Controls</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Negative</td>
<td>Positive</td>
</tr>
<tr>
<td>Negative</td>
<td>59</td>
<td>14</td>
</tr>
<tr>
<td>Positive</td>
<td>42</td>
<td>0</td>
</tr>
<tr>
<td>Total</td>
<td>101</td>
<td>14</td>
</tr>
</tbody>
</table>

(Chi square test; p value =0.001).

### Table 3. The distribution of the study population by staging and Epstein-Barr virus (EBV) infection.

<table>
<thead>
<tr>
<th>Staging</th>
<th>EBV</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Positive</td>
<td>Negative</td>
</tr>
<tr>
<td>Stage I</td>
<td>2</td>
<td>8</td>
</tr>
<tr>
<td>Stage II A</td>
<td>7</td>
<td>6</td>
</tr>
<tr>
<td>Stage II B</td>
<td>4</td>
<td>11</td>
</tr>
<tr>
<td>Stage III A</td>
<td>10</td>
<td>7</td>
</tr>
<tr>
<td>Stage III B</td>
<td>8</td>
<td>16</td>
</tr>
<tr>
<td>Stage IV</td>
<td>11</td>
<td>25</td>
</tr>
<tr>
<td>Total</td>
<td>42</td>
<td>73</td>
</tr>
</tbody>
</table>

(Chi square test; p value > 0.05).
Conclusion
In conclusion, the data obtained in this study demonstrated that EBV was present in a high percentage of our study population; however, the exact role of EBV in Sudanese breast cancer needs to be studied more in depth.

Data availability
Underlying data
Harvard Dataverse: Replication Data for: EBV and Breast cancer Data among Sudanese patients. https://doi.org/10.7910/DVN/94YOCV

EBV_and_Breast_cancer_Data.tab (diagnoses, PCR results and ages of patients and controls)

References


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