In vitro efficacy of a copper iodine complex PPE disinfectant for SARS-CoV-2 inactivation [version 2; peer review: 2 approved]

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Abstract
Background: The ability to protect workers and healthcare professionals from infection by SARS-CoV-2, the virus that causes coronavirus disease 2019 (COVID-19), is of great concern. Hospitals, nursing homes and employers are adopting infection control strategies based on guidance from leading public health organizations such as the CDC, OSHA, FDA, and other government bodies. Certain hard surface disinfectants are effective against SARS-CoV-2 but are not suitable for use on skin or personal protective equipment (PPE) that comes into contact with skin. Furthermore, near-universal alcohol-based hand sanitizers are acceptable for use on skin, but they are not suitable for use on PPE. PPE, especially masks, are also commonly being used for longer durations than normal. There is a need for new products and techniques that can effectively disinfect PPE during wear time without having detrimental effects on surrounding skin. Clyraguard spray is a novel copper iodine complex designed to be used on non-critical PPE.

Methods: In this study, the Clyraguard copper iodine complex was tested for its ability to inactivate SARS-CoV-2 in solution.

Results: These data indicate the product to be effective in reducing SARS-CoV-2 titers in a time-dependent manner, with the virus being reduced below the detection limits within 30 minutes.

Conclusions: These results suggest that Clyraguard may be an effective tool for mitigating cross-contamination of non-critical PPE that may come into contact with SARS-CoV-2.

Keywords
COVID, PPE, iodine, disinfection, decontamination, virus
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Author roles: Mantlo E: Data Curation, Investigation, Methodology, Visualization, Writing – Original Draft Preparation; Rhodes T: Conceptualization, Formal Analysis, Methodology, Writing – Review & Editing; Boutros J: Formal Analysis, Writing – Original Draft Preparation, Writing – Review & Editing; Patterson-Fortin L: Writing – Original Draft Preparation, Writing – Review & Editing; Evans A: Methodology, Project Administration, Writing – Original Draft Preparation, Writing – Review & Editing; Paessler S: Conceptualization, Formal Analysis, Investigation, Methodology, Project Administration, Resources, Supervision, Validation, Writing – Review & Editing

Competing interests: Studies conducted in the laboratory of Dr. Slobodan Paessler at Galveston National Laboratory at the University of Texas Medical Branch described herein were funded by Clyra Medical Technologies, Inc., who developed and manufactures Clyraguard, and were requested to be conducted by Clyra Medical Technologies, Inc. Clyra staff who are listed as authors on this paper do not themselves have any financial commitments to the research described herein, but are employed as staff or consultants by Clyra.

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Introduction

In late 2019, the world saw the spread of a novel coronavirus (SARS-CoV-2) that caused a global pandemic (ongoing as of this writing) that has claimed the lives of more than 260,000 people and infected 3.8 million as of May 2020 (Johns Hopkins Coronavirus Resource Center, 2020). The spread of this virus has led governments worldwide to implement unprecedented and far-reaching public health guidelines and lockdowns in an effort to flatten the infection curve and reduce the burden on their respective healthcare systems. Healthcare workers currently have the highest risk of exposure and can easily become vectors of viral transmission. On April 16th, 2020, the United Kingdom cabinet reported that 16.2% of positive cases were critical workers in NHS (Centre for Evidence-Based Medicine, 2020), and the United States-based CDC reported that up to 11% of positive cases were healthcare workers in some states (CDC, 2020a).

Symptomatic COVID-19 patients display symptoms including a dry cough, fever, shortness of breath and sore throat (CDC, 2020b), and in severe cases can experience severe pneumonia, pulmonary edema, organ failure, and death (Chen et al., 2020). The virus incubation period varies from 3 to 20 days, during which time a patient can be infectious while asymptomatic, as some evidence suggests (Bai et al., 2020). While all modes of transmission for SARS-CoV-2 are not completely understood, it is generally believed that the primary mechanism of transmission is through micron-sized droplets and aerosols produced when an infected person, sneezes, coughs, talks or breathes (WHO, 2020a). Accordingly, public health organizations around the world are urging citizens to practice physical distancing (i.e., “social distancing”), use facemasks when in public, frequently wash their hands, and disinfect commonly touched areas in their surroundings (CDC, 2020c; WHO, 2020b).

In most countries, many front-line healthcare professionals are required to wear approved N95 masks as well as face shields to protect themselves against infectious droplets and cross-contamination (CDC, 2020d). However, due to shortages of N95 masks and other protective PPE, health organizations including the CDC are now recommending extended use and reuse of PPE including N95 masks. These new recommended practices may put healthcare workers at additional risk if the PPE cannot be effectively disinfected between uses. Therefore, effective disinfection of protective masks and other PPE before reuse is critical (CDC, 2020c). One recent study (Fischer et al., 2020) validated several CDC-recommended PPE decontamination techniques by demonstrating effective disinfection of N95 masks using UV-radiation, dry heat, and hydrogen peroxide vapor application. However, authors showed that while ethanol was effective in disinfecting N95 masks, it also led to reduced filtration capacity in the mask. Another study suggested that ethanol- and chlorine-based disinfectants are not suitable for decontamination due to their capacity to adsorb on fabric material and interfere with their electrostatic properties, thereby reducing filtration capacity (Liao et al., 2020). In keeping with these findings, bleach, sanitizing wipes, and ethyl oxides are not recommended for use to decontaminate masks (SAGES, 2020). Recommended PPE decontamination methods (UV, dry heat, hydrogen peroxide vapors) cannot, however, be easily applied during times of extended daily use of PPE, warranting the need for new or additional disinfection methods that can be applied “on the go” in these settings without causing harm to adjacent skin.

Iodine-containing solutions such as povidone iodine (PVP-I) have been proven effective as surface disinfectants against a wide variety of viruses including influenza A, poliovirus, adenovirus type 3, mumps, SARS, MERS, and HIV (Eggers et al., 2015; Kawana et al., 1997; Wada et al., 2016), with some indication of greater viricidal spectrum of activity compared to other commercially available disinfectants. Furthermore, iodine-based formulations have also been shown to be an effective preventative method to reduce upper respiratory tract infections and oral tract pathogens (Eggers et al., 2018; Sakai et al., 2008).

Clyraguard copper iodine complex, developed by Clyra Medical Technologies, Inc., is a novel FDA-registered product intended to be used for decontaminating non-critical PPE. The formula has proven antimicrobial activity (FDA submission data, see here) and is cleared for use on skin and wounds. In contrast, other iodine-based products, such as Lugol’s Iodine and PVP-I, may cause staining and skin sensitivity.
In this study, Clyraguard copper iodine complex was assessed for its efficacy in inactivating SARS-CoV-2 using a Vero cell monolayer infection model to provide a basis for whether or not Clyraguard may represent a potentially effective tool to disinfect and prolong the useful protective life of PPE, as well as to provide additional antiviral protection to PPE such as masks.

**Methods**

**Cell lines and cell growth**

Vero cells were obtained from ATCC and grown in DMEM (Corning) containing 1x L-glutamine and 1% MEM vitamins, and supplemented with 10% FBS (Gibco) and 1% penicillin/streptomycin (Gibco). Cells were incubated at 37°C in 5% CO₂. Cells were used at 85–95% confluent growth.

**Virus stocks**

The SARS-CoV-2 (USA-WA1/2020) virus was obtained from The World Reference Center for Emerging Viruses and Arboviruses (WRCEVA), University of Texas Medical Branch, Galveston, TX. All experiments involving infectious virus were conducted by S.P.’s laboratory at the University of Texas Medical Branch (Galveston, TX) in approved biosafety level 3 laboratories in accordance with institutional health and safety guidelines and federal regulations.

**Preparation of SARS-CoV-2 virus stocks**

From the original stock, SARS-CoV-2 was propagated for one passage in infection medium (DMEM containing 1x L-Glutamine and 1% MEM vitamins, supplemented with 1% penicillin/streptomycin (Gibco) and 2% FBS) at 37°C in 5% CO₂. Briefly, SARS-CoV-2 was added at a MOI of 0.01 to Vero cells and incubated 1 hour at 37°C. Viral inoculum was removed, and fresh infection medium added. Cells were incubated for an additional 48 hours before supernatant was collected. Supernatant was centrifuged for 5 minutes at 3000 rpm to remove cell debris. Virus stocks were stored at -80°C at a concentration of 1x10⁵ median tissue culture infectious dose (TCID₅₀) per mL.

**Determination of viral titers**

Viral titers were measured by TCID₅₀ on Vero Cells in 96-well plates. Each log₁₀ dilution (10⁴ through 10⁰) of the virus was inoculated in quadruplicates. On day 4 post-infection, the cells were fixed with 10% formalin for 45 minutes and subsequently stained with crystal violet. Cleared wells were quantified to calculate titer.

**Virus inactivation using Clyraguard or controls**

Aliquots of stock virus (10 μl) were mixed with 90 μl of Clyra, diluted Clyraguard, or control saline solutions. Clyra was mixed either 1:10 or 1:100 with sterile saline for dilution. Room temperature water was used as a negative control for virus inactivation, while boiling water (water pre-heated to 100°C) was used as a positive control. All mixtures were incubated for 30 seconds, 10 minutes, 30 minutes, or 60 minutes at room temperature, at which time 900 μl infection medium was added to neutralize antiviral activity. Subsequently, the SARS-CoV-2 viral titer (TCID₅₀/mL) for each test substance was determined. The experiment was conducted in triplicate.

**Statistical analysis**

Statistical significance was determined using a two-way ANOVA with Dunnett’s multiple comparisons test comparing against the control saline group. This was conducted using GraphPad Prism (software version 8.3.0). A p-value of 0.05 or below was considered statistically significant.

**Results**

In this study, the ability of Clyraguard to inactivate SARS-CoV-2 at various timepoints and at various concentrations was assessed (Figure 1; Table 1). While diluted Clyraguard was unable to

![Figure 1. Inactivation of SARS-CoV-2 using Clyraguard.](image)

**Table 1. Mean and standard deviation for the SARS-CoV-2 titer (TCID₅₀/ml) calculated from three replicates.**

<table>
<thead>
<tr>
<th></th>
<th>30 sec.</th>
<th></th>
<th>10 min.</th>
<th></th>
<th>30 min.</th>
<th></th>
<th>60 min.</th>
<th></th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Mean</td>
<td>Std. Dev.</td>
<td>Mean</td>
<td>Std. Dev.</td>
<td>Mean</td>
<td>Std. Dev.</td>
<td>Mean</td>
<td>Std. Dev.</td>
</tr>
<tr>
<td>Saline</td>
<td>2.25 x 10⁵</td>
<td>1.95 x 10⁵</td>
<td>6.08 x 10⁴</td>
<td>3.91 x 10⁵</td>
<td>2.17 x 10⁵</td>
<td>2.00 x 10⁵</td>
<td>2.25 x 10⁵</td>
<td>1.95 x 10⁵</td>
</tr>
<tr>
<td>Clyraguard</td>
<td>1.92 x 10⁵</td>
<td>8.25 x 10⁴</td>
<td>1.12 x 10⁴</td>
<td>1.01 x 10⁴</td>
<td>&lt;7.5 x 10¹</td>
<td>0</td>
<td>&lt;7.5 x 10¹</td>
<td>0</td>
</tr>
<tr>
<td>Clyraguard 1:10</td>
<td>4.17 x 10⁵</td>
<td>2.36 x 10⁴</td>
<td>6.67 x 10⁴</td>
<td>2.36 x 10⁴</td>
<td>1.25 x 10⁵</td>
<td>8.90 x 10⁴</td>
<td>5.83 x 10⁴</td>
<td>2.36 x 10⁴</td>
</tr>
<tr>
<td>Clyraguard 1:100</td>
<td>2.00 x 10⁵</td>
<td>7.07 x 10⁴</td>
<td>5.00 x 10⁴</td>
<td>0</td>
<td>3.33 x 10⁵</td>
<td>1.18 x 10⁵</td>
<td>5.00 x 10⁴</td>
<td>2.04 x 10⁴</td>
</tr>
<tr>
<td>Boiling saline</td>
<td>4.17 x 10⁴</td>
<td>1.18 x 10⁴</td>
<td>&lt;7.5 x 10¹</td>
<td>0</td>
<td>&lt;7.5 x 10¹</td>
<td>0</td>
<td>&lt;7.5 x 10¹</td>
<td>0</td>
</tr>
</tbody>
</table>
inactivate SARS-CoV-2 after any length of time, undiluted Clyraguard was effective at significantly reducing viral titers after just 10 minutes of incubation. After incubation with undiluted Clyraguard for 10 minutes, viral titers dropped by 2 logs (p-value < 0.0001). Furthermore, after incubation with undiluted Clyraguard for either 30 minutes or 60 minutes, viral titers dropped below the limit of detection (<75 TCID\textsubscript{50} per ml).

**Discussion**

In this study, Clyraguard copper iodine complex was assessed for its ability to inactivate SARS-CoV-2 in liquid suspension. These data demonstrate that the product has significant viricidal activity against SARS-CoV-2 within 10 minutes and yields complete SARS-CoV-2 deactivation by 30 minutes.

Antiviral activity against human CoV viruses including SARS-CoV and MERS-CoV has been previously been demonstrated using iodine (Eggers et al., 2015; Eggers et al., 2018). The mechanism of action is not widely understood but researchers have hypothesized that iodine attacks viruses in multiple ways, attacking the protein structure and interfering with hydrogen bonding associated with cysteine, histidine, and tyrosine, thus altering virus membrane structure and causing inhibition of viral release and spread from infected cells (Qin, 2009).

Iodine-based materials such as PVP-I and Lugol’s solution have shown effective antiviral activity but also exhibit toxicity concerns and can only be tolerated on skin for short periods of time. The copper iodine complex investigated in this study has also been tested according to ISO 10993 standards (FDA submission data, see here) and found to be safe for both skin and wounds, suggesting that it may represent a potential alternative to current iodine-based regimens.

Long-term antimicrobial performance studies with organisms including *Enterococcus faecium*, *Staphylococcus aureus*, *Klebsiella pneumoniae*, *Acinetobacter baumannii*, *Pseudomonas aeruginosa*, *Enterobacter aerogenes*, *Bacillus fragilis* (FDA submission data, see here) have also been conducted with the Clyraguard formula, wherein it was found to be efficacious for up to 72 hours. It is therefore recommended that further studies be carried out under good laboratory practices to directly determine any extended activity against the SARS-CoV-2 virus. This would verify the potential extended use for PPE against SARS-CoV-2, potentially providing additional protection for healthcare professionals working during the COVID-19 pandemic.

In addition, further studies to assess the impacts (or lack thereof) of this copper iodine complex in combination with common face masks such as N95 masks on facial skin and wearer comfort are planned once a suitable and acceptable test method has been established.

Common existing decontamination practices used on PPE such as face masks include UV radiation, dry heat (ovens), and hydrogen peroxide vapor machines such as those approved for use by the FDA manufactured by Battelle and Steris (Rubio-Romero et al., 2020). While substantial evidence exists that supports the consistent antiviral efficacy of hydrogen peroxide vapor PPE decontamination devices (Steinberg et al., 2020), these devices typically require PPE to be reprocessed overnight due to the time required for adequate decontamination. The distinction between these decontamination techniques, which have commonly been used in hospitals during the SARS-CoV-2 pandemic and previous pandemics, and spray-on decontamination products such as the copper iodine complex Clyraguard is that the former require removal of the PPE followed by comparatively lengthy decontamination (typically overnight), whereas the latter is intended to be additional protection on the mask and can be used on-the-go without significant time commitment during a regular work day.

This study demonstrates clear evidence to the potential suitability of the copper iodine complex (Clyraguard) for decontaminating non-critical PPE to help mitigate cross-contamination of SARS-CoV-2. However, this study examined the efficacy of Clyraguard against SARS-CoV-2 only in liquid suspension, and therefore follow-up studies wherein this copper iodine complex is assessed for its efficacy against SARS-CoV-2 (and other viruses) adhered to materials used in personal protective equipment should also be conducted.

**Data availability**

Underlying data


This project contains the raw viral titers for each repeat produced in this experiment.

Data are available under the terms of the Creative Commons Zero "No rights reserved" data waiver (CC0 1.0 Public domain dedication).

**Acknowledgements**

We thank Drs. Kenneth Plante (The World Reference Center for Emerging Viruses and Arboviruses, UTMB) and Natalie Thornburg from the CDC for providing the SARS-CoV-2 stock virus. E.K.M was supported by NIH T32 training grant AI060549.
References


Centers for Disease Control and Prevention: Decontamination and Re-use for SARS-CoV-2. Reference Source


Open Peer Review

Current Peer Review Status:  ✔️  ✔️

**Version 2**

Reviewer Report 14 October 2020

https://doi.org/10.5256/f1000research.30025.r72577

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✔️ Paul Dabisch
National Biodefense Analysis and Countermeasures Center, Operated by BNBI for the US Department of Homeland Security Science and Technology Directorate, Frederick, MD, USA

The authors revisions are satisfactory and I have no additional comments.

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** My research focuses on the examination of factors influencing transmission of infectious diseases, including the influence of environmental conditions on the persistence of infectious material on surfaces and in aerosols.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

**Version 1**

Reviewer Report 18 August 2020

https://doi.org/10.5256/f1000research.27191.r68324

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❓ Paul Dabisch
National Biodefense Analysis and Countermeasures Center, Operated by BNBI for the US Department of Homeland Security Science and Technology Directorate, Frederick, MD, USA
This study describes novel data on the efficacy of a copper iodine complex for inactivation of SARS-CoV-2 in liquid suspension. The results demonstrate that the undiluted copper iodine complex causes a significant decrease in viral infectivity in a cell-based microtitration assay after 10 minutes, with greater than a 5-log reduction in 30 minutes. The methodologies and conclusions are generally appropriate. However, there are several limitations to the study:

1. The study only investigates the ability of the copper iodine complex to inactivate the virus in a liquid suspension. Thus, while the present study provides useful preliminary data, additional testing with virus deposited as droplets or aerosol particles on relevant surfaces needs to be conducted.

2. As a justification for the use of copper iodine complex, the authors discuss the incompatibility of certain disinfectants for N95 masks, as well as the difficulty of using other decontamination methods while PPE is being worn. However, while the authors cite some evidence that copper iodine complex is safe on skin, the present study does not perform testing to demonstrate the benefit of copper iodine complex in this regard, specifically its compatibility with N95 masks or ability to be used while PPE is being worn.

3. The manuscript would benefit from additional discussion/comparison of the results obtained in the study to those published for other decontamination methods for SARS-CoV-2, SARS-CoV-1, and MERS.

4. The methods state that a t-test was used for statistical comparisons. However, it is unclear from the text what comparisons were made. In looking at the data, it appears that either a one- or two-way ANOVA with a multiple comparisons post-test would be more appropriate.

5. Regarding data presentation, while Table 1 provides mean values, the standard deviation would also be helpful.

Is the work clearly and accurately presented and does it cite the current literature?
Yes

Is the study design appropriate and is the work technically sound?
Yes

Are sufficient details of methods and analysis provided to allow replication by others?
Partly

If applicable, is the statistical analysis and its interpretation appropriate?
Partly

Are all the source data underlying the results available to ensure full reproducibility?
Partly

Are the conclusions drawn adequately supported by the results?
Yes

Competing Interests: No competing interests were disclosed.
Reviewer Expertise: My research focuses on the examination of factors influencing transmission of infectious diseases, including the influence of environmental conditions on the persistence of infectious material on surfaces and in aerosols.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Author Response 16 Sep 2020
Alex Evans, BioLargo, Inc., Westminster, USA

Thank you for your review. Your comments have provided valuable insights, and we are presently working to edit the paper to address each of the issues raised in your comments. We hope to re-submit this new version in the coming weeks.

Competing Interests: No competing interests were disclosed.

Reviewer Report 06 July 2020
https://doi.org/10.5256/f1000research.27191.r66377

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Ilya Frolov
Department of Microbiology, University of Alabama at Birmingham, Birmingham, AL, USA

This manuscript presents new data, which are important to the field and public health. However, to make the results more reproducible by other groups, the authors need to provide the source of Vero cell and catalog number, number of cells seeded into the wells of 96-well plates and to clarify "infection medium."

Is the work clearly and accurately presented and does it cite the current literature?
Yes

Is the study design appropriate and is the work technically sound?
Yes

Are sufficient details of methods and analysis provided to allow replication by others?
Partly

If applicable, is the statistical analysis and its interpretation appropriate?
Yes
Are all the source data underlying the results available to ensure full reproducibility?
Partly

Are the conclusions drawn adequately supported by the results?
Yes

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** Virology

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

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