Cytoplasmic nanojunctions between lysosomes and sarcoplasmic reticulum are required for specific calcium signaling [version 1; peer review: 2 approved, 1 not approved]

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Abstract

Herein we demonstrate how nanojunctions between lysosomes and sarcoplasmic reticulum (L-SR junctions) serve to couple lysosomal activation to regenerative, ryanodine receptor-mediated cellular Ca²⁺ waves. In pulmonary artery smooth muscle cells (PASMCs) it has been proposed that nicotinic acid adenine dinucleotide phosphate (NAADP) triggers increases in cytoplasmic Ca²⁺ via L-SR junctions, in a manner that requires initial Ca²⁺ release from lysosomes and subsequent Ca²⁺-induced Ca²⁺ release (CICR) via ryanodine receptor (RyR) subtype 3 on the SR membrane proximal to lysosomes. L-SR junction membrane separation has been estimated to be < 400 nm and thus beyond the resolution of light microscopy, which has restricted detailed investigations of the junctional coupling process. The present study utilizes standard and tomographic transmission electron microscopy to provide a thorough ultrastructural characterization of the L-SR junctions in PASMCs. We show that L-SR nanojunctions are prominent features within these cells and estimate that the junctional membrane separation and extension are about 15 nm and 300 nm, respectively. Furthermore, we develop a quantitative model of the L-SR junction using these measurements, prior kinetic and specific Ca²⁺ signal information as input data. Simulations of NAADP-dependent junctional Ca²⁺ transients demonstrate that the magnitude of these signals can breach the threshold for CICR via RyR3. By correlation analysis of live cell Ca²⁺ signals and simulated Ca²⁺ transients within L-SR junctions, we estimate that “trigger zones” comprising 60–100 junctions are required to confer a signal of similar magnitude. This is compatible with the 110 lysosomes/cell estimated from our ultrastructural observations. Most importantly, our model shows that increasing the L-SR junctional width above 50 nm lowers the magnitude of junctional [Ca²⁺] such that there is a failure to breach the threshold for CICR via RyR3. L-SR junctions are therefore a pre-requisite for efficient Ca²⁺ signal coupling and may contribute to cellular function in health and disease.
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Introduction

The importance of cytoplasmic nanojunctions to cellular signaling and thus to the modulation of cell function was recognised several decades ago\(^\text{1,2}\), henceforth the extent to which cellular nanospaces may contribute to the regulation of cell function received little attention. Nevertheless there is now a growing recognition of the widespread occurrence and functional significance of cytoplasmic nanospaces in cells from species across several kingdoms\(^\text{3,4}\).

In this respect, membrane-membrane junctions between lysosomes and the sarcoplasmic reticulum (L-SR junctions) are of particular interest; not least because they have been hypothesized to couple calcium signaling between these two organelles\(^\text{5,6}\).

That L-SR junctions may play an important role in cellular \(\text{Ca}^{2+}\) signaling was uncovered through early studies on the \(\text{Ca}^{2+}\) mobilizing messenger nicotinic acid adenine dinucleotide phosphate (NAADP)\(^\text{7}\), which demonstrated that NAADP released \(\text{Ca}^{2+}\) from a store other than the sarco/endoplasmic reticulum (S/ER)\(^\text{8}\), that could then trigger further \(\text{Ca}^{2+}\) release from the S/ER by \(\text{Ca}^{2+}\)-induced \(\text{Ca}^{2+}\) release (CICR)\(^\text{9,10}\). A major advance in our understanding was then provided by the demonstration that the NAADP-released \(\text{Ca}^{2+}\) was from an acidic lysosome-related store\(^\text{11-13}\) in a manner that requires two pore segment channel subtype 2 (TPC2)\(^\text{14}\). However, studies on pulmonary arterial smooth muscle cells (PASMCs) had also identified a significant specialization, namely L-SR nanojunctions. It was hypothesized not only that these nanojunctions were necessary for coupling between lysosomes and the SR, but that they could both coordinate and restrict their relationship to the SR by preferentially targeting ryanodine receptors while excluding inositol 1,4,5-trisphosphate (IP\(_3\)) receptors\(^\text{15,16}\). Importantly, NAADP-dependent \(\text{Ca}^{2+}\) bursts primarily arise from lysosomes in the perinuclear region of PASMCs and appear to promote rapid, local \(\text{Ca}^{2+}\) transients that are of sufficient size to activate clusters of SR resident ryanodine receptor subtype 3 (RyR3) and thus initiate, in an all-or-none manner, a propagating global \(\text{Ca}^{2+}\) wave\(^\text{17}\).

The specialization of the proposed L-SR junction is likely important in smooth muscle cell physiology, e.g., in vasomotion, and in the recycling of organelles and programmed cell death by autophagy\(^\text{21}\). However, L-SR junctions may also make as yet unforeseen contributions to vascular pathologies as highlighted by the fact that Niemann-Pick disease type C1 results, in part, from dysregulation of lysosomal \(\text{Ca}^{2+}\) handling\(^\text{22}\) and is known to precipitate portal hypertension\(^\text{23}\), while other lysosomal storage diseases (e.g., Pompe and Gaucher disease) accelerate pulmonary arterial hypertension\(^\text{24-26}\). Moreover, observed hypertension is often associated with dysfunction of cholesterol trafficking\(^\text{27}\), increased plasma cholesterol levels, vascular lesion formation, atherosclerosis/thrombosis and medial degradation\(^\text{25,27}\). Therefore lysosomal \(\text{Ca}^{2+}\) signaling is of considerable clinical interest. That L-SR junctions may be of further significance to pathology is also evident, for example, from the fact that in the pulmonary artery smooth muscle L-SR junctions underpin \(\text{Ca}^{2+}\) waves initiated by endothelin 1, the levels of which are elevated in pulmonary hypertension, systemic hypertension and atherosclerosis\(^\text{28,29}\). An understanding of how specific \(\text{Ca}^{2+}\) signals are functionally initiated therefore has important translational implications.

Lysosomal \(\text{Ca}^{2+}\) regulation has been of current interest in several recent research and review articles (e.g.,\(^\text{30-34}\)), yet a quantitative elucidation of the mechanism by which \(\text{Ca}^{2+}\) signals are generated by the endolysosomal system is still lacking. Given the likely importance of L-SR junctions to \(\text{Ca}^{2+}\) signaling in health and disease, we sought to determine whether this nano-environment would indeed be able to effectively generate the previously observed NAADP-induced \(\text{Ca}^{2+}\) signals. In particular, we set out to verify the following hypotheses in PASMCs: (1) L-SR nanojunctions should be observable in the ultrastructure of these cells, (2) NAADP induces sufficient \(\text{Ca}^{2+}\) release from the lysosome to initiate activation of RyR3 embedded in the junctional SR, and (3) the combined effect of activation of L-SR nanojunctions in a cytoplasmic “trigger zone” determines the threshold of global [\(\text{Ca}^{2+}\)] for the biphasic release process.

Due to the minute spatial scale of the nanojunctions generating the primary \(\text{Ca}^{2+}\) signals, accurate investigation of dynamic signaling within these spaces cannot be achieved with currently available instrumentation. To overcome this issue, we took an integrative approach by combining our own electron microscopy of L-SR nanojunctions, existing kinetic data on the \(\text{Ca}^{2+}\) transporters and buffers, and prior knowledge of the NAADP-induced \(\text{Ca}^{2+}\) signal features into a quantitative model of a typical L-SR nanojunction. This model is based on stochastic simulations of intracellular \(\text{Ca}^{2+}\) diffusion by Brownian motion implemented using the particle simulator MCell\(^\text{35-37}\).

In the present manuscript we have verified the existence of L-SR nanojunctions within the ultrastructure of PASMCs, and shown that lysosomes can release sufficient \(\text{Ca}^{2+}\) to activate CICR via RyR3 clusters embedded in the junctional SR. Perhaps most importantly, we show that L-SR coupling is determined both by the integrity of L-SR junction on the nanoscale and the quantal summation of \(\text{Ca}^{2+}\) release from multiple, activated junctional complexes.

Materials and methods

All the experiments and procedures were carried out in accordance with the guidelines of the University of British Columbia Animal Care Committee, and in accordance with the United Kingdom Animals (Scientific Procedures) Act 1986. The Research Ethics Committees from both these institutions specifically and independently approved this study.

Electron microscopy

An adult male Wistar rat (about 12 months old, weighing 220–250 g, obtained from Charles River and housed in the institutional animal facility at the University of British Columbia Child and Family Research Institute under standard animal room conditions of 12 h light-12 h dark cycles, at 25°C, with two animals in a cage) was anesthetized with 3 ml of Thiotal (Vétouquin, Lavaltrie, QC, Canada). For the purpose of this phase of the study, one representative healthy animal usually suffices. This choice also minimizes unnecessary animal sacrifice. The thoracic cavity was opened and flooded with warm physiological saline solution (PSS) containing 145 mM
NaCl, 4 mM KCl, 1 mM MgCl₂, 10 mM HEPES, 0.05 mM CaCl₂, and 10 mM glucose (pH 7.4; all chemicals were purchased from Sigma-Aldrich Canada unless otherwise stated). After extraction of the heart and lungs and their placement in HEPES buffer, several rings from the primary and secondary branches of the pulmonary artery were dissected and immediately immersed in fixative solution. The primary fixative solution contained 2.5% glutaraldehyde (Ted Pella, Redding, CA, USA) in 0.1 M sodium cacodylate buffer (Canemco & Marivac, Gore, QC, Canada) at room temperature. The artery rings were then washed three times in 0.1 M sodium cacodylate (30 min in total). In the process of secondary fixation, the tissue rings were fixed with 1% OsO₄ (Ted Pella, Redding, CA, USA) in 0.1 M sodium cacodylate buffer for 1 h followed by three 10-minute washes with distilled water and en bloc staining with 2% uranyl acetate (Ted Pella, Redding, CA, USA). The samples were then dehydrated in increasing concentrations of ethanol (25, 50, 75, 80, 90, and 95%). In the final process of dehydration, the samples underwent 3 washes in 100% ethanol. The artery rings were then resin-infiltrated in increasing concentrations (30, 50, and 75% in ethanol) of a 1:1 mix of Epon and Spurr’s resins (Canemco & Marivac, Gore, QC, Canada). The infiltration process was completed by three passages in 100% resin. All of the ethanol dehydration and resin infiltration steps were carried out by using a laboratory microwave oven (Pelco 3441, Ted Pella, Redding, CA, USA). The blocks were then dehydrated in increasing concentrations of ethanol (25, 50, 75%, 80, 90, and 95%). In the final process of dehydration, the samples underwent 3 washes in 100% ethanol. The artery rings were then resin-infiltrated in increasing concentrations (30, 50, and 75% in ethanol) of a 1:1 mix of Epon and Spurr’s resins (Canemco & Marivac, Gore, QC, Canada). The infiltration process was completed by three passages in 100% resin. All of the ethanol dehydration and resin infiltration steps were carried out by using a laboratory microwave oven (Pelco 3441, Ted Pella, Redding, CA, USA). The blocks were then resin-embedded in molds and polymerized overnight in an oven at 60°C.

2D imaging

For standard (2D) electron microscopy imaging, 80-nm sections were cut from the embedded sample blocks on a Reichert Ultracut-E microtome (Diatome, Biel, Switzerland) and were collected on uncoated 100- and 200- mesh copper grids (hexagonal or square meshes; Ted Pella, Redding, CA, USA). The sections were then post-stained with 1% uranyl acetate (Canemco & Marivac, Gore, QC, Canada; pH not recorded) and Reynolds lead citrate (Fisher Scientific Company, USA; pH not recorded) for 12 and 6 minutes, respectively. Electron micrographs at various magnifications were obtained with a Hitachi 7600 transmission electron microscope (Hitachi High Technologies, Japan) at 80 kV.

Lysosomes in these images were identified according to their well-established appearance and features: they are bound by a single lipid-bilayer membrane, with a granular, more or less uniform laminar matrix that is more electron dense than the surrounding cytosol. Secondary lysosomes may also contain less granular structures within the finer matrix. Moreover, lysosomes are normally distinguishable from endosomes by their larger size, hence we set a threshold “diameter” of > 200 nm for acceptance of a lysosome, below which all vesicles were excluded.

Tomography (3D imaging)

To obtain electron microscopic tomograms, we cut 200-nm-thick sections from the same sample blocks used for standard imaging. The sections were then collected on Formvar (Canemco & Marivac, Gore, QC, Canada) coated slot copper grids (Ted Pella, Redding, CA, USA) and post-stained with 1% uranyl acetate and Reynolds lead citrate for 20 and 10 minutes, respectively. We surveyed the sample sections using a FEI Tecnai G2 200 kV transmission electron microscope (FEI Technologies, Eindhoven, The Netherlands) and identified regions of interest containing L-SR junctions. We then acquired tomograms of several of those regions by taking 2D scans through the sample every 5° of inclination as it was tilted between −60° and +60° with respect to horizontal. The scans were reconstructed with the Inspect3D (FEI Technologies, Eindhoven, The Netherlands) software tools and structures of interest, primarily lysosomal and SR membranes in the same cellular neighbourhood, were traced out using Amira (FEI Technologies, Eindhoven, The Netherlands) software.

Image and data analysis

The images of the samples were further processed using GIMP (GNU Imaging Manipulation Program, open source, available at gimp.org) to enhance membrane contrast in the interest of improving our characterization of the L-SR junctions.

The SR and lysosomal membranes were outlined, highlighted and measured in pixels using the Inkscape software program (open source, available at http://www.inkscape.org), converting the pixel measurements to nm using the scale bar in the recorded micrographs. By modifying the Inkscape script for measuring lengths, we were able to output the measurements directly into a text file, which we used to produce the histograms in Figure 3B–D. We used the Gnuplot program (open source, available at http://www.gnuplot.info) to produce the histograms and plots presented herein.

Ca²⁺ imaging

To obtain sufficient cell isolation to appropriately test significance, 1–3 adult male Wistar rats (weighing 150–300 g, obtained from Charles River and housed in the institutional animal facility at the Centre for Integrative Physiology of the University of Edinburgh under standard animal room conditions of 12 h light-12 h dark cycles, at 25°C, with 2–4 animals in a cage) were sacrificed by cervical dislocation. The heart and lungs were removed en bloc and placed in physiological salt solution (PSS) of the following composition (mM): 130 NaCl, 5.2 KCl, 1 MgCl₂, 1.7 CaCl₂, 10 glucose, and 10 HEPES, pH 7.45 (all chemicals were purchased from Sigma-Aldrich). Single arterial smooth muscle cells were isolated from second-order branches of the pulmonary artery. Briefly, arteries were dissected out and placed in low Ca²⁺ solution of the following composition (mM): 124 NaCl, 5 KCl, 1 MgCl₂, 0.5 NaH₂PO₄, 0.5 KH₂PO₄, 15 NaHCO₃, 0.16 CaCl₂, 0.5 EDTA, 10 glucose, 10 taurine and 10 HEPES, pH 7.4. After 10 min the arteries were placed in the same solution containing 0.5 mg/ml papain (Fluka) and 1 mg/ml bovine serum albumin (Sigma) and kept at 4°C overnight. The following day 0.2 mM 1,4-dithio-DL-threitol (Sigma) was added to the solution, to activate the protease, and the preparation was incubated for 1 h at room temperature (22°C). The tissue was then washed at 3°C in fresh low Ca²⁺ solution without enzymes, and single smooth muscle cells were isolated by gentle trituration with a fire-polished Pasteur pipette. Cells were stored in suspension at 4°C until required.

PASMCs were incubated for 30 min with 5 µM Fura-2-AM in Ca²⁺-free PSS in an experimental chamber on a Leica DMIRBE inverted microscope and then superfused with Fura-2 free PSS for at least
30 min prior to experimentation. Intracellular Ca\(^{2+}\) concentration was reported by Fura-2 fluorescence ratio (F340/F380 excitation; emission 510 nm). Emitted fluorescence was recorded at 22°C with a sampling frequency of 0.5 Hz, using a Hamamatsu 4880 CCD camera via a Zeiss Fluar 40×, 1.3 n.a. oil immersion lens and Leica DMIRBE microscope. Background subtraction was performed on-line. Analysis was done via Openlab imaging software (Improvision, UK).

NAADP was applied intracellularly in the whole-cell configuration of the patch-clamp technique, and in current clamp mode (I = 0) as described previously\(^{17}\). The pipette solution contained (in mM): 140 KCl, 10 HEPES, 1 MgCl\(_2\), and 5 µM Fura-2, pH 7.4. The seal resistance, as measured using an Axopatch 200B amplifier (Axon Instruments, Foster City, CA), was ≥ 3 GΩ throughout each experiment. Series resistance and pipette resistance were ≤ 10 MΩ and ≤ 3 MΩ, respectively. All experiments were carried out at room temperature (≈ 22°C).

Quantitative modeling
The main stages of the quantitative modeling approach are:

1. The design of 3D software mesh objects (nets of interconnected triangles by which surfaces can be represented in computer graphics) representing a typical L-SR region, including a whole lysosome and a portion of neighbouring SR network. These objects are built to-scale following the ultrastructural characterization of the L-SR junctional regions as it results from our electron microscopy image analysis; this phase was carried out using the “3D content creation suite” Blender (open source, available at blender.org);

2. The positioning of the relevant transporters on the reconstructed membranes (TPC2 complexes on the lysosome, SERCA2a and RyR3 on the SR) according to information gathered from the literature on their typical membrane densities and the implementation of the transporter’s known kinetics and multi-state models and of the ion diffusivities (Ca\(^{2+}\); mobile Ca\(^{2+}\) buffers);

3. The simulation of molecular Brownian motion in the cytosol by random walk algorithms; this phase was performed by writing appropriate code for the stochastic particle simulator MCell (freely available at http://www.mcell.org)\(^{10,17}\). In a nutshell, MCell reproduces the randomness of the molecular trajectories, of the ion transporter flickering and of the relevant chemical reactions by probabilistic algorithms governed by pseudo-random number generators (iterative mathematical algorithms, which produce a pseudo-random sequence of numbers once initiated by a given number called seed). This enables the simulation of a number of microphysiological processes, all stochastically different from one another. The average outcome of the processes, to mimic the instrumental output during experimental measurements, is obtained by taking the average of a desired quantity, e.g., [Ca\(^{2+}\)], over a large number of simulations all initiated with a different seed;

4. The measurement of simulated [Ca\(^{2+}\)] in the L-SR junctions from the process of Ca\(^{2+}\) release via TPC2-related signaling complex on the lysosome and SR Ca\(^{2+}\) uptake by the SERCA2a pumps, and static as well as dynamic visualization of the simulations; this stage is part of MCell’s output.

Results
The work presented in this article is based on earlier Ca\(^{2+}\) fluorescence imaging observations by optical microscopy, of which the data in Figure 1 is a representative example, and immunofluorescence results. These studies led to the proposal that, for the lysosomal Ca\(^{2+}\) release to trigger CICR, L-SR nanojunctions are required and that they consist of apposing patches of lysosomal and SR membranes separated by a narrow space of nano-scale dimension\(^{10,17}\). The relevant background findings that stimulated the development of the work exposed here were first reported by Evans’ group\(^{10,17}\), and are summarized in Figure 1.

The example record in Figure 1 highlights the fact that NAADP appears to activate a two-phase Ca\(^{2+}\) signal, which can exhibit an identifiable “shoulder” during the initial rising phase of the signal (Figure 1, time point 1), followed by a second faster phase of signal amplification (Figure 1, time point 2). Previous studies have demonstrated that this two-phase response results from initiation by NAADP of Ca\(^{2+}\) bursts from lysosome-related stores\(^{17}\) in a manner that requires TPC2\(^{38}\) and that Ca\(^{2+}\) bursts are subsequently amplified by CICR from the SR via clusters of RyR3\(^{10,17}\). The studies cited above provided calibrated estimates of the changes in intracellular [Ca\(^{2+}\)] input into the model below. We propose herein that L-SR junctions do indeed represent cellular nanojunctions and that they

![Figure 1](image-url)
might play a role of accentuating Ca\(^{2+}\) gradients, akin to that of plasma membrane (PM)-SR junctions that are pivotal in the process of SR Ca\(^{2+}\) refilling during asynchronous [Ca\(^{2+}\)] waves\(^{39}\). We furthermore hypothesize that in order for these nanojunctions to appropriately regulate Ca\(^{2+}\) signaling, they must be separated by a distance of approximately 20 nm and be of the order of a few hundred nm in lateral dimensions, as inferred from previous studies on PM-SR junctions\(^{10,11}\).

Ultrastructural characterization of L-SR nanojunctions

To identify lysosomes, SR regions and L-SR nanojunctions, we recorded and surveyed 74 electron micrographs of rat pulmonary arterial smooth muscle taken from samples prepared as described in the Materials and methods section. The images in Figure 2 provide a set of examples. Since we were aiming to detect L-SR junctions, namely close appositions of the lysosomal and SR membranes, immuno-gold labeling of lysosomes was prohibited, since this technique compromises membranes definition by electron microscopy to the extent that we would be unable to assess junctional architecture. Instead, in images like those in Figure 2, lysosome identification was accomplished by relying on the knowledge of lysosomal ultrastructural features, which has accumulated over the past 50 years since the discovery of the lysosomes\(^{40}\). In standard (2D) transmission electron microscopy (TEM) images, lysosomes typically appear as elliptical structures bound by a single lipid bilayer, a feature that distinguishes them from mitochondria. Depending on the lysosomal system stage, they also tend to have a more or less uniformly electron-dense interior as compared to the

![Figure 2](image-url). Representative electron micrographs of rat pulmonary artery SMC regions containing lysosomes (L), several SR cisterns, and including several examples of L-SR junctions (arrows). Also indicated are nuclei (N), Golgi apparatus (G), mitochondria (M), a multivesicular body (MVB) and extra-cellular space (ECS). Scale bars: 500 nm. Magnifications: A, C 80,000×, B, D 70,000×.
surrounding cytosol. They can be distinguished from endosomes by their larger size and darker lumen and they differ from peroxi-
somes, since the latter usually display a geometrically distinct and
markedly darker structure called “crystalloid” in their luminal area.
Moreover, it would appear that peroxisomes are found far more fre-
quently in liver, kidney, bronchioles and odontoblasts than in other
cell types (see, for example, http://www.uni-mainz.de/FB/Medi-
zin/Anatomie/workshop/EM/EMPeroxisomE.html).

Occasionally, organellar remnants are still visible inside these
ovas, a characteristic that identifies them as multi-vesicular bod-
ies (“MVB” in Figure 2A). As it is at times questionable whether
MVB’s are late endosomes or endosome-lysosome hybrids, we
have excluded organelles (3 in total) with such characteristics from
our statistical count.

From each of the relevant smooth muscle regions surveyed, we
obtained high-resolution images of areas containing lysosomes and
L-SR junctions in order to quantitatively characterize them (arrows
in Figure 2 and Figure 3A). Using a software graphics editor
(inkscape.org) and the image scale bar as a calibration gauge, we
measured the lysosome size, as the length of the major and minor
axes of their elliptical 2D projections (in orange and grey, respecti-
vely, in Figure 3A), the L-SR widths, that is the distance between
lysosomal and SR membranes at places where the two were about
30 nm or closer to each other (in purple in Figure 3A), and the
L-SR extensions as a percentage ratio between the junctional SR
and the lysosomal membranes (in turquoise in Figure 3A). From
these measurements, we extrapolated the 3D junctional SR exten-
sion, both as a percentage of the lysosomal surface and as a length
in nm. The histograms displayed in Figure 3B–D visually sum-
marize the data collected from the image analysis. The mean and
standard deviation values of the measured parameters are reported
in Table 1.

Estimates of the various parameters gathered in this phase of the
study were used as a basis to build a 3D software object to represent a
typical L-SR junction. This reconstruction was then used to design
the simulations mimicking Ca²⁺ diffusion in the L-SR nanojunc-
tions, as is described below.

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**Figure 3.** A, High magnification (150,000×) electron micrograph of a region of Figure 2B containing 3 L-SR junctions (arrows); coloured tracings as shown were used to measure lysosome dimensions, L-SR widths and extensions. Scale bar: 100 nm. B–D, Histograms showing distribution of several relevant lysosomal and L-SR junctional parameters, used to characterize the junctions and inform the quantitative model. B, lysosomal dimensions as major and minor axes of oval shapes in micrographs; C, L-SR junctional width; D, percentage apposition between junctional SR and lysosome perimeter as projected in 2D micrographs.
Tomography

To gather more direct information on the 3D morphology of L-SR junctions, we acquired a set of tomograms of those regions from the same sample blocks used to obtain the images in Figure 2. In Figure 4, we report snapshots from one of the tomograms; in these stills, we have also traced out parts of one lysosome and the closely apposed SR region that together form a L-SR junction. These tomograms are very helpful in clarifying the detailed morphology of L-SR junctions and informing on the spatial variability of the SR network. For example, while the SR segment shown in a single 2D tomographic scan (Figure 4A) would appear to be continuous and part of a large SR compartment, it actually branches out into narrower cisterns as revealed by 3D tomographic reconstruction (Figure 4B). Furthermore, it is interesting to observe the fact that one extension of the SR appears to couple with multiple organelles. Thus, the 3D views generated by tomography are paramount for demonstrating the presence of a true junctional complex and for the design of a prototypical L-SR environment as a software mesh object, on which we may simulate the NAADP-mediated localized Ca\(^{2+}\) release.

Quantitative model

The model aims to verify whether NAADP-induced Ca\(^{2+}\) release from the lysosomal system could be responsible for the localized Ca\(^{2+}\) signal preceding the global Ca\(^{2+}\) wave (see Figure 1), which triggers a propagating wave by CICR via RyR3s localized at L-SR junctions, as predicted by earlier observations.

Generation of the signal's “shoulder” by lysosomal Ca\(^{2+}\) bursts

To understand the generation of the signal “shoulder”, such as that observed at time point 1 in the lower panel of Figure 1, by Ca\(^{2+}\) bursts within L-SR nanojunctions, let us note that its magnitude corresponds to the difference in [Ca\(^{2+}\)] between the resting level of \(\approx 100\) nM prior to NAADP stimulation (up to the point, at which NAADP enters the cytoplasm under the whole-cell configuration (WC) in Figure 1) and the value of \(\approx 400\) nM, corresponding to [Ca\(^{2+}\)] at time point 1 in the example record shown in Figure 1, lower panel (these concentration values were obtained via a standard calibration procedure). This leads to a \(\Delta[Ca^{2+}]_{\text{shoulder}}\) of approximately 300 nM. Let us now estimate the number of lysosomes required to generate a \(\Delta[Ca^{2+}]_{\text{shoulder}}\) of such magnitude in a PASMC, given the following assumptions:

1. That the luminal [Ca\(^{2+}\)] of a lysosome, [Ca\(^{2+}\)]\(_{\text{lys}}\), is in the range of 400–600 µM, as determined in mouse macrophages and that it is homogeneous across the lysosomal population;
2. That the Ca\(^{2+}\) release rate during bursts is \(\approx 10^6\) ions/s (based on other experimenters’ findings, but see next section for a detailed analysis on the rate time-variation) and that it becomes negligible for values of [Ca\(^{2+}\)]\(_{\text{lys}}\) below \(\approx 80\) µM, as suggested by the single channel kinetics of TPC2 signaling complexes in lipid bilayer studies;
3. That lysosomes are spheroids with dimensions gathered from our EM characterization (see Figure 2, Figure 3, Figure 4).

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<tr>
<th>Table 1. L-SR junction characterization parameters.</th>
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<td><strong>Parameter</strong></td>
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<tr>
<td>lysosome minor axis</td>
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<tr>
<td>lysosome major axis</td>
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<tr>
<td>L-SR width</td>
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<tr>
<td>L-SR overlap</td>
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<tr>
<td>L-SR lateral extension</td>
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<td>3D L-SR overlap</td>
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Mean and standard deviation values of L-SR junction parameters, calculated from data as in Figure 3B–D.

Figure 4. A. Snapshot from a TEM tomogram of a L-SR region of rat pulmonary artery smooth muscle, illustrating, among other things, a single SR extension apparently forming junctions with several lysosomes. Magnification: 62,000x. B. Same snapshot shown in A, but with a lysosome (orange) and a portion of SR (turquoise) partially traced out in 3D. These pseudocolor tracings underscore how the SR can appear as a large cistern in a given plane, but can actually branch out in different directions when viewed in 3D. Scale bars \(\approx 100\) nm.
and Table 1), and hence that their volume is on average \( V = 2.2 \times 10^{-17} \) L;

4. That a smooth muscle cell cytosolic volume can be calculated as \( V_{\text{cyt}} = 2.4 \times 10^{-13} \) L by modeling a cell as a 130-µm-long cylinder, 6 µm in diameter\(^2\), and accounting for nuclear, SR, mitochondrial and lysosomal volumes as described in the supplementary material.

With these assumptions accepted, from points 1. and 2. above we gather that a lysosome can release a potential \([\text{Ca}^{2+}]_{\text{lys}} = (400 \text{ to } 600) \, \mu{\text{M}} – 80 \, \mu{\text{M}} = 3.2 \times 10^{-4} \, \text{M} \) into the cytosol (as we elaborate in the next section, this should take \( \approx 0.03 \) s, a time frame that ensures that released Ca\(^{2+}\) can be considered unbuffered).

Each lysosome contribution to \([\text{Ca}^{2+}]_i\), \([\text{Ca}^{2+}]_{\text{lys}}\), can then simply be calculated by taking the lysosome-to-cell volume ratio into account:

\[
\Delta[\text{Ca}^{2+}]_{\text{lys}} = \Delta[\text{Ca}^{2+}]_i \frac{V_{\text{lys}}}{V_{\text{cyt}}} = 3 \text{ to } 5 \, \text{nM} \tag{1}
\]

Therefore, we can calculate the number of lysosomes that may contribute to the magnitude of \( \Delta[\text{Ca}^{2+}]_{\text{shoulder}} \) as

\[
\frac{\Delta[\text{Ca}^{2+}]_{\text{shoulder}}}{\Delta[\text{Ca}^{2+}]_{\text{lys}}} = 60 \text{ to } 100 \text{ lysosomes} \tag{2}
\]

In summary, between 60 and 100 lysosomes would be necessary (and possibly sufficient) to provide a \( \Delta[\text{Ca}^{2+}]_{\text{shoulder}} \) of 300 nM, which is typically observed during the localized Ca\(^{2+}\) release phase of the NAADP-induced Ca\(^{2+}\) signal.

How many lysosomes do we actually expect to be in a PASM cell of our sample tissue? We can obtain a rough estimate of this number from the TEM imaging we performed for this study. In each 80-nm-thick TEM sample section, we see 5–10 lysosomes/cell. Lysosomes are predominantly localized to the perinuclear region of the cytoplasm, but seldom in the subplasmalemmal area, consistent with previous observations by optical microscopy\(^{7,11}\). If we simplify the geometry of a typical smooth muscle cell to a 130-µm-long cylinder with radius of 6 µm, and considering that lysosome radii are around 180 nm, it is reasonable to assume that separate sets of lysosomes would be observable in TEM images taken at distances into the sample of about 360 nm from each other. Neglecting the slices within one lysosomal diameter of the cylinder surface—given the near total lack of observed lysosomes in those subvolumes—then our images suggest that we can expect a total of about 110 lysosomes/cell. It is encouraging that we obtain from this count a higher number than the 60–100 lysosome range we derived in Equation (2), in that on the one hand it is plausible to think that not all of the lysosomes in a cell may be activated in synchrony, nor may they all be involved in NAADP-mediated signaling, and on the other hand experience tells us that evolution has built in some redundancy of function in order to provide a threshold and also a margin of safety for the generation of this type of Ca\(^{2+}\) signals. Moreover, the estimated number of junctions is based on a value for \([\text{Ca}^{2+}]_{\text{lys}}\) determined in macropatches, and it is plausible that the total releasable \([\text{Ca}^{2+}]_{\text{lys}}\) in a PASM may differ from that value and that it may also vary over the lysosome population. Lastly, the uncertainty in the number of lysosomes/cell emerging from the electron micrographs as described above may also contribute to this discrepancy.

**L-SR junctional Ca\(^{2+}\) signal**

In the hypothesized model outlined previously\(^{10,11}\), Ca\(^{2+}\) bursts activate SR resident RyR3 channels within L-SR junctions and thus initiate a propagating Ca\(^{2+}\) wave by CICR. The stochastic simulations developed here attempt to reproduce the phenomenon of the generation of \([\text{Ca}^{2+}]_i\) transients within individual L-SR junctions, considering the Ca\(^{2+}\) release kinetic requirements for lysosome-resident TPC2 signaling complexes and the rate of Ca\(^{2+}\) capture by the SERCA2a localized on the neighbouring SR membrane\(^{15}\), and to link these junctional transients to the observed bursts.

The thorough quantitative image analysis described in the previous section yields critical information for our first modeling phase, in which we built a dimensionally accurate virtual lysosome and a portion of the SR system, closely apposing the lysosome (Figure 5), so as to reproduce a NAADP-triggered Ca\(^{2+}\) signal within the nanospace of a representative L-SR junction as faithfully as possible.

From the available literature we estimated the number of TPC2 and SERCA2a likely distributed on the lysosome and SR membranes, respectively. We obtained the former number by dividing the macroscopic whole-lysosome conductance, calculated from the current values reported in a recent study on TPC2-mediated Ca\(^{2+}\) current in isolated lysosomes\(^{48}\), by the single channel conductance\(^{45}\), thus estimating that a typical lysosomal membrane may contain \(\approx 20\) TPC2. To obtain this value, we used experimental condition data\(^{46}\) to extract values for the ion potential, \(E_{\text{ion}}\), across the lysosomal membrane. We then employed the authors’ current-voltage (\(I-V_m\)) data to compute whole-lysosome conductance values (\(g_{\text{m,ion}}\)) as a function of the applied membrane potential, \(V_m\), from Ohm’s law:

\[
I = g_{\text{m,ion}}(V_m - E_{\text{ion}})\tag{3}
\]

We report details on this calculation in the supplementary material.

Moreover, we have estimated the density of SERCA2a on the SR within L-SR junctions to be equivalent to that previously predicted for PM-SR junctions as approximately 6250/µm\(^2\). Other input data for the model are the estimates of lysosomal volume and of the \([\text{Ca}^{2+}]_{\text{lys}}\), from which we calculated the actual number of ions in the lysosome prior to the beginning of Ca\(^{2+}\) release (see previous section). This, in turn, was used in the extrapolation of the TPC2 complex Ca\(^{2+}\) release rate as a function of time, as follows.

A recent electrophysiological study of the Ca\(^{2+}\) conductance of TPC2 signaling complex provides valuable information regarding its biophysical properties and, importantly, highlights the fact that, for a given relatively low activating concentration of NAADP (10 nM, as in Figure 1), the channel open probability appears to depend on \([\text{Ca}^{2+}]_{\text{lys}}\) (this is likely due to a partial neutralization of the electrochemical potential across the lysosomal membrane)\(^{45}\). We used the Ca\(^{2+}\) conductivity measured in that study and values of the lysosome membrane potential\(^{49}\) to calculate the maximal Ca\(^{2+}\) current, \(I_{\text{max}}\), as \(2.4 \times 10^9\) ions/s. We then applied a weighted quadratic fit to the channel’s open probability \(P_o\) as a function of the reported \([\text{Ca}^{2+}]_{\text{lys}}\) data with constraints that the \(P_o\) would tend to zero at \([\text{Ca}^{2+}]_{\text{lys}} = 80 \, \mu{\text{M}}\), based on the observation that below
\[ [\text{Ca}^{2+}]_{\text{lys}} = 100 \, \mu\text{M} \]

essentially no single channel openings were observed\(^4\). Another constraint for the fit was that the curve be within the standard deviation value of \( P_\circ \) at the highest reported \([\text{Ca}^{2+}]_{\text{lys}} = 1 \, \text{mM} \). From the quadratic fit, we then obtained our own \( P_\circ\)-vs-\([\text{Ca}^{2+}]_{\text{lys}} \) table (plotted in Figure 6A) and assumed that at the beginning of \( \text{Ca}^{2+} \) release, the \( \text{Ca}^{2+} \) current would be \( P_{\circ,\text{max}} \times I_{\max} \) at the maximal luminal concentration until the luminal concentration decreased to the next point in the \( P_\circ\)-vs-\([\text{Ca}^{2+}]_{\text{lys}} \) relationship. At this point, the release rate decreases to a new (lower) \( P_\circ \times I_{\max} \) until the luminal concentration reaches the next lower point in the table, and so on until \( P_\circ = 0 \) at \([\text{Ca}^{2+}]_{\text{lys}} = 80 \, \mu\text{M} \). The \( \text{Ca}^{2+} \) release rate as a function of time obtained in this manner is shown in Figure 6B.

To implement an approximated SERCA pump action, we used a simplified version of a multi-state kinetic model developed earlier\(^5\), which was further informed by more recent isoform-specific studies\(^51,52\). In particular, since we are interested in simulating the SERCA2a \( \text{Ca}^{2+} \) uptake only in terms of its influence on the shaping of the junctional \([\text{Ca}^{2+}]_\text{NJ} \) transient, we have opted not to implement the steps of the multistate model that deal with the \( \text{Ca}^{2+} \) unbinding from the SERCA on the SR luminal side of the pump. In brief, the reactions SERCA2a undergo are: (1) binding/unbinding of the first \( \text{Ca}^{2+} \); (2) binding/unbinding of the second \( \text{Ca}^{2+} \).

The \( \text{Ca}^{2+} \) diffusivity was obtained from studies, in which it was concluded that given the known kinetics of typical \( \text{Ca}^{2+} \) buffers, the range of free \( \text{Ca}^{2+} \) after it enters a cell can be up to 200 nm. Therefore, due to the nano-scale of our system, the trajectories of \( \text{Ca}^{2+} \) released by TPC2 signaling complexes in the simulations are governed by the measured diffusivity of free \( \text{Ca}^{2+} \) for a displacement comparable to the nanojunction extension \((2.23 \times 10^{-10} \, \text{m}^2/\text{s})\)\(^53,54\). Once \( \text{Ca}^{2+} \) are buffered they acquire the measured diffusivity of the buffers \((8.4 \times 10^{-11} \, \text{m}^2/\text{s})\)\(^55\).

We summarize the quantitative model input data in Table 2.

---

**Figure 5.** A and B, 3D software reproduction of a lysosome closely apposed to a portion of SR, thereby forming an \( \approx 20\)-nm-wide L-SR nanojunction; this rendering was inspired by a series of observations from micrographs as in Figure 2 (grey object: SR, blue object: lysosome). Included are relevant molecules traversing Brownian motion trajectories produced by the model simulations (see symbol legend below the panels). B, enlarged view of the L-SR junctional region, in which we have displayed the volume object (rust-coloured box) used to measure the \([\text{Ca}^{2+}]_\text{NJ} \) transients like the ones reported in Figure 7. Scale bar: 100 nm. The model geometry and code files are available from the corresponding author.

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**Figure 6.** A, open probability \( P_\circ \) of the TPC2-dependent \( \text{Ca}^{2+} \) conductance reproduced from a quadratic fit to the data reported in the literature\(^4\). B, \( \text{Ca}^{2+} \) release rate calculated as explained in the text, based on the \( P_\circ \) in A.
Using MCell as a stochastic particle simulator, we ran a number of simulations to represent the NAADP-mediated Ca\(^{2+}\) release that is supposed to occur at L-SR junctions in PASM experiments. Released Ca\(^{2+}\) is assumed to undergo Brownian motion in the surrounding space, including the L-SR nanospace. SERCA2a placed on the neighbouring SR surface may capture Ca\(^{2+}\) according to our approximation of their known multistate model. To determine how this regenerated cellular environment can shape a Ca\(^{2+}\) transient we “measured” the junctional [Ca\(^{2+}\)] by counting the ions within the L-SR volume at any given time and dividing the number by that volume. The snapshots in Figure 5 are part of the visual output of this phase of the work. The data to determine the simulated Ca\(^{2+}\) transients in the L-SR junctions were collected in a measuring volume placed between the lysosomal and SR membranes in the virtual L-SR junction (rust-coloured box in Figure 5B).

Previous experimental findings about VSMC PM-SR junctions indicate that disruption of nanojunctions can have profound consequences on Ca\(^{2+}\) signaling properties of the cells. For example, when calyculin A was used to separate superficial SR portions from the plasmalemma of the rabbit inferior vena cava, it was observed that [Ca\(^{2+}\)] oscillations would cease\(^{26}\). This was later corroborated by a quantitative model of the PM-SR junction’s role in the refilling of SR Ca\(^{2+}\) during oscillations\(^{39}\). In another study by the van Breemen laboratory, it was observed that the mitochondria-SR junctions of airway SMC displayed a variable average width as a function of the state of rest or activation of the cell\(^{46}\). It makes sense then to study the effect of changes in the L-SR junctional geometry on the transients generated by our simulations. Therefore, we ran several simulations, in which the separation between the lysosomal and SR membranes was increased from 10 nm to 100 nm in steps of 10 nm. In Figure 7A, we report three sample transients from this set of simulations obtained using three different junctional membrane separations, as indicated in the inset legend. The value of each of the points graphed in this plot is the average value of 100 simulations, in each of which the random number generator within MCell is initiated with a different seed (see Materials and methods). We report representative error bars as 3x the standard error, to convey the > 99% confidence interval of the data. As one would expect, the transient nonjunctional (NJ) Ca\(^{2+}\) concentration, [Ca\(^{2+}\)]\(_{\text{NJ}}\), decreases in magnitude, as the junctional L-SR membrane-membrane separation increases, simply because the released Ca\(^{2+}\) has a larger junctional volume available over which to spread. However, it is important to make a quantitative comparison between this change in [Ca\(^{2+}\)]\(_{\text{NJ}}\), and the [Ca\(^{2+}\)] requirements to activate the putative RyR3 population of the junctional SR. This can be attained by analyzing the time scale of both the recorded and simulated Ca\(^{2+}\) signals.

Reconciling the temporal scales in simulation and experimental results

In the final step of the development of our model, we analyzed the relationship between the time scale of the [Ca\(^{2+}\)]\(_{\text{NJ}}\), transients resulting from our model (Figure 7) and that of the observed Ca\(^{2+}\) signal shoulder in Figure 1.

Let us note that the typical duration of the simulated transients, \(t_{\text{transient}}\), is about 0.06 s and recall that these represent \(\Delta[\text{Ca}^{2+}]\) above the resting [Ca\(^{2+}\)]. On other hand, the build up to the maximum value of \(\Delta[\text{Ca}^{2+}]_{\text{shoulder}}\) takes about 5 s (we refer to this time as \(t_{\text{shoulder}}\)). This interval is about two orders of magnitude larger than the duration of the simulated individual transients (Figure 7A). One way to reconcile the hypothesis that L-SR junctions are at the base of the observed NAADP-induced Ca\(^{2+}\) signals such as the ones in Figure 1 and in particular that the signal shoulder emerges from lysosomal Ca\(^{2+}\) release at L-SR junctions, is to bring forward the assumption that the signal shoulder may be the result of a sequential summation effect over many L-SR junctions, each working according to an all-or-none mechanism of Ca\(^{2+}\) release, and that the

<table>
<thead>
<tr>
<th>Table 2. Quantitative model input data.</th>
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<tr>
<td><strong>Quantity</strong></td>
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<tr>
<td>(\Delta[\text{Ca}^{2+}]_{\text{shoulder}})</td>
</tr>
<tr>
<td>[Ca(^{2+})](_{\text{lys}})</td>
</tr>
<tr>
<td>lysosomal average radius</td>
</tr>
<tr>
<td>(cylindrical) SMC radius</td>
</tr>
<tr>
<td>SMC length</td>
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<tr>
<td>SMC cytosolic volume</td>
</tr>
<tr>
<td>TPC2 complexes per lysosome</td>
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<tr>
<td>TPC2 complex Ca(^{2+}) release rate</td>
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<tr>
<td>SERCA2a surface density</td>
</tr>
<tr>
<td>SERCA2a first Ca(^{2+}) binding (unbinding) rate</td>
</tr>
<tr>
<td>SERCA2a second Ca(^{2+}) binding (unbinding) rate</td>
</tr>
<tr>
<td>free Ca(^{2+}) diffusivity, (D_\text{free})</td>
</tr>
<tr>
<td>Ca(^{2+}) buffer diffusivity, (D_\text{buffer})</td>
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Summary of input parameters used in various phases of the quantitative model, and references to the origin of their values.
“firing” of one junction may cause a cascading effect across the set of junctions that produces the shoulder. Then the duration of the shoulder upstroke can be expressed as:

\[ t_{\text{shoulder}} = N_{\text{nj}} \times t_{\text{transient}} \]  \hspace{1cm} (3)

where \( N_{\text{nj}} \) is the number of L-SR nanojunctions that yield \( \Delta [\text{Ca}^{2+}]_{\text{nj}} \). Since we calculated above that \( \text{Ca}^{2+} \) release from \( N_{\text{nj}} \approx 60-100 \) lysosomes would be necessary to produce the observed signal shoulder magnitude and have shown in the previous section that such release would need to take place at L-SR junctions, we can estimate \( t_{\text{transient}} \), the duration of the \( [\text{Ca}^{2+}]_{\text{nj}} \) transient, by reversing Equation (3) and obtaining \( t_{\text{transient}} = 0.05-0.08 \) s. It is noteworthy that this range of values is obtained in a manner completely independent of our simulation results, which yielded a similar range of values.

To gain quantitative insight into the effects of varying the junctional width, we then calculated the temporal average of the \( [\text{Ca}^{2+}]_{\text{nj}} \) over \( t_{\text{transient}} \) (using the middle value of the range calculated via Equation (3)) and graphed it as a function of the junctional width. The result of this analysis is reported in Figure 7B (solid blue circles). As we have anticipated at the end of the previous section, the decrease in magnitude of these data is to be expected; however, in this plot we also indicated the range of \( [\text{Ca}^{2+}] \) values (shaded area) over which maximum SR \( \text{Ca}^{2+} \) release via RyR3 is reported to occur in skeletal muscle. This comparison underscores the important constraint played by the width of the L-SR junctions and indicates that, unless the membrane separation is kept below about 30 nm, it is not possible for \( [\text{Ca}^{2+}]_{\text{nj}} \) to breach the threshold for RyR3 \( \text{Ca}^{2+} \) release. Let us also point out that the \( [\text{Ca}^{2+}]_{\text{nj}} \) data in Figure 7B would shift upward, toward concentration values that would make the junction more prone to promote RyR3 release, if the temporal average were taken over a shorter transient time, \( t_{\text{transient}} < t_{\text{transient}} \) peak. However, in that case Equation (3) indicates that a larger \( N_{\text{nj}} \) (than 80, picked as the middle of the 60–100 range) would have to contribute to the signal summation that results in a 5-second \( t_{\text{shoulder}} \).

Interestingly, this possibility agrees with our lysosome count from TEM images (110 lysosomes/cell) and with the argument of natural redundancy we contemplated to explain the discrepancy between the calculated and observed lysosome number estimates. As an exercise we have recalculated the \( [\text{Ca}^{2+}]_{\text{nj}} \) time-averaged over \( t_{\text{transient}} \) obtained using 110 lysosomes in Equation (3) (empty purple circles in Figure 7B), and this indeed shows that RyR3 \( \text{Ca}^{2+} \) release threshold would be cleared more readily. These observations suggest that activation of RyR3s in the L-SR junctions not only depends on the concentration of \( \text{Ca}^{2+} \) near them, but also on their exposure time to this concentration.

L-SR nanojunction study data

7 Data Files

http://dx.doi.org/10.6084/m9.figshare.988699

Discussion

Inter-organellar nanospaces

We have recently introduced the concept of the “panjunctional SR”, which states that \( \text{Ca}^{2+} \) release and uptake at a family of specific nanojunctions connected by a continuous but variable SR lumen integrates cellular control over multiple functions. The lysosome/endoplasmic reticulum (L-S/ER) junction is the most current junction to be considered in this context and exhibits perhaps the highest degree of plasticity of the family of nanojunctions of the SR; the mechanism and function of lysosomal \( \text{Ca}^{2+} \) signaling is currently hotly debated.

By means of a thorough ultrastructural study in rat pulmonary artery smooth muscle, we have observed and characterized L-SR nano-junctions, which had been previously hypothesized on the basis of optical measurements of \( \text{Ca}^{2+} \) signals and optical immunocytochemistry. Our observations corroborate the previously reported finding that lysosomes in PASMCs tend to cluster in the
perinuclear region, as is evident in our micrographs (e.g., Figure 2A). We find that L-SR junctions are on average 15 nm in width (equivalent to our preliminary reports and to recent observations in cultured fibroblasts) and extend approximately 300 nm in lateral dimensions, thereby involving about 15% of the lysosomal membrane (Table 1).

**Mechanism of NAADP Ca²⁺ signaling**

In an effort to achieve quantitative understanding of the phenomenon of NAADP-mediated Ca²⁺ transients and verify the proposal that these may be generated in L-SR junctions, we focused on one of the prominent features of these Ca²⁺ signals, namely the localized Ca²⁺ bursts that precede the propagating Ca²⁺ wave (Figure 1), which we refer to as the signal shoulder ($[\Delta(Ca^{2+})]_{shoulder}$). In the first instance, we have estimated the potential contribution of local bursts of Ca²⁺ release from individual lysosomes to the elevation of global [Ca²⁺]i observed during this shoulder in the experimental records. From this, and using the dimensions of a typical smooth muscle cell, we calculated that 60–100 lysosomes would be required to cause an elevation of comparable magnitude to the signal shoulder. This is lower than the estimate for the total number of lysosomes per cell, 110, we obtained from our ultrastructural study, but comparable in order of magnitude. We have already mentioned above a number of factors in favour of observing a greater number of lysosomes/cell than the estimated number required to generate the signal shoulder. In addition, several other elements contribute a degree of uncertainty to those estimates, such that their discrepancy may not be as large. We need to consider that published $P_o$ data for the Ca²⁺ conductance associated with the TPC2 signaling complex, on which we based our TPC2 rate table, show some variability according to the standard deviation bars, which, in turn, implies an uncertainty in the interpolated $P_o$ (Figure 5A). Moreover, we cannot exclude the possibility that these data reflect a contribution from multiple channels ($N P_o$) rather than a purely single channel $P_o$. Lastly, the standard deviation of the simulated transient (only the standard error is shown in Figure 7A) and the variability in the experimental determination of the [Ca²⁺] sensitivity of RyR may allow for some uncertainty in the estimated number of nanojunctions.

To take this study further and understand whether the observed L-SR junctions could give rise to [Ca²⁺], transients of appropriate magnitude to trigger Ca²⁺ release from RyR3 channels on the junctional SR, we developed a quantitative stochastic model of Ca²⁺ dynamics in the junctional nanospaces. We have previously published a simplistic version of this model, which nonetheless captured the essential features of the problem and yielded an indication that such Ca²⁺ transients in the L-SR junctions would be possible. However, the simulated transients we obtained displayed unphysiological features, such as an abrupt onset and decay. We show here that this was largely due to lack of a faithful representation of the open probability for the Ca²⁺ conductance of the TPC2 signaling complex. We have now combined experimental information on the biophysical properties of conductance and open probability and on the luminal [Ca²⁺]i of the lysosomes to implement a more realistic Ca²⁺ release rate model in the simulations. As a consequence, we are able to output a physiologically meaningful junctional transient profile (Figure 7A), and observe that the [Ca²⁺]i transients generated within our model junctions—in turn, based on those observed in our TEM images—reach peaks of about 20 µM. While we are still not able to measure these transients in individual L-SR junctions (peri-lysosomal Ca²⁺ probes are only recently becoming available and so far have not been used in smooth muscle cells, vascular or otherwise), it is worth noticing that the values we find are comparable to those recently measured in so-called Ca²⁺ hot spots in the mitochondria-ER junctions of neonatal ventricular cardiomyocytes (rat culture) and in RBL-2H3 and H9c2 cells earlier. These results therefore suggest that the hypothesis presented for the role of L-SR junctions in cellular Ca²⁺ signaling is certainly plausible.

**Lysosomal trigger zone**

Although our simulations were successful, comparison of the Ca²⁺ transients in Figure 1 and Figure 7 not surprisingly reveals a striking difference between the simulated Ca²⁺ kinetics of a single L-SR junction and those of the whole cell. This illustrates important aspects related to the concept of lysosome-SR “trigger zone” previously introduced by one of us (AME) and recently proposed as a facilitating factor in lysosomal-ER signaling in reverse, whereby Ca²⁺ release from ER compartments enables NAADP-mediated activation of acidic organelles.

Briefly, this concept suggests that in PSMCs clusters of lysosomes and closely apposed SR regions containing sets of RyRs may act together to form specialized intracellular compartments that transform NAADP-stimulated localized lysosomal Ca²⁺ release into cell-wide Ca²⁺ signals via RyR-supported CICR. If the firing of one junction were sufficient for subsequent initiation of the CICR across the entire SR, the experimentally observed threshold (Figure 1) would be much lower and the rate of junctional coupling by CICR faster. In other words, due to the high [Ca²⁺]i threshold of about 10 µM for Ca²⁺ activation of RyR3, CICR engendered by a single L-SR junction is likely to die out, unless reinforced by a process of quantal releases by other L-SR junctions within a cluster. Therefore it seems more likely that multiple L-SR junctions work in concert in a process characterized by both additive and regenerative elements to provide the necessary threshold and margin of safety required to ensure the propagation by CICR of the global wave via the more widely distributed RyR2 along extra-junctional SR, once the Ca²⁺ release wave escapes an RyR3-enriched SR region, as previously suggested.

In this respect, our results also allowed us to establish a time interval for the duration of the transient ($t_{shoulder}$) that would be compatible with the hypothesis that sequential summation of all-or-none lysosomal Ca²⁺ release events from a set of individual L-SR junctions is responsible for generating $[\Delta(Ca^{2+})]_{shoulder}$. Remarkably, this value is comparable to the average duration of the simulated transients, which was determined on the basis of ultrastructural details and completely independently of the summation effect hypothesis. The assumption of summation may imply that the role of the NAADP as a stimulus is limited to the initial lysosomal Ca²⁺ release, while the recruitment of subsequent L-SR junctions may be governed by further lysosomal calcium release, by SR Ca²⁺ release via Ca²⁺ activated junctional RyR3, and/or propagation and combination of calcium signals via inter-junctional clusters of RyR3. Therefore, from our model we envision the intriguing possibility of an
Disruption of nanojunctions, plasticity and pathology

Carrying the signal time-scale analysis further and using the calculated $r_{nano}$ to take a temporal average of the simulated $[\text{Ca}^{2+}]_{\text{el}}$ at different junctional widths, we find that above a width of about 30 nm the transients would be unable to trigger Ca$^{2+}$ release from the RyR3s (Figure 7B). Thus it is possible that heterogeneity and plasticity are controlled by a variable width of the junctional nanospace. For example, in atrial myocytes it has been proposed that NAADP evokes Ca$^{2+}$ release from an acidic store, which enhances general SR Ca$^{2+}$ release by increasing SR Ca$^{2+}$ load and activating RyR sites. This functional variant may be provided by either: (1) An increase in junctional distance such that $[\text{Ca}^{2+}]_{\text{el}}$ is insufficient to breach the threshold for activation of RyR2, yet sufficient to allow for increases in luminal Ca$^{2+}$ load of the SR via apposing SERCA2 clusters; or (2) L-SR junctions in cardiac muscle formed between lysosome membranes and closely apposed regions of the SR which possess dense SERCA2 clusters and are devoid of RyR2. Further ultra-structural studies on cardiac muscle and other cell types may therefore provide a greater understanding of how L-SR junctions may have evolved to provide for cell-specific modalities within the calcium signaling machinery.

Under metabolic stresses, such as hypoxia, lysosomal pathways have been proposed to provide for autophagic glycolgen metabolism via acid maltase in support of energy supply, and significant levels of protein breakdown during more prolonged metabolic stress. Moreover, although controversial, in some cell types it has also been suggested that lysosomes may contribute to the energy supply by providing free fatty acids for beta-oxidation by mitochondria. It may be significant, therefore, that TPC2 gating and thus autophagy may be modulated by mTOR. This may well speak to further roles for lysosomal calcium signaling and L-SR junctional plasticity during hypoxic pulmonary vasoconstriction and even during the development of hypoxia pulmonary hypertension, not least because AMP-activated protein kinase has been shown to modulate autophagy through the phosphorylation and inhibition of mTOR.

The process of autophagy serves not only to regulate programmed cell death, but also to recycle organelles, such as mitochondria, through a process of degradation involving lysosomal hydrolases. For this to occur L-SR junctions would likely be disrupted and thus select for local rather than global Ca$^{2+}$ signals in order to facilitate fusion events among lysosomes, endosomes, autophagosomes and amphisomes, which follows from the fact that Ca$^{2+}$ plays a pivotal role in vesicle trafficking and fusion. Active non-synchronous movements of TPC2-expressing vesicles have been detected in live-cell imaging experiments using GFP-tagged proteins. Thus, lysosomal Ca$^{2+}$ signaling may modulate plasticity in the manner required at all stages of multiple membrane fusion events that are dependent on Ca$^{2+}$ for the effective formation of the SNARE complexes. In short, transport of proteins between lysosomes, Golgi apparatus, and plasma membrane via lysosomes and endosomes may be coordinated at different stages by both global Ca$^{2+}$ signals and spatially restricted Ca$^{2+}$ release from acidic stores in a manner that is in some way determined by L-SR junctional integrity.

Loss of integrity of L-SR junctions may also contribute to disease, given that lysosomal Ca$^{2+}$ release both depletes luminal Ca$^{2+}$ and causes intraluminal alkalinization. Therefore, disruption of L-SR junctions may modulate the activity of pH-sensitive hydrolytic lysosomal enzymes, such as glucocerebrosidase and acid sphingomyelinase, which exhibit a marked loss of function at pH $> 5.5$ that could lead to accumulation of macromolecules such as glucocerebrosides and sphingomyelin. As mentioned previously, dysfunction of these enzyme systems consequent to L-SR junctional abnormalities could also contribute to pathologies associated with subclasses of lysosome storage disease such as Niemann-Pick disease type CI, Pompe and Gaucher, which may include hepatic portal, or pulmonary hypertension, dysfunctions in cholesterol trafficking and consequent increases in plasma cholesterol levels, vascular lesion formation, atherosclerosis/thrombosis and medial degradation.

Conclusion

We have determined that L-SR junctions, about 15 nm in width and extending to $\sim 300$ nm in lateral dimensions are a regularly occurring feature in rat pulmonary arterial smooth muscle cells, in which L-SR junctions were first proposed. The present study provides a mechanistic basis for the observed NAADP-induced Ca$^{2+}$ signals and strong support for the proposal that these Ca$^{2+}$ signals are generated at L-SR junctions. Even within the variability of our recorded values of L-SR junctional widths and extension, our results suggest that localized $[\text{Ca}^{2+}]_{\text{el}}$ transients due to junctional Ca$^{2+}$ release can without fail reach values required to breach the threshold for CICR from junctional RyR3s. Perhaps most significantly, disruption of the nanojunctions decreases $[\text{Ca}^{2+}]_{\text{el}}$ below the value for CICR via junctional RyR3s. Therefore, consistent with previous studies on the PM-SR membrane, we have established that L-SR junctions are required to allow such signals to be generated and that there is a 30–50 nm limit on junctional width, above which there is loss of junctional integrity and inadequate control of ion movements within the junctional space. This suggests that the observed L-SR junctions in PAMSCs are not only capable of delivering localized Ca$^{2+}$ bursts of the required magnitude, but are also necessary if lysosomes are to fulfill this identified role in Ca$^{2+}$ signaling. In other words, L-SR nanojunctions are a necessary and sufficient condition for generating local Ca$^{2+}$ bursts essential for NAADP-induced Ca$^{2+}$ waves. Most importantly, however, this study demonstrates the importance of junctional architecture on the nanoscale to the capacity for coupling across cytoplasmic nanospaces, tight regulation of ion transport and thus signal transduction. In turn, this highlights the possibility that alterations in the dimensions and architecture of intracellular nanojunctions lead to cell dysfunction and hence disease.
Data availability

figshare: L-SR nanojunction study data, doi: 10.6084/m9.figshare.988699

Author contributions

NF and CvB conceived the electron microscopy work, quantitative model, simulations and quantitative analysis of the results. AME and OAO obtained the background optical microscopy study. NF, OAO, CvB and AME contributed to writing the manuscript. All authors were involved in the revision of the draft manuscript and have agreed to the final content.

Competing interests

No competing interests were disclosed.

References


Grant information

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Acknowledgements

We are very grateful to Garnet Martens and the University of British Columbia Bioimaging Facility for their assistance. We also acknowledge the help of David Walker and Arash Tehrani with TEM image analysis and acquisition. This research has been enabled by the use of computing resources provided by WestGrid and Compute/Calcul Canada (http://www.westgrid.ca, http://www.computecanada.ca).
of mouse cardiac cells points to a critical role of SERCA's affinity for Ca2+. Biophys J. 2011; 100(5): 1252. 


Supplementary material

1 Estimating the number of TPC2 complexes per lysosome

The whole-lysosome conductance was calculated based on the work reported by Schieder and collaborators [1], in which current voltage relationships of TPC2-mediated Ca\(^{2+}\) currents were determined for single lysosomes (see Figure 1 therein [1]). A set of these values, as well as the experimental conditions relevant for this calculation, are reported in Table S1. From these data, we first calculated the electrochemical potentials of K\(^{+}\) (\(E_{c}^{+}\)) and Ca\(^{2+}\) (\(E_{c}^{2+}\)) across the lysosomal membrane using the general expression

\[
E_{c}^{z+} = \frac{RT}{zF} \ln \left( \frac{[C^{z+}]_{\text{out}}}{[C^{z+}]_{\text{in}}} \right)
\]

where \(R\) is the universal gas constant, \(T\) the absolute temperature, \(C^{z+}\) a \(z\)-valent cation, \(F\) is Faraday’s constant. We then obtained the whole-lysosome, \(g_{\text{wl}}\), from Ohm’s law:

\[I = g_{\text{wl}} \left( V_{m} - E_{syt} \right)\]

where \(I\) is the measured current, \(V_{m}\) is the applied membrane voltage, and \(E_{syt} = E_{c}^{+}(P_{K}/P_{\text{ca}})E_{\text{ca}}\) with \(P_{K}/P_{\text{ca}}\) the permeability ratio. The calculated conductance values are listed in Table S1.

If we assume that the whole-lysosome conductance under these conditions is due to a set of TPC2 complexes working in parallel, then \(g_{\text{wl}} = N_{\text{TPC2}} g_{\text{TPC2}}\). \(N_{\text{TPC2}}\) is the number of complexes in that set and \(g_{\text{TPC2}}\) is the single TPC2 determined as 15 pS [2]. We can then find \(N_{\text{TPC2}}\) by dividing the calculated conductance, \(g_{\text{wl}}\), by the single channel conductance, \(g_{\text{TPC2}}\). For our calculation, we picked the whole-lysosome conductance value \(g_{\text{wl}} = 316\) pS corresponding to \(V_{m} = -20\) mV, in accordance to the most recent measurements of lysosomal membrane potential [3]. Hence, our estimate of the number of TPC2 signaling complexes in a representative lysosome is:

\[N_{\text{TPC2}} = \frac{g_{\text{wl}}}{g_{\text{TPC2}}} = \frac{316}{15} = 21\] TPC2 complexes

2 Estimating the cytosolic volume of a vascular smooth muscle cell

We model a typical smooth muscle cell as a long and narrow cylinder 3 \(\mu\)m in radius and 130 \(\mu\)m long [4]. Its volume is then \(V_{\text{SMC}} = \pi(3 \mu m)^2 (130 \mu m) \approx 3.7 \times 10^{-12}\) L. To calculate an upper-limit estimate of the cell cytosolic volume, \(V_{\text{cyt}}\), available to Ca\(^{2+}\) released from lysosomes, we need to account for the space occupied by the nucleus, the sarcoplasmic reticulum (SR), the mitochondria and the set of lysosomes we observe (\(V_{\text{nu}}, V_{\text{SR}}, V_{\text{mito}}, V_{\text{mito}}\) and \(V_{\text{all lys}}, \) respectively), so that

\[V_{\text{cyt}} = V_{\text{SMC}} - V_{\text{nu}} - V_{\text{SR}} - V_{\text{mito}} - V_{\text{all lys}}\]

The nuclear volume, \(V_{\text{nu}}\), is estimated from a sample of 10 cells in one of our lower resolution micrographs, by measuring the minor and major axes of the oval-shaped nuclei, approximating their 3D shape as a prolate spheroid and calculating the volume of the spheroid from the measured values of the axes (Figure 2 and Table 1 in the main article). We calculate it as

\[V_{\text{nu}} = 5 \times 10^{-11}\] L

(about 15% of the cell volume).

To estimate \(V_{\text{mito}}\), we resort once again to our own micrographs, in which we observe of the order of 10 mitochondria per sample section per cell. Using a similar procedure to the one employed to obtain the approximate number of lysosomes in a pulmonary artery smooth muscle cell (described in the main article, section “Generation of the signal’s “shoulder” by lysosomal Ca\(^{2+}\) bursts’’), and pretending that mitochondria are spheres 0.8 \(\mu\)m in radius, we arrive at an estimate of 43 mitochondria/cell. Thus,

\[V_{\text{mito}} = 43 \times \frac{4}{3} \pi (0.8 \mu m)^3 = 10^{-13}\] L

\[V_{\text{all lys}} = V_{\text{nu}} + V_{\text{mito}} + V_{\text{mito}} = 6 \times 10^{-12}\] L

\[V_{\text{cyt}} = V_{\text{SMC}} - V_{\text{nu}} - V_{\text{mito}} - V_{\text{mito}} = 9 \times 10^{-13}\] L

Table S1. Data for whole-lysosome conductance calculation.

<table>
<thead>
<tr>
<th>Experimental conditions</th>
<th>(V_{m}) (mV)</th>
<th>(I) (pA)</th>
<th>(g_{\text{wl}}) (pS)</th>
</tr>
</thead>
<tbody>
<tr>
<td>([\text{Ca}^{2+}]_{i})</td>
<td>60 mM</td>
<td>100</td>
<td>15</td>
</tr>
<tr>
<td>([\text{Ca}^{2+}]_{o})</td>
<td>2 mM</td>
<td>60</td>
<td>6</td>
</tr>
<tr>
<td>([\text{K}^{+}]_{i})</td>
<td>70 mM</td>
<td>20</td>
<td>-7</td>
</tr>
<tr>
<td>([\text{K}^{+}]_{o})</td>
<td>60 mM</td>
<td>-20</td>
<td>-20</td>
</tr>
<tr>
<td>(T)</td>
<td>298.16 K (23°C)</td>
<td>-60</td>
<td>-35</td>
</tr>
<tr>
<td>(P_{\text{ca}}/P_{\text{K}})</td>
<td>1000</td>
<td>-100</td>
<td>-50</td>
</tr>
<tr>
<td>(R)</td>
<td>8.3 J/(mol K)</td>
<td></td>
<td></td>
</tr>
<tr>
<td>(F)</td>
<td>9.65 \times 10^{4} \text{J}/(\text{mol})</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Data on experimental conditions by Schieder et al. [1], used to calculate whole-lysosome TPC2-mediated Ca\(^{2+}\) conductance, \(g_{\text{wl}}\), as explained in the text. The subscripts “i” and “o” refer to the inside and outside of a lysosome, respectively.
The volume of the SR in vascular smooth muscle has been estimated previously \cite{5} and expressed as a percentage of the cell volume deprived of the nuclear and mitochondrial volume. These findings suggest that

\[
\frac{V_{\text{SR}}}{V_{\text{SMC}} - V_{\text{nuc}} - V_{\text{mito}}} = 0.75 \tag{4}
\]

Which corresponds to

\[
V_{\text{SR}} = 2 \times 10^{-13} \text{ L} \tag{5}
\]

using the values for $V_{\text{SMC}}$, $V_{\text{nuc}}$, and $V_{\text{mito}}$ found above.

$V_{\text{all lys}}$ can be obtained from the value of $V_{\text{lys}}$ we found in the main article multiplied by the 110 lysosomes/cell we already estimated. However, at a value of $\approx 3 \times 10^{-15} \text{ L}$ it is negligible when compared to the other terms.

Finally, putting Equation (5), Equation (3) and Equation (2) into Equation (1), we find

\[
V_{\text{cyt}} = 2.4 \times 10^{-12} \text{ L} \tag{6}
\]

Clearly, the value in (6) is an upper limit, since we haven’t accounted for all of the organellar volumes in our calculations. However, the values above suggest that to a first approximation nucleus, SR, and mitochondria are by and large the main contributors to the difference between the gross cellular volume and the cytosolic volume.

3 Line-and-point plot of NAADP-induced Ca\(^{2+}\) fluorescence trace

The plot in the figure below was obtained from the same data, from which we produced the Ca\(^{2+}\) trace in Figure 1 in the manuscript. We report it again here in line-and-point form to highlight that the sampling frequency was 0.5 Hz. In a separate supplementary spreadsheet file, we also provide the raw data.

References


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NAADP mediated two-phase Ca\(^{2+}\) release responses were observed in smooth muscle cells, including pulmonary artery smooth muscle cells (PASMCs). Lysosomes and sarcoplasmic reticulum (L-SR) junctions had been proposed as the structural bases in facilitating local lysosomal Ca\(^{2+}\) bursts (first phase) followed by whole cell Ca\(^{2+}\) release responses from SR (second phase) through a calcium-induced calcium release (CICR) mechanism. In their studies reported here, Nicola Fameli et al. attempted to define the L-SR junctions in PASMCs with a nano-level resolution using electron microscopy (EM), and proceeded to build a quantitative L-SR junctional model. Unfortunately, all these efforts will add little to our further understanding of these L-SR junctional structures, if not at all, due to the lack of solid experimental EM results in reconstructing L-SR junctions, some dogmatic presumptions in L-SR model build up, the inadequacy of validation of the model and a failure to include information on the current disputes over the identity of NAADP-associated Ca\(^{2+}\) release channels in lysosomes. Some major concerns are as follows:

1. Data are not convincing and lack quality. In defining L-SR junctions, the assurance of lysosome and SR identities is fundamental. Immune-gold staining has been widely applied in EM studies to ascertain different cell organelles as well as to conduct 3D reconstruction of cell structure \(^1\ 2\). However, such a standard practice was excluded in this study as a 'precaution' to prevent compromising lysosome membrane definition. In line with applying empirical judgments for identifying organelles to the EM data analysis, the authors extrapolated the opposing and proximal organelles adjacent to the nucleus as lysosomes and SR, and deemed them to be worth studying (Figure 2). In addition, the authors stated that there were few lysosomes found in the subplasmalemmal region, which implied that L-SR junctions were profoundly located around the nucleus. However, the fura-2 fluorescence images in Figure 1 as well as in the two citations used as basis of this study (10,11)\(^3\ 4\) exclusively demonstrated that the first phase lysosomal Ca\(^{2+}\) release initiated from a region around the cell membrane, suggesting that L-SR junctions should
1. have cytoplasmic locations near the plasma membrane. It is clear that more reliable methods such as immune-gold staining are needed to resolve the contradictions over their own results.

2. The modeling is based on many assumed parameters and lacks reliable validation. To establish the L-SR nanojunctional model, the present study has introduced many assumptions to some critical parameters such as the distance between the lysosome and SR, the size of lysosome, and total number of lysosomes; many of which lack proper judgments. In the abstract section, the authors rationalized the application of EM in investigating L-SR junctions by stating “L-SR junction membrane separation has been estimated to be < 400 nm and thus beyond the resolution of light microscopy”. In defining the L-SR nanojunctions, they declared that lysosomes and SR “must be separated by a distance of approximately 20 nm and be of the order of a few hundred nm in lateral dimensions” by citing PM-SR junctions studies (10, 11)3 4. However, these two citations originated from the light microscope-based studies of L-SR junctions by the same group. Given the resolution limitation, these light microscopy results have no way to give an estimation as accurate as 20 nm. It is therefore confusing to find that these references fail to back up such claims, in my opinion. In reference 39 5, it has been proposed that the extracellular Ca2+ entered the cell through Na+/Ca2+ exchanges, traversed PM-SR junctions to refill SR via Ca2+-ATPase and sustained cell Ca2+ oscillations. For the current L-SR model, this PM-SR modeling may be used as a comparison or reference system leading to the conclusion “that L-SR had a similar 20 nm gap measured as that in PM-SR junctions.” Given that there are no similarities of PM-SR to L-SR junctions, in terms of potency of Ca2+ movement due to Ca2+ gradient across PM and lysosomal membrane, biophysical property of Na+/Ca2+ exchangers and TPC2 channels, and the actions of Ca2+ -ATPase and RyR3 Ca2+ receptors, such a conclusion is not reliable from the modeling. In addition, a major concern is that this model is too simplistic as it excludes many other Ca2+ regulatory mechanisms in both lysosomes and SR, which may influence the accuracy of junction width or prediction in modeling.

3. Some assumptions contradict well-established concepts due to a lack of consideration of lysosome biology. In explaining the shoulder Ca2+ signal (Figure 1), the authors introduced up to four assumptions and calculated that ~60-100 lysosomes were needed to generate this shoulder peak. They verified these lysosome numbers with their TEM images that “can expect a total of about 110 lysosomes/cell”. Considering these two calculations however, it is obviously problematic because almost all lysosomes (100 out of 110 each cell) will need to functionally contribute to the L-SR junctional structure formation. In addition, TEM measurements showed the lysosome dimensions (Table 1) to be major axis of 398 ± 91 nm and minor axis of 325 ± 65 nm. Given the L-SR junction model features “junctonal membrane separation and extension are about 15 nm and 300 nm”, it could be postulated that all the lysosomes that participated in L-SR structure formation also have to align well with the SR in order to produce a 300 nm junctional extension. Therefore, this L-SR model actually describes a scenario whereby in PASMCs the majority of lysosomes act as the components of L-SR junctions and uniformly have impeccable arrangements to oppose the SR at 20 nm apart. However, this idealistic model largely contradicts the well-established concepts that lysosomes are active organelles that dynamically exert their different functions by their movements and other activities.

4. TPCs as lysosomal Ca2+ release channels are questionable. In quantifying the L-SR model, the authors arbitrarily assumed TPC2 as the only lysosomal NAADP-activated Ca2+ channel by totally disregarding recent findings that TPC2 is not a lysosomal NAADP-associated Ca2+ channel, but a Na+ channel6, 7. They also ignored the contribution of other lysosomal channels or transporters
that may contribute to such L-SR junction activity. This prejudice in channel selection inevitably invites doubts on the reliability of this proposed L-SR model. In addition, the authors collectively adopted electrophysiological data from other research groups without taking into consideration any incompatibilities with the model. For example, in obtaining the total number of TPC2 channels on a lysosome, the authors “…dividing the macroscopic whole lysosome conductance, calculated from the current values reported in a recent study on TPC2-mediated Ca\(^{2+}\) current in isolated lysosomes (48)\(^8\), by the single channel conductances(45)\(^9\), thus estimating that a typical lysosomal membrane may contain ~20 TPC2”. Unfortunately, the lysosomes used in the whole lysosome conductance recording were not normal lysosomes, but lysosomes manipulated by overexpression of TPC2 and were undergoing size enlargement. With these changes, the modified lysosomes would have a much higher TPC2 conductance compared with normal lysosomes with a natural biogenesis. It is important to use a TPC2 conductance and other parameters of channels from lysosomes of normal cells without an artificial transgene for such modeling. Otherwise, the modeling prediction will be inaccurate and the relevance of such a model will be vague.

In conclusion; aside from the issue that the current L-SR nanojunction model fails to contribute any new insights to our understanding of the L-SR architecture, my concerns over the reliability of data and modelling parameters are rather serious – this work is therefore ‘Not Approved’ in its current version.

References

Abstract I Publisher Full Text

Competing Interests: No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to state that I do not consider it to be of an acceptable scientific standard, for reasons outlined above.

Author Response 04 Dec 2014

Nicola Fameli, University of British Columbia, Vancouver, Canada

Dr. Fan Zhang we respond to your concerns below, by reporting your comments in quotes first and adding our response below them in bold.

"Unfortunately, all these efforts will add little to our further understanding of these L-SR junctional structures, if not at all, due to the (1) lack of solid experimental EM results in reconstructing L-SR junctions, (2) some dogmatic presumptions in L-SR model build up, (3) the inadequacy of validation of the model and (4) a failure to include information on the current disputes over the identity of NAADP-associated Ca2+ release channels in lysosomes."

We address points (1) to (4) in detail below (numbering ours).

1. "Data are not convincing and lack quality."

We fully disagree with this statement. Our set of data represents the most complete survey of these structures in vascular smooth muscle currently available in the literature. The micrographs are of high quality with very well defined membranes and identifiable organelles.

"In defining L-SR junctions, the assurance of lysosome and SR identities is fundamental. Immune-gold staining has been widely applied in EM studies to ascertain different cell organelles as well as to conduct 3D reconstruction of cell structure. However, such a standard practice was excluded in this study as a 'precaution' to prevent compromising lysosome membrane definition."

Immuno-gold staining was avoided not as a 'precaution', but because, while certainly enabling us to identify the desired organelles, it reduces visualization of their membranes, since the fixing procedure aimed at preserving tissue antigenicity and the staining protocol for label visualization seriously compromise the resolution of membrane structure required for accurate quantitative characterization of the L-SR junctions. The references this reviewer provides only corroborate our point. As explained in detail by first reference, SEM is less appropriate in this kind of study for a number of reasons: 1) One would need to expose lysosomes buried underneath the PM to visualize their surface membrane. This, as the article suggests, is best done by isolating the organelles. Hardly a technique that can serve the purpose of identifying L-SR junctions, whether we use immuno-gold or not; (2) As is evident from Fig 3 of the second reference, therein no membranes were visible in the micrograph with immuno-gold labeling.
"In line with applying empirical judgments for identifying organelles to the EM data analysis, the authors extrapolated the opposing and proximal organelles adjacent to the nucleus as lysosomes and SR, and deemed them to be worth studying (Figure 2)."

Figure 2, as clearly explained in the text, contains some representative examples from our set of micrographs, but even from the subset in Fig. 2 it is evident that we did not only consider regions neighbouring the nucleus in our survey. This reviewer's statement that "the authors extrapolated the opposing and proximal organelles adjacent to the nucleus as lysosomes and SR" mistakenly implies that the authors simply picked any two objects whose membrane were close enough to be deemed a junctional complex between lysosome and SR. This was most certainly not the case, as explained in detail in the article. The authors can add however that, in spite of our own laboratory's proven experience in electron microscopy (see for example references 39, 56, 57, 92, 98 in our article), we consulted with an expert in the field of electron microscopy, Prof. David Walker, and drew his immense (>30 years) experience (including smooth muscle) to our own. We not only sought and received guidance, but were enlightened with respect to common practices regarding the identification of lysosomes and their junctions with SR.

"In addition, the authors stated that there were few lysosomes found in the subplasmalemmal region, which implied that L-SR junctions were profoundly located around the nucleus. However, the fura-2 fluorescence images in Figure 1 as well as in the two citations used as basis of this study (10,11) exclusively demonstrated that the first phase lysosomal Ca2+ release initiated from a region around the cell membrane, suggesting that L-SR junctions should have cytoplasmic locations near the plasma membrane. It is clear that more reliable methods such as immune-gold staining are needed to resolve the contradictions over their own results."

One does not exclude the other for a couple of reasons. First, the typical diameter and length of vascular smooth muscle cells are about 5 μm and 150 μm, respectively, which ensures that even so-called perinuclear lysosomes are never too far from the periphery. This fact alone would break the apparent contradiction raised. Moreover, the definition of cell periphery that one can gather from fluorescence measurements can easily cover more than 500 nm in depth. Therefore, at lower resolution fluorescence signals could appear close to the membrane yet be generated by perinuclear lysosomes.

"2. The modeling is based on many assumed parameters and lacks reliable validation. To establish the L-SR nanojunctional model, the present study has introduced many assumptions to some critical parameters such as the distance between the lysosome and SR, the size of lysosome, and total number of lysosomes; many of which lack proper judgments."

This point raised by the reviewer verges on the ridiculous. None of the stated parameters was assumed, as should be clear upon reading the article. The L-SR distance was measured, not assumed. The size of the lysosomes observed in micrographs was measured, not assumed. The total number of lysosomes was estimated from our observations, not assumed. We find it difficult to see where proper judgment is lacking, at least with respect to our contribution.

"In the abstract section, the authors rationalized the application of EM in investigating L-SR junctions by stating "L-SR junction membrane separation has been estimated to be < 400 nm and thus beyond the resolution of light microscopy". In defining the L-SR nanojunctions, they declared
that lysosomes and SR “must be separated by a distance of approximately 20 nm and be of the order of a few hundred nm in lateral dimensions” by citing PM-SR junctions studies (10, 11). However, these two citations originated from the light microscope-based studies of L-SR junctions by the same group. Given the resolution limitation, these light microscopy results have no way to give an estimation as accurate as 20 nm. It is therefore confusing to find that these references fail to back up such claims, in my opinion.

We thank the reviewer for drawing our attention to an unfortunate (and clearly confusing) typographical error with respect to in-text numbering of citations. Our intention was to refer to Lee CH et al 2002 and Fameli N et al 2007, or references 92 and 39, respectively, in the present version. This typographical error will be rectified in a future version of the manuscript.

“In reference 39 5, it has been proposed that the extracellular Ca2+ entered the cell through Na+/Ca2+ exchanges, traversed PM-SR junctions to refill SR via Ca2+-ATPase and sustained cell Ca2+ oscillations. For the current L-SR model, this PM-SR modeling may be used as a comparison or reference system leading to the conclusion “that L-SR had a similar 20 nm gap measured as that in PM-SR junctions.” Given that there are no similarities of PM-SR to L-SR junctions, in terms of potency of Ca2+ movement due to Ca2+ gradient across PM and lysosomal membrane, biophysical property of Na+/Ca2+ exchangers and TPC2 channels, and the actions of Ca2+-ATPase and RyR3 Ca2+ receptors, such a conclusion is not reliable from the modeling.”

The sentence in quotes (underlined above) does not appear in our article. Nevertheless let us provide the best rebuttal possible under these circumstances, and address the somewhat confusing comments made by this reviewer in terms of considerations of the relationship between junctional architecture and the biophysical properties of channels and transporters within different junctions. We measured the distance of observed junctional widths between lysosomes and the sarcoplasmic reticulum with closely apposed “junctional” membranes. The calculated average for the size of measured junctional width was not significantly different from the averaged size of PM-SR junctions previously published and cited in this manuscript. We draw the reader to this striking similarity, and relate this to the capacity of the “nanojunction” to hold and concentrate released calcium. In this respect the precise behaviour of each junction will of course be determined by the density and biophysical properties of local pumps and channels, much like the behaviours vary between other previously defined nanojunctions.

“In addition, a major concern is that this model is too simplistic as it excludes many other Ca2+ regulatory mechanisms in both lysosomes and SR, which may influence the accuracy of junction width or prediction in modeling.”

It is always best to start with a minimal model and to make the least number of assumptions possible while not compromising the capacity to inform by way of outcomes. This is what the authors delivered. A more complex model would require the addition of a far greater number of assumptions and, in some cases, these would be difficult to justify. Given earlier comments by this reviewer regarding the limitations of assumptions, it is surprising that such minima are not better appreciated, not least because the model in its current form is sufficient to verify a hypothesis based on a substantial body of peer reviewed work (ref. 10, 11 in the article). As the strength of supporting experimental data grows, it would of course be beneficial to the field if the present model were developed
further to incorporate other “regulatory mechanisms” to assess with greater accuracy these and similar signalling mechanisms that lie beyond the reach of current technologies.

"3. Some assumptions contradict well-established concepts due to a lack of consideration of lysosome biology. In explaining the shoulder Ca\textsuperscript{2+} signal (Figure 1), the authors introduced up to four assumptions and calculated that ~60-100 lysosomes were needed to generate this shoulder peak."

In constructing quantitative (computational) models, and particularly with respect to those that are applied to systems beyond the resolution of current technologies, one is necessarily required to make justified assumptions. All assumptions made are both appropriate and plausible, and are justified by reference to the literature.

"They verified these lysosome numbers with their TEM images that “can expect a total of about 110 lysosomes/cell”. Considering these two calculations however, it is obviously problematic because almost all lysosomes (100 out of 110 each cell) will need to functionally contribute to the L-SR junctional structure formation."

This is a biased reading of our findings, in that we suggest that a range of 60 to 100 lysosomes would have to contribute to the shoulder magnitude, under our model’s conditions, not exactly 100.

"In addition, TEM measurements showed the lysosome dimensions (Table 1) to be major axis of 398 ± 91 nm and minor axis of 325 ± 65 nm. Given the L-SR junction model features “junctional membrane separation and extension are about 15 nm and 300 nm”, it could be postulated that all the lysosomes that participated in L-SR structure formation also have to align well with the SR in order to produce a 300 nm junctional extension."

This reviewer’s postulate forcing the lysosomes to “align well” does not emerge naturally from our model. Would this reviewer explain what relationship between the size of the observed lysosomes (major and minor axes) and the portion of the lysosomal surface participating in the L-SR junction suggests that “all the lysosomes that participated in L-SR structure formation also have to align well with the SR in order to produce a 300 nm junctional extension”? This is most unclear from this reviewer’s comments. It is instead clear upon reading the article that nothing in our model nor in our conclusions depends on such postulate.

"Therefore, this L-SR model actually describes a scenario whereby in PASMCs the majority of lysosomes act as the components of L-SR junctions and uniformly have impeccable arrangements to oppose the SR at 20 nm apart."

This last sentence is a conclusion that this reviewer makes and stems directly from this reviewer’s postulate—which I must re-emphasize we do not make—and not from our model.

"However, this idealistic model largely contradicts the well-established concepts that lysosomes are active organelles that dynamically exert their different functions by their movements and other activities."
The contradiction arises from this reviewer postulated interpretation of our findings. We certainly don’t claim anything that goes against the established observations of lysosomal mobility.

“4. TPCs as lysosomal Ca2+ release channels are questionable. In quantifying the L-SR model, the authors arbitrarily assumed TPC2 as the only lysosomal NAADP-activated Ca2+ channel by totally disregarding recent findings that TPC2 is not a lysosomal NAADP-associated Ca2+ channel, but a Na+ channel6, 7. They also ignored the contribution of other lysosomal channels or transporters that may contribute to such L-SR junction activity. This prejudice in channel selection inevitably invites doubts on the reliability of this proposed L-SR model.”

The authors used a minimal model to examine junctional communication at the L-SR nanjunction. The designation of TPC2 was supported by the fact that TPC2: (1) Is primarily targeted to lysosomes; (2) Is required to support NAADP-dependent calcium signalling; (3) Has been shown to be permeable to calcium by at least two previous studies. Controversy in the field is not ignored, as the authors cite articles that provide detailed coverage on disputes relating to TPC2 calcium permeability. That there is conflict in the field does not negate the use of data derived from papers that demonstrate calcium permeability of these channels.

“In addition, the authors collectively adopted electrophysiological data from other research groups without taking into consideration any incompatibilities with the model. For example, in obtaining the total number of TPC2 channels on a lysosome, the authors “…dividing the macroscopic whole lysosome conductance, calculated from the current values reported in a recent study on TPC2-mediated Ca2+ current in isolated lysosomes (48) 8, by the single channel conductances(45) 9, thus estimating that a typical lysosomal membrane may contain ~20 TPC2”. Unfortunately, the lysosomes used in the whole lysosome conductance recording were not normal lysosomes, but lysosomes manipulated by overexpression of TPC2 and were undergoing size enlargement. With these changes, the modified lysosomes would have a much higher TPC2 conductance compared with normal lysosomes with a natural biogenesis. It is important to use a TPC2 conductance and other parameters of channels from lysosomes of normal cells without an artificial transgene for such modeling. Otherwise, the modeling prediction will be inaccurate and the relevance of such a model will be vague.”

The authors used the best evidence available in support of the present model. At no point was it stated that values used were anything more than estimates derived by best practice from cited and peer reviewed research publications by others. Assuming something other than that which can be derived falls in to the realm of unjustifiable assumption. With present information, assuming a higher TPC2 conductance would be just that.

“In conclusion; aside from the issue that the current L-SR nanjunction model fails to contribute any new insights to our understanding of the L-SR architecture, my concerns over the reliability of data and modelling parameters are rather serious – this work is therefore ‘Not Approved’ in its current version.”

We think that, within the limits of the available techniques and knowledge, our work not only contributes "new insights to our understanding of the L-SR architecture", but also a novel approach to the study of nanojunctional signals.
Alexei Tepikin
Department of Cellular and Molecular Physiology, University of Liverpool, Liverpool, UK

This is certainly an interesting study. The characterisation of lysosomal-SR junctions is novel and conceptually important. The data look convincing but I would recommend the authors to further expand the structural part of the study. It would perhaps be useful to further verify that the organelles participating in the formation of the contacts are indeed lysosomes and that the junctions are not simply the result of accidental collisions of the two organelles. Identifying/visualising tethers between the organelles (by analogy with tethers between mitochondria and the ER) would be particularly advantageous.

Modelling part of the paper is also useful. In this respect it would be valuable to clarify the Ca^{2+} buffering capacity of lysosomes (and its possible effect on the outcomes of the model). One could also perhaps consider measuring this important parameter using isolated lysosomes.

References

Competing Interests: No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.
Anthony P Albert  
Pharmacology and Cell Physiology, Biomedical Sciences Research Centre, Division of Biomedical Sciences, St George’s, University of London, London, UK

This is an interesting and thought provoking study which simulates NAADP-dependent Ca$^{2+}$ transients; these are produced by release of Ca$^{2+}$ from lysosome stores that induce Ca$^{2+}$-induced Ca$^{2+}$ release from SR stores via stimulation of RyR3. It is increasingly apparent that answers to many issues concerning cellular Ca$^{2+}$ signalling will be found by recording local Ca$^{2+}$ events – for instance the recently identified Ca$^{2+}$ sparklets representing Ca$^{2+}$ influx through opening of plasma membrane ion channels.

Simulation studies are always difficult to review and evaluate as they must make many assumptions. However, this work is considered and may be a useful model for how other local Ca$^{2+}$ events might be modeled between discrete organelles. Moreover, unlike many simulation manuscripts, I find this one clear and well-written.

Do the authors think that the use of proximity ligation assay techniques might be appropriate to measure local interactions between TPC2 and RyR3? Also, how are lysosomes refilled with Ca$^{2+}$ following NAADP/TPC2 activation – a role for a lysosome store-operated Ca$^{2+}$ influx?

**Competing Interests:** No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.