Neurosharing: large-scale data sets (spike, LFP) recorded from the hippocampal-entorhinal system in behaving rats
[version 2; referees: 4 approved]

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Abstract

Using silicon-based recording electrodes, we recorded neuronal activity of the dorsal hippocampus and dorsomedial entorhinal cortex from behaving rats. The entorhinal neurons were classified as principal neurons and interneurons based on monosynaptic interactions and wave-shapes. The hippocampal neurons were classified as principal neurons and interneurons based on monosynaptic interactions, wave-shapes and burstiness. The data set contains recordings from 7,736 neurons (6,100 classified as principal neurons, 1,132 as interneurons, and 504 cells that did not clearly fit into either category) obtained during 442 recording sessions from 11 rats (a total of 204.5 hours) while they were engaged in one of eight different behaviours/tasks. Both original and processed data (time stamp of spikes, spike waveforms, result of spike sorting and local field potential) are included, along with metadata of behavioural markers. Community-driven data sharing may offer cross-validation of findings, refinement of interpretations and facilitate discoveries.
This article is included in the Data: Use and Reuse collection.

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Introduction
The hippocampus and entorhinal cortex are essential structures for memory and spatial navigation\(^{-4}\). Position-tuned cells (‘place cells’) are present in CA1, CA3 and dentate gyrus regions\(^{5,6}\). Grid cells, head direction cells, and border cells have been described in the dorsomedial entorhinal cortex, and are critical ingredients of navigation systems\(^{6,7,8,9,10,11}\). The temporal coordination across the entorhinal cortex and hippocampus is secured by various oscillations, especially theta, gamma and sharp wave ripples\(^{14,21}\).

We recorded activity of neurons in these brain regions while animals performed various tasks, such as linear track, open maze, T-maze with wheel running delay, plus maze and zigzag maze, as well as recordings during sleep in the home cage. Extensive technical descriptions of the data sets described in this document are available in several published papers\(^{6,21-27}\). Several questions related to memory, navigation, spike time patterns, population coding, neuronal interactions, neuronal classification, replay, sleep homeostasis and oscillations have been studied based on this dataset\(^{6,21-27}\). However, this dataset may provide valuable information if subjected to yet further analyses. Improved spike sorting, neuron classification and more sophisticated analyses may extend and refine the initial conclusions and offer insights that were previously missed. For these reasons we provide both unprocessed (wide band) and processed versions of our data. In our experience, all methods have limitations and must undergo continuous revision.

We believe that community-driven data sharing, cross-validation of data, unified data formats and large collaborative efforts will facilitate discovery and benefit future progress in neuroscience. We are well aware of the potential risks and arguments against data sharing\(^{41}\) but are of the opinion that the benefits of sharing counteract the risks.

Material and methods
Animal surgery
All protocols were approved by the Institutional Animal Care and Use Committee of Rutgers University (protocol No. 90-042), and all experiments were performed at Rutgers University. Before surgery, one to four rats were housed in a single home cage (made of plastic; size L = 45 cm, W = 23.5 cm, H = 20 cm). Wood shavings were used as bedding and dry pellets were provided as food. The animals were housed in a temperature controlled (68°F), environment under 12:12-hours light:dark cycle where light cycle was from 7AM to 7PM. After surgery, the rats were housed individually, and highly absorbent paper (Techboard, Shepherd Specialty Papers) was used as bedding, and the animal’s health was assessed daily by the experimenters.

Details of surgery and recovery procedures have been previously described in detail\(^{35,46}\). Eleven Long Evans rats (male, 3–8 months old, 250–400 g) were deeply anesthetized with isoflurane (1–1.5%). In two rats (f01_m and g01_m), two silicon probes were implanted (one in each hemisphere) and targeted CA1 region. In three rats (gor01, pin01 and vvp01), two probes (32- and/or 64-site silicon probes) were implanted in the left dorsal hippocampus, targeted to CA1 and CA3 separately, and advanced over sessions and days through overlying neocortical and hippocampal tissue. The probe positions were: rat pin01: CA3: a plane formed by probe shanks was at a 35° angle to coronal plane approximately along the septo-temporal axis, probe shanks were centered on 2.8 mm posterior and 2.6 mm lateral to bregma. CA1: a plane formed by probe shanks was at a 26.5° angle to vertical, at a 35° angle to coronal approximately along the septo-temporal axis, probe shanks were centered on 4.6 mm posterior and 2.4 mm lateral to bregma; rat vvp01: CA3: a plane formed by probe shanks was at a 26.5° angle to coronal plane, probe shanks were centered on 2.8 mm posterior and 2.6 mm lateral to bregma. CA1: a plane formed by probe shanks was at a 26.5° angle to vertical, parallel to sagittal plane, probe shanks were centered on 4.4 mm posterior and 2.3 mm lateral to bregma; rat gor01: CA3: a plane formed by probe shanks was at a 26.5° angle to coronal plane, probe shanks were centered on 3.1 mm posterior, and 3.0 mm lateral to bregma. CA1: a plane formed by probe shanks was at a 26.5° angle to vertical, at a 45° angle to coronal plane, probe shanks were centered on 4.9 mm posterior and 1.5 mm lateral to bregma. In four rats (ec013, ec014, ec016 and i01_m), 32- or 64-site silicon probe(s) were implanted in the right dorsal hippocampus and recorded from CA1, CA3 or dentate gyrus, and another 4-shank silicon probe was implanted in the right dorsocaudal medial entorhinal cortex. In one rat (ec012), one 4-shank silicon probe was implanted in the right dorsocaudal medial entorhinal cortex. In rat ec012, ec013, ec014, and ec016, the probe targeting the entorhinal cortex was positioned such that the different shanks recorded from different layers\(^{21}\) (4.5 mm lateral from the midline; 0.1 mm anterior to the edge of the transverse sinus at a 20–25° angle in the sagittal plane with the tip pointing toward the anterior direction). In rat i01_m, the EC probe had 4 shanks and was positioned such that all shanks recorded from the same layer. For the hippocampus probe in rats ec013, ec014 and ec016, the shanks were aligned parallel to the septo-temporal axis of the hippocampus (45° parasagittal), positioned centrally at 3.5 mm posterior from bregma and 2.5 mm lateral from the midline. During recordings, CA1 electrodes could be recognized by characteristic ripples in the pyramidal layer, whereas uniform ripples were not detected on the DG or CA3 electrodes.

For all silicon probes used, each shank had eight recording sites (160 µm² each site, 1–3-MΩ impedance), and intershank distance was 200 µm. Recordings sites were staggered to provide a two-dimensional arrangement (20 µm vertical separation)\(^{37,48}\). The individual silicon probes were attached to respective microdrives and moved independently and slowly to the target. Two stainless steel screws inserted above the cerebellum were used as indifferent (reference) and ground electrodes during recordings. At the end of the physiological recordings during the behavioural tasks, a small anodal DC current (2–5 µA, 10 s) was applied to recording sites 1 or...
2 days before rats were deeply anesthetized and euthanized by perfusion with 10% formalin solution. The positions of the electrodes were confirmed histologically and reported previously in detail\textsuperscript{21,24}. Histological pictures of rat ec012, ec013, ec014 ec016 are reported in the original publications, using the same animal identifiers\textsuperscript{21}. Histological analysis of rat pin01, vvp01 and gor01 was also reported previously using animal identifier rat1, 2 and 3 respectively\textsuperscript{24}. Histological verification of rats f01_m, g01_m, i01_m and j01_m is not available, but the physiological characterization of the recording sites provide a reliable indication of their locations.

**Behavioural testing**

After the animals recovered from surgery (1 to 2 weeks), physiological signals were recorded during eight different types of behaviours mostly during light cycles (see Table 1).

(1) On an elevated linear track (250 cm × 7 cm), the animals were required to run back and forth to obtain water reward at both ends\textsuperscript{21}. In three animals (gor01, pin01, and vvp01), a similar elevated track was used (170 cm × 6.2 cm, with 22 cm × 22 cm end platforms) that was shortened to 79 or 125 cm in some trials\textsuperscript{23,24}. The same linear track was used in linearOne and linearTwo but at different locations in the same recording room. The center of the track was at the same position for linearOne and linearTwo, but the track was at fixed at a 74° angles from each other, corresponding to the diagonals of the 480 × 640 pixel camera.

(2) In the open field task, the rats chased randomly dispersed drops of water or pieces of Froot Loops (25 mg; Kellogg’s) on an elevated open platform\textsuperscript{21} (180 cm × 180 cm, 120 cm × 120 cm or 100 cm × 200 cm).

(3) In the rewarded wheel-running task, a wheel (diameter = 29 cm) was attached to a rectangular-shape box (39 cm × 39 cm × 39 cm). The rat was required to run in the wheel continuously for 10 seconds, after which time a piece of Froot Loop was dropped in the box as reinforcement\textsuperscript{21}.

(4) In the alternation task in the T-maze (100 cm × 120 cm) with wheel running delay, the animal was required to run on a wheel attached to the waiting area for 10 sec, after which time the animal had access to the central arm of the T-maze, at the end of which the animal chose to turn right or left. The animal was rewarded with water if the choice was opposite to the previous one\textsuperscript{6}.

(5) The elevated plus maze (100 cm × 100 cm) consisted of two open and two closed arms, and each arm had a water cup at the outer end of the corridor. Before the session started, all the four cups were filled with water (~20ul). Once the animal consumed water in all of the water cups, water was given in

<table>
<thead>
<tr>
<th>Behaviour</th>
<th>Behaviour subclass</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>elevated linear track</td>
<td>linear</td>
<td>Linear track, 250 cm × 7 cm.</td>
</tr>
<tr>
<td>elevated linear track</td>
<td>linearOne</td>
<td>Linear track (170 cm × 6.2 cm, with 22 cm × 22 cm end platforms) that was shortened to 79 or 125 cm in some trials\textsuperscript{23,24} (Usually shortened but sometimes also lengthened). The same linear track was used in linearOne and linearTwo but at different locations in the same recording room. The center of the track was at the same position for linearOne and linearTwo, but the track was at fixed at a 74° angles from each other, corresponding to the diagonals of the 480 × 640 pixel camera.</td>
</tr>
<tr>
<td>elevated linear track</td>
<td>linearTwo</td>
<td>Exactly the same as linearOne but the linear track was at different locations in the same recording room. See linearOne.</td>
</tr>
<tr>
<td>open field</td>
<td>bigSquare</td>
<td>180 cm × 180 cm.</td>
</tr>
<tr>
<td>open field</td>
<td>bigSquarePlus</td>
<td>180 cm × 180 cm square open field, divided by plus shaped walls put in the center of the field.</td>
</tr>
<tr>
<td>open field</td>
<td>midSquare</td>
<td>120 cm × 120 cm.</td>
</tr>
<tr>
<td>open field</td>
<td>Open</td>
<td>100 cm × 200 cm.</td>
</tr>
<tr>
<td>rewarded wheel-running task</td>
<td>wheel</td>
<td>Operant wheel running task, See Mizuseki et al., 2009\textsuperscript{21}.</td>
</tr>
<tr>
<td>alternation task in T-maze</td>
<td>Mwheel</td>
<td>Alternation task in T-maze (100 cm × 120 cm) with wheel running delay. See Pastalkova et al., 2008\textsuperscript{6}.</td>
</tr>
<tr>
<td>alternation task in T-maze</td>
<td>Tmaze</td>
<td>Alternation task in T-maze, the same as Mwheel but without delay period. There were 2.78 camera pixels/cm, which converts to 22.24 units/cm for the .whl files (8x compression of pixels).</td>
</tr>
<tr>
<td>elevated plus maze</td>
<td>plus</td>
<td>Plus maze. 100 cm × 100 cm.</td>
</tr>
<tr>
<td>zigzag maze</td>
<td>Zigzag</td>
<td>100 cm × 200 cm zigzag maze. See Royer et al., 2010\textsuperscript{49}.</td>
</tr>
<tr>
<td>wheel-running in home cage</td>
<td>wheel_home</td>
<td>Wheel running in home cage with free access to a wheel with no reinforcement.</td>
</tr>
<tr>
<td>sleep</td>
<td>sleep</td>
<td>Sleeping in home cage.</td>
</tr>
</tbody>
</table>
two water cups either in the closed or open arms depending on the last visited arm. If the animal consumed the water in the closed arm lastly, water was provided in water cups in the open arms (and vice versa) and this regime continued to encourage the rat to visit all arms with similar probability.

(6) In the zigzag maze (100 cm × 200 cm) with 11 corridors, the animals learned to run back and forth between two water wells; 100 µl of water was delivered at each well\textsuperscript{21,22,23,45}.

(7) In the wheel-running in home cage, a wheel (diameter = 29 cm) was attached to a rectangular-shape box (39 cm × 39 cm × 39 cm) which was used as a home cage during the experiment. Rats had free access to the wheel, and ran on the wheel with no reinforcement.

(8) In the sleeping session, the rat slept in the home cage.

For recording of behaviour (1) to (6), animals were water-scheduled for 23 hours prior to the experiment. Otherwise, both dry food and water were provided ad libitum. For tracking the position of the animals, two small light-emitting diodes, mounted above the headstage, were recorded by a digital video camera (SONY) at 30 Hz resolution.

Data collection and cell-type classification

Detailed information about the recording system and spike sorting has been previously described\textsuperscript{21,22,45}. Briefly, signals were amplified (1,000×), bandpass-filtered (1 Hz–5 kHz) and acquired continuously at 20 kHz (DataMax system; RC Electronics) or 32,552 Hz (NeuraLynx, MT) at 16-bit resolution. After recording, the signals were down-sampled to 1,250 Hz (DataMax system) or 1,252 Hz (NeuraLynx, MT) at 16-bit resolution.

In electrophysiological recordings, positive polarity is from zero toward positive values. To maximize the detection of very slowly discharging (‘silent’) neurons\textsuperscript{22}, clustering was performed on concatenated files of several behavioural and sleep sessions recorded at the same electrode position on the same recording day\textsuperscript{22,25,27}. We made extensive use of publicly available analytical and display programs, which were developed in our laboratory (KlustaKwik\textsuperscript{51}, available at http://sourceforge.net/projects/klustakwik/, Neuroscope\textsuperscript{52}, available at http://neuroscope.sourceforge.net/, Klusters\textsuperscript{53} available at http://klusters.sourceforge.net/, NDmanager\textsuperscript{52} available at http://ndmanager.sourceforge.net/). The latest available version at the time was used in each case. Spike sorting was performed automatically, using KlustaKwik\textsuperscript{51}, followed by manual adjustment of the clusters, with the help of autocorrelogram, cross-correlogram and spike wave-shape similarity matrix (Klusters software package\textsuperscript{52}). Because none of the existing spike sorting algorithms is completely automated, manual adjustment is necessary\textsuperscript{51}. This inevitably leads to some operator-dependent variability\textsuperscript{51}; therefore, provided clusters are not always identical to those used in our previous publications. Hippocampal principal cells and interneurons were separated based on their burstiness, waveforms and short-term monosynaptic interactions\textsuperscript{5,17,21,24,45}. Classification of principal neurons and interneurons of entorhinal cortical neurons was based on waveforms and short-term monosynaptic interactions, and described previously in detail\textsuperscript{21}. A total of 3,113 (CA1), 882 (CA3), 66 (DG), 491 (EC2), 568 (EC3) and 551 (EC5) principal neurons and 420 (CA1), 198 (CA3), 52 (DG), 85 (EC2), 215 (EC3) and 91 (EC5) interneurons were identified and included in this data set (see Table 2–Table 4).

To quantify the quality of spike sorting, for each neuron, we calculated an isolation distance\textsuperscript{55}, an interspike interval index \textit{R}\textsubscript{200}\textsuperscript{56} and a fraction of interspike intervals less than 2 msec\textsuperscript{56} using the concatenated files described above. This information is provided in metadata tables included with the data set so that the user can identify sessions containing the desired cells based on these isolation

Table 2. Number of cells recorded. Top row: animal identifier. Left column: brain region. Brain region EC4 indicates either entorhinal cortex layer 3 or 5 (could not be determined which); region EC7 indicates in entorhinal cortex, but without layer assignment.

<table>
<thead>
<tr>
<th>Brain region</th>
<th>ec012</th>
<th>ec013</th>
<th>ec014</th>
<th>ec016</th>
<th>f01_m</th>
<th>g01_m</th>
<th>gor01</th>
<th>l01_m</th>
<th>j01_m</th>
<th>pin01</th>
<th>vvp01</th>
<th>total</th>
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<tbody>
<tr>
<td>EC2</td>
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<td>180</td>
<td>112</td>
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<tr>
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<td>439</td>
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<td>2622</td>
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<tr>
<td>CA1</td>
<td>1185</td>
<td>1136</td>
<td>661</td>
<td>99</td>
<td>145</td>
<td>50</td>
<td>309</td>
<td>23</td>
<td>116</td>
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<td>1561</td>
<td>1840</td>
<td>99</td>
<td>145</td>
<td>203</td>
<td>394</td>
<td>90</td>
<td>68</td>
<td></td>
<td>7736</td>
</tr>
</tbody>
</table>
quality measurements. An initial quality control filtering was done in selecting cells to include in the data set. Specifically, there were 7943 cells that were detected, but of those, only 7736 were included in the data set. 207 were not included because they were judged to have insufficient quality.

Our classification of principal neurons and interneurons is supported by short-term spike cross-correlogram and is consistent with previous reports\(^5^7\)–\(^5^9\). However, mis-classification of pyramidal cells and interneurons is inevitable to some extent by our methods. For example, certain types of interneurons with wider wave shapes (e.g., subgroups of somatostatin positive interneurons) might have been falsely classified as pyramidal cells by our method\(^6^0\). Examining the bursting properties of the putative pyramidal cells may help identifying these neurons. The cell-type classification method should be verified and refined by optogenetical tools in the future\(^6^0\)–\(^6^5\).

The tip of the probe either moved spontaneously relative to the brain or was moved by the experimenter between recording days to record from potentially different sets of neurons. However, we cannot exclude the possibility that some neurons recorded on different days were identical, because spikes recorded on each day were clustered separately, though in some instances neurons were recorded over multiple days. When we moved the electrodes, we waited for at least an hour before recording in order to stabilize the position of electrodes.
Data description

The data are available at CRCNS.org (http://dx.doi.org/10.6080/K09G5JRZ). Details of the data collection, processing and storage of data into files are included with the data set, including scripts useful for processing the data. Here, we briefly summarize the data description.

The number of cells recorded from each animal and brain region is shown in Table 2.

Most of the recorded cells were classified as principal neurons or interneurons. The number of cells classified as principal and interneuron are shown in Table 3 and Table 4.

The 8 types of behaviours (see Behavioural Testing section) were further subdivided into 14 behaviour subclasses based on minor differences (e.g. size of maze) and used as behaviour identifiers in the dataset (Table 1).

The data were obtained during 442 recording sessions. During each session the animal performed one of the 14 behaviour subclasses. The number of recording sessions and behaviour subclasses used with each animal is shown in Table 5. The description of each behaviour subclass is given in Table 1.

The 442 sessions with neural responses included in the data set have a total duration of 204.5 hours. They are a subset of 1,538 recording sessions that were used for the spike sorting. Most of the sessions for which neural responses are not included in the data set have behaviour "sleep".

Data file organization

The data files for each recording session are stored in separate compressed tar archive files (i.e. with extension “.tar.gz”). These files are organized into top-level directories, each of which contains data for sessions recorded on the same day using the same animal and electrode placement combination. Data from all sessions recorded from the same animal on the same day were merged for spike sorting. All merged sessions are stored in the same top-level directory in the data set at CRCNS.org. Therefore, the cell identification numbers assigned by the spike sorting are common to all sessions within a top-level directory, and are not specific to individual sessions. Details of the file organization are provided in the document “CRCNS.org hc3 data description” which is included with the data set.

Metadata organization

The metadata describing the data is stored in six tables that are included with the data set. Table cell has information about each spike sorted cell. Table session has information about the 442 experimental sessions that have data files included in the data set. Table session_a has information about all sessions (1,538) that were used to do spike sorting. Table spike_count has the number of spikes recorded from each cell in each of the 1,538 sessions. Table epos contains information about the position of the electrodes. And table file has information about the “.tar.gz” and video files that are included the data set.

These tables are provided in CSV (comma-separated values) format, Excel format, and as tables in an SQLite database. SQLite (http://www.sqlite.org/) is a free, open source, SQL data base engine.

<table>
<thead>
<tr>
<th>Behaviour subclass</th>
<th>ec012</th>
<th>ec013</th>
<th>ec014</th>
<th>ec016</th>
<th>f01_m</th>
<th>g01_m</th>
<th>gor01</th>
<th>l01_m</th>
<th>j01_m</th>
<th>pin01</th>
<th>vvp01</th>
<th>Total</th>
</tr>
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<tbody>
<tr>
<td>bigSquare</td>
<td>24</td>
<td>45</td>
<td>4</td>
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<td>1</td>
<td>4</td>
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<td>8</td>
<td>91</td>
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available for all common operating systems. Tables `cell`, `session`, `session_a`, `epos` and `file` are related to each other through the common field "topdir" which has the name of top-level directories described above. Table `spike_count` is related to tables `cell`, `session`, and `session_a` by the id field in those tables. The table fields are given in Listing 1 (except for `session_a` which is similar to `session`).

Document “crcns-hc3-data-description” has examples showing how the SQLite command interface can be used with these tables to generate summary statistics from the metadata and to identify data files of interest (for example, find sessions with a maximum number of simultaneously recorded cells from a particular brain region that have a desired isolation characteristics) prior to downloading the data. Sessions of interest can also be identified using file “crcns-hc3-cell-counts.zip” which has tables giving the number of cells recorded from each region in each session (for both all cells and well isolated units) and also the sessions with the largest number of recorded cells from each region (for both all cells and well isolated).

**Listing 1: Fields in metadata tables.** Fields for each table are documented in the comments.

```sql
create table cell
  id integer,       -- Id used to match original row number in MatLab PyrIntMap.Map matrix
topdir string,    -- top level directory containing data
animal string,    -- name of animal
ele integer,      -- electrode number
clu integer,      -- ID # in cluster files
region string,    -- brain region
nexciting integer,   -- number of cells this cell monosynaptically excited
ninhibiting integer, -- number of cells this cell monosynaptically inhibited
exciting integer,    -- physiologically identified exciting cells based on CCG analysis
inhibiting integer,  -- physiologically identified inhibiting cells based on CCG analysis
  -- (Detailed method in Mizuseki Sirota Pastalkova and Buzsaki., 2009 Neuron paper.)
excited integer,     -- based on cross-correlogram analysis, the cell is monosynaptically excited by other cells
inhibited integer,   -- based on cross-correlogram analysis, the cell is monosynaptically inhibited by other cells
fireRate real,  -- meanISI=mean(bootstrap(100, 'mean',ISI)); fireRate = SampleRate/MeanISI; ISI is interspike intervals.
totalFireRate real,  -- num of spikes divided by total recording length for a period during which the neuron was stably recorded.
cellType string      -- 'p'=pyramidal, 'i'=interneuron, 'n'=not assigned as pyramidal or interneuron
eDist float,      -- "isolation distance" (see Harris, Hirase, Leinekugel, Henze and Buzsaki, Neuron, 2001^1^)
RefracRatio float, -- This is an interspike interval index "R2/10" (in Fee, Mitra and Kleinfeld, Journal of Neuroscience Methods, 1996^6^). R2/10 = (fraction of ISI < 2ms)/(fraction of ISI < 10 ms)*9.15/1.15 (our shortest interval between spikes allowed by our spike sorting method is 0.85 ms, (10-0.85)/(2-0.85) = 9.15/1.15)
RefracViol float  -- Fraction of interspike intervals less than 2 msec.
);

create table session (  -- this is actually an SQL view on session_a, see crcns-hc3-data-description
  id integer,     -- matches row in original MatLab Beh matrix
topdir string,  -- directory in data set containing data (tar.gz) files
session string, -- individual session name (corresponds to name of tar.gz file having data)
behavior string, -- one of: Mwheel, Open, Tmaze, Zigzag, bigSquare, bigSquarePlus,
  -- linear, linearOne, linearTwo, midSquare, plus, sleep, wheel, wheel_home
familiarity integer, -- number of times animal has done task, 1=animal did task for first time,
  -- 2=second time, 3=third time, 10=10 or more
duration real   -- recording length in seconds
);

create table file (  -- information about files in hc3 dataset
  topdir string,  -- directory in data set containing data (tar.gz) files
session string, -- individual session name (corresponds to name of tar.gz file having data)
size integer,   -- number of bytes in tar.gz file
...
create table epos (  
   -- has electrode positions for each top level directory  
   -- Note, some regions do not match that in cell table.  
   -- Those that differ have following meanings:  
   --   DGCA3: not sure if the electrode is DG or CA3.  
   --   Ctx: somewhere in the cortex (above the hippocampus)  
   --   CA: somewhere in the hippocampus (do not know if it is CA1, CA3 or DG)  
   topdir string,  -- directory in data set containing data (tar.gz) file  
   animal string,  -- animal name  
   e1 string,      -- region for electrode 1  
   e2 string,      -- region for electrode 2  
   -- ... (e3 through e15 fields not shown)  
   e16 string      -- region for electrode 16  
);

create table spike_count (  
   -- contains number of spikes each cell has in each session (if cell could have fired).  
   cellId integer,   -- id in cell table (row in original MatLab PyrIntMap.Map)  
   sessId integer,   -- id in session table (row in original MatLab Beh table)  
   nSpikes integer   -- number of spikes for cell in the session  
);

Data availability
CRCNS: Multiple single unit recordings from different rat hippocampal and entorhinal regions while the animals were performing multiple behavioral tasks, http://dx.doi.org/10.6080/K09G5JRZ

Terms of data usage: Data on this site is made available only for scientific purposes. Redistribution of the data is not permitted (except to direct collaborators working with you on the data or to students in your classroom). Any publications derived from the data must state the data contributors and CRCNS.org as being the source of the data and cite the original paper(s) in which the data contributors have first described the data. Unnecessary downloading of large data files is not permitted. (To minimize demands on the server, only data expected to be useful for your research or teaching should be downloaded).

Privacy notice: Occasionally the researchers who contribute data wish to know who has downloaded their data. Upon request we will provide this information to the data contributors. So, if you download data, there is a possibility that your name and email address will be provided to the data contributor. We request that the data contributors only use the information for legitimate scientific purposes (such as determining the frequency of downloads, or contacting users to providing updated information about the data or to explore possible collaborations).

Author contributions
KM, KD, EP and GB designed the experiments. KM, KD and EP carried out experiments and collected the data. KM collected data from rats ec012, ec013, ec014 and ec016. KD collected data from rats gor01, pin01 and vvp01. EP collected data from rats f01_m, g01_m, i01_m and j01_m. KM carried out all spike sorting and classification of cell types in this dataset. JT prepared documentations for public data release (data sets hc-2 and hc-3) at CRCNS.org. AS prepared an earlier version of documentations for data set hc-2 at CRCNS.org. KM, JT and GB wrote the paper. All authors were involved in the revision of the draft manuscript and have agreed to the final content.

Competing interests
No competing interests were disclosed.

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The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

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Open Peer Review

Current Referee Status: ✔ ✔ ✔ ✔ ✔

Version 2

Referee Report 29 July 2014
doi:10.5256/f1000research.4461.r5500

Yoshikazu Isomura
Brain Science Institute, Tamagawa University, Tokyo, Japan

I have no concerns on this paper. This is an excellent work that will be useful for many other researchers. I appreciate it.

Competing Interests: No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Version 1

Referee Report 07 May 2014
doi:10.5256/f1000research.4171.r4603

James Knierim
Zanvyl Krieger Mind/Brain Institute and Department of Neuroscience, Johns Hopkins University, Baltimore, MD, USA

This data set will be a valuable resource for investigators who wish to test hypotheses about hippocampal function and interaction with entorhinal cortex at the level of single unit and LFP physiology. I have not investigated the data base carefully to ensure its utility, as I assume the authors have done so. My comments are limited to a few questions about their F1000Research article describing the data base.

1. ‘Animal surgery’ - second paragraph:

Do the authors mean “in the coronal plane?” It is not clear whether the tetrodes were angled medially or laterally in that plane, or whether the authors mean that the tetrodes were angled anteriorly or posteriorly to the coronal plane. Please clarify here and in other locations in the text how the tetrodes were angled.

2. ‘Animal surgery’ - final paragraph:
Does the database contain histological figures? If not, are they easily identifiable and accessible from published reports? It would be very useful to ensure that the precise location of tetrodes could be made available to investigators.

3. Table 1, *elevated linear track*:

Shouldn't the fixed angles be 74 degrees (i.e. +/- 37 degrees)?

4. *Behavioural testing* item (5):

Were the rats motivated to run the corridors in any particular order? Was there any working memory component to the trials?

**Competing Interests:** No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

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**Reader Comment 14 Jul 2014**

Kenji Mizuseki, Osaka City University Graduate School of Medicine, Japan

1. *Do the authors mean "in the coronal plane?" It is not clear whether the tetrodes were angled medially or laterally in that plane, or whether the authors mean that the tetrodes were angled anteriorly or posteriorly to the coronal plane. Please clarify here and in other locations in the text how the tetrodes were angled."

   Indeed the description of probe implantation in the previous manuscript was not detailed enough. What we meant by 'to the coronal plane' was 'a plane formed by silicone probe shanks was 35 degrees to the coronal plane, roughly along the septo-temporal axis'. We provide accurate and complete descriptions of probe implantation in the revised manuscript.

2. *Does the database contain histological figures? If not, are they easily identifiable and accessible from published reports? It would be very useful to ensure that the precise location of tetrodes could be made available to investigators."

   The data set does not contain histological figures, but all available histological figures are reported in the previous publications. In the revised manuscript, we described the relationship between the animal identifiers in this paper and the animal identifiers in the previous publications so that one can easily identify the histology of the animal.

3. *Shouldn't the fixed angles be 74 degrees (i.e. +/- 37 degrees)?"

   Yes, indeed this should be 74 degrees and we fixed it.

4. *Were the rats motivated to run the corridors in any particular order? Was there any working memory component to the trials?"

   The plus maze had two open and two closed arms, and each arm had a water cup at the outer end of it. Before the session started, all the four water cups were filled with water.
Once the animal consumed water in all of the water cups, water was given in two water cups in closed or open arms depending on which type of arms the animal visited and consumed the water lastly, that is, if the animal consumed the water in the closed arm lastly water was provided in water cups in the open arms (and vice versa). Afterword, when the animal consumed the water in the two water cups, water was given in the two water cups in the opposite type of arms to encourage the animal to visit all the wells with similar probability. We added this information in the revised manuscript.

**Competing Interests:** No competing interests were disclosed.
make up a small battalion. Mizuseki and colleagues invite critics to their doorstep. Perhaps this is more similar to posing nude as a model for artist. By placing data online, it comes with judgment and the potential to be proved wrong. What do the authors believe the convention is if others follow in their footsteps? For example, if group A shares their database after publishing their results, group B downloads and analyzes their data in new light and finds the opposite results, what is the appropriate manner in handling the situation? The self-correction aspect of science is also accelerated when data is openly-shared. It remains to be seen how situations like this should/will be handled.

3. There is also a chance for the data to be used in order to stifle or impede publication when the result is dubious. That is, should it be considered “fair-play” for a reviewer to use the same database and conduct a similar analysis with results that contradict a submitted manuscript’s results? A tactic such as this only seems appropriate in “open-review formats”.

4. “A final barrier to sharing digital reconstructions relates to the reluctance to lose or give a competitive edge” (Ascoli, 2006). The release of this immense database will surely be the stronghold of many new assistant professors who are still in the initial stages of setting up a physiology laboratory. Moreover, I can see these data being used in laboratories that are heavily analysis driven and limited in their own capacity for high-density in vivo recordings. This increase in the number of people analyzing data invites competitors for the authors as well as their neutral peers. Sheer logic dictates that the authors are not afraid of competition (otherwise, why share the data? Please note that I am a cynic and have been taught by many reviewers that “scientific altruism” is as abundant as snarks and unicorns). Why should researchers not be afraid of others using this database to compete? Is it believed that these data will be used for collaboration? What can be done to emphasize collaboration across laboratories when using the database? How is authorship handled when multiple groups use these data? If I spend my time analyzing these data, only to have it published under a “group project name” (similar to Sir William Timothy Gowers’ Polymath project), do I put it on my CV?

5. The paper explicitly states that large downloads are prohibited. Does this mean that I should not download all the data? Is it OK to use these data for a class project? If so, is it more appropriate for the professor to disseminate it to the students or should the students make their own CRCNS account? Finally, more of an afterthought but along similar lines: should there be a collaborative processing code library that should be developed and maintained in parallel with the use of these data (similar to GitHub or SourceForge)?

I absolutely do not expect the authors to have complete answers to these questions nor should they carry the sole responsibility of determining the general conventions of what constitutes use versus misuse, but I do think that it is worth hearing their general thoughts. As this is the largest and most comprehensive database of in vivo hippocampal and entorhinal physiology to become available to the general scientific community, there will be an immense ramification. Scientific replication/external validation is an immediate and positive application of this database. As I have cited Timothy Gowers above, I think it would be best to leave off with his opinion of open data-sharing and collaboration: “It feels as though this process is to normal research as driving is to pushing a car.” (http://gowers.wordpress.com/2009/02/01/questions-of-procedure/).

As a field we have the opportunity to compete or collaborate. I hope that these data facilitate cross-laboratory collaboration where two groups are reticent to share their own data. For those that are interested in embracing the collaborative spirit, the CRCNS website also has a “marketplace” section
where ideas and potential collaborations can be discussed. Finally, I applaud the authors for this unprecedented act of scientific altruism. I hope this will be a platform that accelerates our understanding of the entorhinal-hippocampal circuitry through collaboration.

**Competing Interests:** No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

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Reader Comment 15 Jul 2014

Kenji Mizuseki, Osaka City University Graduate School of Medicine, Japan

1. **One barrier to sharing data** is “the fear of being scooped” (Ascoli, 2006). For example, scientific progress will be dramatically increased through parallel (and hopefully, collaborative) data analysis. Are the authors concerned about being “scooped”? What about a group of researchers unknowingly using the database to conduct an analysis that is also a student’s PhD project? This hypothetical student may still be acquiring the expertise to keep pace with more seasoned researchers.

   We absolutely agree that “the fear of being scooped” is a big concern and for this reason some people feel strong resistance against data sharing. On the other hand, this fear is not specific to data sharing: similar experiments and analysis are underway by many groups independently every day with or without knowing that that is also a student or postdoc’s project in other groups. Our tentative feeling is that, if the research group (principal investigator and his/her students and postdocs) is happy to share data publicly, we do not see any reason not to do so. If the group is not willing to share data due to the fear of being scooped, we do not see any reason to force them to share the data.

2. “**An often unspoken resistance to the sharing… data is born out of concern for criticisms and mistakes**” (Ascoli, 2006). This is an unprecedented event that Mizuseki and colleagues have set forth, by providing a comprehensive catalog of data in an unabashed manner. The first point I want to touch upon is the level of raw exposure in releasing data. I have never met a neuroscientist who believes that anyone else could've conducted their analyses better than themselves (save a modest few), but to find those who believe that the authors “didn’t look for the right thing” could make up a small battalion. Mizuseki and colleagues invite critics to their doorstep. Perhaps this is more similar to posing nude as a model for artist. By placing data online, it comes with judgment and the potential to be proved wrong. What do the authors believe the convention is if others follow in their footsteps? For example, if group A shares their database after publishing their results, group B downloads and analyzes their data in new light and finds the opposite results, what is the appropriate manner in handling the situation? The self-correction aspect of science is also accelerated when data is openly-shared. It remains to be seen how situations like this should/will be handled.

   We fully agree that releasing data may be judged as “risky” by many. We have added this information to the text and cite Ascoli (2006). However, we think that this ‘self-correction’ process is a sound one and accelerates the progress in science in the long run. Since the
scientific community has only a limited history of data sharing, good manner of data sharing should be formed in the community through experience.

3. There is also a chance for the data to be used in order to stifle or impede publication when the result is dubious. That is, should it be considered “fair-play” for a reviewer to use the same database and conduct a similar analysis with results that contradict a submitted manuscript’s results? A tactic such as this only seems appropriate in “open-review formats”.

Of course this should be judged case by case, but in general we think the situation the reviewer described is a fair-play and even a sound one. Tactics and politics should not be part of scientists’ vocabulary. We need to assume that all of our colleagues are honest and they assume the same about us.

4. “A final barrier to sharing digital reconstructions relates to the reluctance to lose or give a competitive edge” (Ascoli, 2006). The release of this immense database will surely be the stronghold of many new assistant professors who are still in the initial stages of setting up a physiology laboratory. Moreover, I can see these data being used in laboratories that are heavily analysis driven and limited in their own capacity for high-density in vivo recordings. This increase in the number of people analyzing data invites competitors for the authors as well as their neutral peers. Sheer logic dictates that the authors are not afraid of competition (otherwise, why share the data? Please note that I am a cynic and have been taught by many reviewers that “scientific altruism” is as abundant as snarks and unicorns). Why should researchers not be afraid of others using this database to compete? Is it believed that these data will be used for collaboration? What can be done to emphasize collaboration across laboratories when using the database? How is authorship handled when multiple groups use these data? If I spend my time analyzing these data, only to have it published under a “group project name” (similar to Sir William Timothy Gowers’ Polymath project), do I put it on my CV?

We do not have the answers to all these questions, of course. We can talk only about our opinions. Sharing data has two compelling reasons (1) The experiments were largely performed using public money, therefore it is only natural to return our results (data) to public at some point. (2) We cannot exhaust the every single possible analysis by ourselves, and other researchers may have much better idea and capability to analyze the data set to provide new insights.

5. The paper explicitly states that large downloads are prohibited. Does this mean that I should not download all the data? Is it OK to use these data for a class project? If so, is it more appropriate for the professor to disseminate it to the students or should the students make their own CRCNS account? Finally, more of an afterthought but along similar lines: should there be a collaborative processing code library that should be developed and maintained in parallel with the use of these data (similar to GitHub or SourceForge)?

CRCNS terms allow downloading all the data. The guidelines (not preconditions) suggest that “Unnecessary downloading of large data files is not permitted.” CRCNS just wants to avoid that industry and for-profit organizations will inappropriately use public data. If someone needs to download all the files for their scientific purpose, then it's OK. We modified the terms to allow sharing of downloaded data amongst direct colleagues working with the data or to students in a classroom. The CRCNS.org website does have forum that
allows users to upload attachments that contain code or to reference code being developed at other sites, such as GitHub. Thanks for the suggestion to have a collaboratively developed code library that is more directly associated with data sets.

**Competing Interests:** No competing interests were disclosed.

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**Referee Report 06 May 2014**

**doi:** 10.5256/f1000research.4171.r4607

**Shuzo Sakata**

Strathclyde Institute of Pharmacy and Biomedical Sciences, University of Strathclyde, Glasgow, UK

This is an important report for the neuroscience community regarding huge data sets recorded from the dorsal hippocampus and dorsomedial entorhinal cortex of behaving rats. Because the authors have already published a number of papers with the data sets, I have no doubt that this paper and their shared data will contribute to the further advancement of this field.

This manuscript is clearly written in details. This article can be improved by clarifying the following minor points in the section Data collection and cell-type classification:

1. **Cluster quality**
   It is not clear whether the quality of single units was assessed objectively, for example, by measuring isolation distance.

2. **Cell-type classification**
   Because now researchers including the authors can classify cell-types optogenetically, it would be better to comment on potential pitfalls of the classification procedure used here. For example, it is difficult to exclude the possibility that some of “principal” neurons can be interneurons.

**Competing Interests:** No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

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**Reader Comment 15 Jul 2014**

**Kenji Mizuseki**, Osaka City University Graduate School of Medicine, Japan

1. *“It is not clear whether the quality of single units was assessed objectively, for example, by measuring isolation distance.”*

To check the quality of spike sorting, we calculated isolation distance[^1], an interspike interval index “R[^2]2/10”[^2] and a fraction of interspike intervals less than 2 msec[^3]. In the revised manuscript, we explain that these values are available for each neuron so that one can choose well-isolated neurons by using these isolation quality measurements before downloading the data. We also mention that an initial quality control filtering was done in
selecting cells to include in the data set: 7943 cells were detected, but only 7736 of them were included in the data set; 207 were not included because they were judged to have insufficient quality.

2. “Because now researchers including the authors can classify cell-types optogenetically, it would be better to comment on potential pitfalls of the classification procedure used here. For example, it is difficult to exclude the possibility that some of “principal” neurons can be interneurons.”

This is an important point. Even though our classification of principal neurons and interneurons is supported by short-term spike cross-correlogram and consistent with previous reports misclassification of pyramidal cells and interneurons is inevitable to some extent by our method. For example, some types of interneurons with wide wave shapes (e.g. somatostatin positive interneurons) might have been classified as pyramidal cells by our method. The cell-type classification method should be verified and refined by optogenetical tools in the future. We discuss this point in the revised manuscript.

References


**Competing Interests:** No competing interests were disclosed.

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Referee Report 06 May 2014

doi:10.5256/f1000research.4171.r4602

Yoshikazu Isomura
Brain Science Institute, Tamagawa University, Tokyo, Japan

In this work, Mizuseki et al. provide valuable information on their large-scale data sets of multi-neuronal and local field potential (LFP) recordings from the hippocampus (dentate gyrus, CA3, and CA1) as well as the entorhinal cortex across the layers (layers 2, 3, 4, 5 etc.) of behaving rats. These data sets have originally been used for a number of their excellent studies for past ten years. Here, they fully summarized the experimental conditions and results in individual recording sessions very clearly for any users to plan to analyze the data. It will be a really useful treasure map for all hippocampus researchers.

I have one minor comment. While they showed the number of isolated neurons in each brain area of each animal (Table 2 to Table 4), it is not clear yet how many neurons, at most, were recorded simultaneously. Some people will probably want to analyze spike activity in a population of many neurons. The data would be more available for the higher-order spike analysis if the authors added additional information on the best (or first choice) data set with a sufficient number of well-isolated neurons in the same brain area.

**Competing Interests:** No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

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Reader Comment 15 Jul 2014

Kenji Mizuseki, Osaka City University Graduate School of Medicine, Japan

Indeed one should be able to know the number of neurons recorded in each recording sessions before downloading the data. Following the reviewer’s advice, we added tables giving the number of spikes recorded from each neuron during each session to the metadata tables describing the data set and we provide examples in document “crcns-hc3-data-description” of how to use these to find sessions useful for the researcher’s purpose before downloading the data. In addition, we include a new set of documents (in file “crcns-hc3-cell-counts.zip”) that lists the sessions with the most cells from each region (for both all cells and well isolated units) and also lists - for all sessions - the number of cells recorded from each region (both all cells and well isolated). These documents can also be used to select sessions with the largest number of simultaneously recorded neurons from particular regions before downloading the data.
Competing Interests: No competing interests were disclosed.

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