Fluoxetine pharmacokinetics and tissue distribution suggest a possible role in reducing SARS-CoV-2 titers [version 1; peer review: 2 approved with reservations]

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Abstract

Background. Recent in vitro studies have shown fluoxetine inhibits the severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) pathogen, including variants B.1.1.7 and B.1.351, SARS-CoV-2 spike mutations (E484K, K417N, N501Y), and one retrospective clinical study reported fluoxetine exposure at a median dose of 20 mg in patients with the SARS-CoV-2 coronavirus disease 2019 (COVID-19) had a significantly lower risk of intubation and death. The aim of this study is to conduct in silico population pharmacokinetic dosing simulations to quantify the percentage of patients achieving a trough level for the effective concentration resulting in 90% inhibition (EC90) of SARS-CoV-2 as reported in Calu-3 human lung cells.

Methods. Population pharmacokinetic parameter estimates for a structural one-compartment model with first-order absorption were used to simulate fluoxetine pharmacokinetic data. A population of 1,000 individuals were simulated at standard fluoxetine doses (20 mg/day, 40 mg/day, and 60 mg/day) to estimate the percentage of the patients achieving a trough plasma level for the EC90 SARS-CoV-2 inhibitory concentration for a 10 day treatment period. All analyses were conducted via statistical programming in R.

Results. Standard fluoxetine antidepressant doses resulted in a range of 81% to 97% of the patient population achieving a trough target plasma concentration of 23.2 ng/ml at day 10 and translates to a lung-tissue distribution coefficient of 60-times higher (EC90 of 4.02 mM). At a dose of 40 mg per day, at least 87% of patients will reach the trough target EC90 concentration within three days.

Conclusion. Overall, the findings of this population
pharmacokinetic dosing study corroborates in vitro and observational clinical studies reporting the first selective serotonin reuptake inhibitor fluoxetine inhibits the SARS-CoV-2 pathogen at commonly treated doses in the practice of psychiatry.

Keywords
Prozac, Sarafem, SARS-COV-2, COVID-19, antidepressant, pharmacokinetics, dose, lungs

This article is included in the Disease Outbreaks gateway.

This article is included in the Coronavirus collection.

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Author roles: Eugene AR: Conceptualization, Data Curation, Formal Analysis, Funding Acquisition, Investigation, Methodology, Project Administration, Resources, Software, Supervision, Validation, Visualization, Writing – Original Draft Preparation, Writing – Review & Editing

Competing interests: No competing interests were disclosed.

Grant information: The author(s) declared that no grants were involved in supporting this work.

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How to cite this article: Eugene AR. Fluoxetine pharmacokinetics and tissue distribution suggest a possible role in reducing SARS-CoV-2 titers [version 1; peer review: 2 approved with reservations] F1000Research 2021, 10:477 https://doi.org/10.12688/f1000research.53275.1

First published: 16 Jun 2021, 10:477 https://doi.org/10.12688/f1000research.53275.1
**Introduction**

The selective serotonin reuptake inhibitor (SSRI) fluoxetine is a racemic mixture of two stereoisomers, R-fluoxetine and S-fluoxetine, and maintains regulatory approvals for a wide-array of clinical indications in the practice of psychiatry. Two recent in vitro studies showed fluoxetine inhibits replication of the Severe Acute Respiratory Coronavirus-2 (SARS-CoV-2) pathogen (Schloer et al., 2020; Zimniak et al., 2020, 2021). Specifically, Zimniak et al. reported that following a three-day incubation period of fluoxetine in Vero cells, inoculated at a multiplicity of infection (MOI) of 0.5, resulted in the median maximal effective concentration (EC50) of 387 ng/ml (1.1 μM) and further found a concentration of 800 ng/ml (2.3 μM) significantly inhibited SARS-CoV-2 replication (Zimniak et al., 2020, 2021). Similarly, Schloer et al. found that fluoxetine significantly decreases SARS-Cov-2 titers, after a 48-hour incubation period, in both African green monkey kidney epithelial Vero E6 cells (EC50 = 0.69 μM and 90% maximal effective concentration [EC90] = 1.81 μM, MOI = 0.01) and human-lung Calu-3 cells (EC50 = 0.82 μM and EC90 = 4.02 μM, MOI = 0.1) (Schloer et al., 2020). Taken together these in vitro studies prove in a dose-dependent manner that the SSRI fluoxetine inhibits the SARS-CoV-2 pathogen known to cause the worldwide pandemic, the novel coronavirus disease 2019 (COVID-19).

Considering the COVID-19 clinical symptoms affecting the lungs, fluoxetine lung concentrations would be an important factor to consider when interpreting any study results. Johnson et al. reported human-tissue concentrations of fluoxetine in airline pilots in whole-blood ranged from 0.021–1.4 μg/ml and lung concentrations ranged from 1.56 μg/ml to 51.9 μg/ml, leading to a fluoxetine distribution coefficient of 60 (Johnson, Lewis & Angier, 2007). Clinically, the fluoxetine SARS-CoV-2 in vitro findings were corroborated by Hoertel et al. who showed in a multicenter observational retrospective cohort study of patients who were treated with fluoxetine and diagnosed with COVID-19, experienced a lower risk of intubation and death (hazard ratio = 0.32; 95% confidence interval, 0.14–0.73, p = 0.007) at a median fluoxetine dose of 20 mg (standard deviation [SD] = 4.82) (Hoertel et al., 2020). In this context, the aim of this study is to conduct in silico population pharmacokinetic dosing simulations to quantify the percentage of patients expected to achieve the trough effective concentration resulting in 90% inhibition of SARS-CoV-2.

**Methods**

**Pharmacokinetic model**

Pharmacometric model estimates for differential equation parameters and respective variances for a structural one-compartment pharmacokinetic model with first-order absorption were used to simulate fluoxetine concentration-time data. Model estimates were derived from drug plasma concentrations in 25 females taking a mean dose of 29.4 mg (7.5–80 mg/day) when fluoxetine plasma levels were at steady-state due to being collected for analysis at a minimum median time of fluoxetine treatment of greater than 40 days (Tanoshima et al., 2014). The following parameters were used: volume of distribution (Vd) value of 20.5 liters (variance [ω], 1.24), clearance rate (CL) value of 13.3 liters/hour (ω = 0.052), and absorption rate (Ka) of 0.016 (1/hour) (ω = 0.231) (Tanoshima et al., 2014).

**Target fluoxetine plasma concentration to achieve EC90 lung concentration**

The molecular weight of fluoxetine hydrochloride is 345.8 g/mol and the reported EC50 (0.82 μM) and EC90 (4.02 μM) values from the Schloer et al. study are equivalent to EC50 = 283.6 ng/ml and EC90 = 1390.1 ng/ml, respectively. The fraction of fluoxetine bound in human plasma is 94%, which leaves only 6% of the compound being unbound in human plasma (Sommi, Crismon & Bowden, 1987). Despite fluoxetine being highly protein bound, a study by Mantinieks et al. reported in paired fluoxetine concentrations of antemortem and postmortem cases (n = 18), fluoxetine has a human whole-blood to plasma ratio of 0.8–1.0, meaning that the whole-blood concentration is actually less than plasma or has up to a 1:1 ratio (Mantinieks et al., 2020). Further, Mantinieks et al. found the postmortem (range: 0.031–1.4 mg/L) to antemortem (range: 0.018–0.51 mg/L) fluoxetine drug concentration ratio is 1.8, but was not statistically significant as the p-value >0.05 and thus the 1.8 ratio is not applicable to this study (Mantinieks et al., 2020). Therefore, this study will directly translate the simulated plasma concentrations and apply the tissue distribution coefficients from the Johnson et al. study and the original preprint version of the manuscript is updated to account for the findings from Mantinieks et al. (Johnson, Lewis & Angier, 2007; Eugene, 2020; Mantinieks et al., 2020). Lastly, for all calculations, the trough target plasma concentration is referenced from the Schloer et al. study who reported after a 48-hour incubation period in Calu-3 lung cells the 90% maximal effective concentration is 4.02 μM (Schloer et al., 2020), which is significantly higher than the EC90 in Vero E6 cells (1.81 μM) and EC50 results from Zimniak et al. and the Schloer et al. studies (Schloer et al., 2020; Zimniak et al., 2020, 2021).

**Dosing simulations**

To estimate the percentage of patients from a population of one thousand simulated patients who would achieve the trough target EC90 concentration, pharmacokinetic dosing of fluoxetine consisted of three dosing trials of fluoxetine: 20 mg/day, 40 mg/day, and lastly 60 mg/day.
Software and statistics
All pharmacokinetic dosing simulations are conducted with a population of 1,000 patients using mrgsolve and pharmacokinetic parameter estimates using PKNCA in R version 3.6.3 (R Core Team, 2015). The overall R script has been adapted from a study published in Clinical Pharmacology and Therapeutics using hydroxychloroquine (Al-Kofahi et al., 2020; Eugene, 2021). Statistical results providing percentage estimates are calculated from trough concentrations of patients achieving the effective concentrations and is referenced from the Schloer et al. study reporting the EC90 value in human-lung Calu-3 cells (Schloer et al., 2020).

Results
The EC90 target fluoxetine lung concentration is 1390.1 ng/ml [4.02 μM] and 1/60 of this concentration is the new EC90-plasma concentration of 23.2 ng/ml [0.067 μM]. The percentage of the 1,000 simulated patients are illustrated in Figure 1 (20 mg/day), Figure 2 (40 mg/day), and Figure 3 (60 mg/day) with a horizontal dashed-line throughout the pharmacokinetic dosing figures showing the required trough EC90-plasma level of 23.2 ng/ml that translates to the EC90 level of 1390.1 ng/ml [4.02 μM] in the lungs.

Figure 1 shows the concentration-time data for a fluoxetine dose of 20 mg per day and results in the maximum plasma concentration (Cmax) with a geometric mean (geometric coefficient of variation, CV%) of 65.8 ng/ml (CV=70.2%), median time at maximum concentration (Tmax) of 220 hours (range, 49–220), area under the concentration-time curve (AUC0→Last) of geometric mean from baseline to 10 days of 10,200 ng·hour/ml, and a half-life (t½) – expressed as arithmetic mean (standard deviation, SD) – of 84.7 hours (SD = 181). These aforementioned pharmacokinetic results translate to 24% of the population reaching the target concentration at the end of day one and 81% of the population achieving the target trough EC90 concentration by end of day 10. Figure 2 shows at a dose of 40 mg per day, the Cmax is 132 ng/mL (CV = 68%), Tmax of 220 hours (range, 49–220), AUC0→Last is 20,500 ng·hour/ml, and population t½ is 81.4 (SD = 113), which is interpreted as 59% of the population achieving the EC90 trough target at day one and 93% by day 10. Moreover, Figure 3 shows a patient population treated with fluoxetine at 60 mg daily results in a Cmax of 191 ng/mL (CV = 71%), 220 hours (range, 49–232), AUC0→Last 29,700 ng·hour/ml, and t½ of 85.4 (SD = 209) allowing 74% of the population to reach the target trough concentration threshold on day one and 97% by day 10 of fluoxetine treatment. Table 1 provides an overview of the pharmacokinetics and pharmacodynamics with blood levels (ng/ml and μM) in plasma as well as calculated organ concentrations (whole-blood, lung, brain, heart, liver, spleen, and kidney) as well as the percent of the population achieving trough EC90 target during a treatment period of 10 days. All underlying
**Figure 2.** Fluoxetine population (n = 1,000) dosing simulation results for an oral dose of 40 mg/day for 10 days. The shaded regions illustrate the 10th (lower) and 90th (upper) percentiles with the solid line within the shaded region representing the median fluoxetine concentration. The dashed horizontal line depicts the effective concentration resulting in 90% inhibition (EC90) of SARS-Cov-2 that will result in 60-times higher level in the lungs.

**Figure 3.** Fluoxetine population (n = 1,000) dosing simulation results for an oral dose of 60 mg/day for 10 days. The shaded regions illustrate the 10th (lower) and 90th (upper) percentiles with the solid line within the shaded region representing the median fluoxetine concentration. The dashed horizontal line depicts the effective concentration resulting in 90% inhibition (EC90) of SARS-Cov-2 that will result in 60-times higher level in the lungs.
Table 1. Fluoxetine pharmacokinetics and pharmacodynamics showing blood levels (ng/mL and μM) and percent of population achieving plasma trough concentration of 23.2 ng/ml leading to a lung-EC90 target of 4.02 μM during a treatment period of 10-days in humans.

<table>
<thead>
<tr>
<th>Time Point</th>
<th>20 mg/day</th>
<th>40 mg/day</th>
<th>60 mg/day</th>
</tr>
</thead>
<tbody>
<tr>
<td>Day 1</td>
<td>24%</td>
<td>59%</td>
<td>74%</td>
</tr>
<tr>
<td>Day 2</td>
<td>47%</td>
<td>79%</td>
<td>90%</td>
</tr>
<tr>
<td>Day 3</td>
<td>60%</td>
<td>87%</td>
<td>93%</td>
</tr>
<tr>
<td>Day 4</td>
<td>67%</td>
<td>90%</td>
<td>94%</td>
</tr>
<tr>
<td>Day 5</td>
<td>72%</td>
<td>90%</td>
<td>95%</td>
</tr>
<tr>
<td>Day 6</td>
<td>76%</td>
<td>92%</td>
<td>95%</td>
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<tr>
<td>Day 7</td>
<td>78%</td>
<td>92%</td>
<td>96%</td>
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<tr>
<td>Day 8</td>
<td>80%</td>
<td>93%</td>
<td>96%</td>
</tr>
<tr>
<td>Day 9</td>
<td>81%</td>
<td>93%</td>
<td>96%</td>
</tr>
<tr>
<td>Day 10</td>
<td>81%</td>
<td>93%</td>
<td>97%</td>
</tr>
</tbody>
</table>

EC90: effective concentration inhibiting 90% of SARS-CoV-2 in Calu-3 lung cells reported as 4.02 uM (Schloer et al., 2020). Maximum plasma concentration (Cmax) is expressed as geometric mean (geometric coefficient of variation, CV%). Fluoxetine tissue distribution coefficients (60 for lung, 15 for brain, 10 for heart, 38 for liver, 20 for spleen, and 9 for kidneys) as reported (Johnson, Lewis & Angier, 2007).

Discussion
According to the United States Food and Drug Administration (FDA) Adverse Events Reporting System (FAERS) during the window period of 1982 to June 30, 2020, fluoxetine was reported to have a total of 79,929 cases, 62,948 serious cases, and 10,043 end of life cases (https://www.fda.gov/drugs/questions-and-answers-fdas-adverse-event-reporting-system-faers/fda-adverse-event-reporting-system-faers-public-dashboard). Females represented 58% of the adverse drug reactions (ADRs), males represented 27% of the ADRs, and 15% of the ADRs did not specify a gender. The most common adverse drug event reported for fluoxetine is Drug Interaction and amounts to 3,798 cases (4.75% of total). Given this information, drug interactions associated with fluoxetine are due to inhibition of the cytochrome P450 (CYP) system. Specifically, CYP2C19 and CYP2D6 may have interactions such as in patients taking tamoxifen for breast cancer by inhibiting conversion to the active endoxifen metabolite via CYP2D6 or in cases of clopidogrel in cardiology by inhibiting the conversion of clopidogrel to the active 2-oxo-clopidogrel metabolite (Spina, Trifirò & Caraci, 2012; Eugene, 2019).

Extrapolating from in vitro to in vivo concentrations are dependent on intracellular versus extracellular concentrations, as well as the methodology of quantifying either whole blood versus plasma concentrations in human pharmacokinetic studies. The EC50 and EC90 target concentrations represent the extracellular fluoxetine concentrations in the SARS-CoV-2 cell culture media. As COVID-19 is known to affect the brain during active infection and in post-COVID-19 states, adequate brain concentrations would be clinically important in patients who may experience depression. Bolo et al. reported fluoxetine brain concentrations, at steady-state, using fluorine magnetic spectroscopy and showed fluoxetine pharmacokinetic study data in an.xlsx format, the one-compartment population pharmacokinetic model file in C++ format, and the R programming script are available (Eugene, 2021).
concentrations were 10-times higher in the brain than in human plasma (Bolo et al., 2000). Specifically, Bolo et al. found in study participants taking oral doses (10mg, n = 1; 20 mg, n = 1; 40 mg, n = 2) with a treatment period ranging from three months to 12-months that fluoxetine human brain concentrations were 13 μM (SD = 7) versus 1.73 μM (SD = 1.00) in human plasma fluoxetine (Bolo et al., 2000). In comparison, Johnson et al. found the coefficients for tissue distribution of fluoxetine relative to whole blood was: 60 for lung, 15 for brain, 10 for heart, 38 for liver, 20 for spleen, and 9 for kidneys (Johnson, Lewis & Angier, 2007).

As patients recover from the acute COVID-19 symptoms, long-term sequelae are being documented and in one of the post-SARS-Cov-2 infection studies in young patients, 92% were found to have ongoing cardiorespiratory symptoms with organ dysfunction and impairment in the lungs (33%), heart (32%), kidneys (12%) (Dennis et al., 2020). In another post-COVID-19 syndrome study, 96% of the patients were female and experienced statistically significant exercise intolerance, dyspnea, and chest pain when compared to those not diagnosed with COVID-19 (Walsh-Messinger et al., 2020). Moreover, Walsh-Messinger et al. found patients with post-COVID-19 syndrome had higher ratings of depression subscale markers of altered sleep and thinking, but depression severity was not significantly different with patients not diagnosed with COVID-19 (Walsh-Messinger et al., 2020).

Direct clinical translation of this current pharmacokinetic study corroborates with a retrospective multicenter observational study by Hoertel et al., who found a median fluoxetine dose of 20mg/day resulted in a significantly lower risk of intubation and death in a population composed of 63% women and 37% men (Hoertel et al., 2020). Comparing the Hoertel et al. and Zimmniak et al. publications, Hoertel et al. found that in addition to fluoxetine, venlafaxine (median dose of 75mg/day) and escitalopram (median dose of 10mg) were also associated with a lower risk of intubation and death, however, Zimmniak et al. showed that neither escitalopram nor paroxetine inhibited SARS-CoV-2 in vitro (Hoertel et al., 2020; Zimmniak et al., 2020, 2021). Of note, as shown in Table 1, a 40 mg or 60 mg daily fluoxetine dose results in 90% inhibition of the SARS-CoV-2 infection due to surpassing the EC90 value of 4.02 μM as found in Calu-3 cells and the EC90 value of 1.81 μM in Vero E6 cells (Schloer et al., 2020).

Antiviral properties of fluoxetine are well reported in the literature. Carpinteiro et al. reported that fluoxetine inhibits acid sphingomyelinase preventing infection of both cultured cells and human nasal epithelial cells in SARS-CoV-2, as well as in vesicular stomatitis virus pseudoviral particles presenting the SARS-CoV-2 spike protein (Carpinteiro et al., 2020). A study by Zuo et al. showed fluoxetine resulted in potent inhibition of the coxsackievirus by reducing both synthesis of viral RNA and protein (EC50 of 2.3 μM) exhibiting peak antiviral properties at 6.25 μM (Zuo et al., 2012). Bauer et al. showed, in a broad-spectrum manner, fluoxetine inhibited enterovirus (picornaviridae family) replication with the R-fluoxetine-EC50 values alone for rhinovirus HRV-A2 (EC50 = 7.95 μM) and HRV-B14 (EC50 = 6.34 μM) (Bauer et al., 2019). Notably, Zimmniak et al. found that individual stereoisomers, R-fluoxetine and S-fluoxetine, inhibited the SARS-CoV-2 viral load; however, in contrast, fluoxetine could not inhibit gene expression of the herpes simplex-1 virus, human herpes virus-8, rabies virus, nor the respiratory syncytial virus (Zimmniak et al., 2020, 2021). Lastly, as shown in Table 1, standard fluoxetine doses are capable of achieving the aforementioned EC50s for all of the aforementioned microbes.

A limitation of this study is associated with the previously validated fluoxetine pharmacometric model being in women and did not include men (Tanoshima et al., 2014). However, as shown from the aforementioned FAERS data, women represented 58% of all ADR cases overall from 1982 to 2020. A significant study strength is that a study from the University of Helsinki did not include men (Tanoshima et al., 2014). However, as shown from the aforementioned FAERS data, women represented 58% of all ADR cases overall from 1982 to 2020. A significant study strength is that a study from the University of Helsinki reported fluoxetine inhibits SARS-CoV-2 variants (B.1.1.7 and B.1.351) and the spike mutations (E484K, K417N, N501Y) (Fred et al., 2021). Overall, from a drug-safety perspective, prior to administering fluoxetine, a careful review of all patient medications and clinical status by a clinical pharmacologist physician would be recommended to avoid drug interactions due to fluoxetine’s ability to strongly inhibit CYP2C19 and CYP2D6 (Hefner, 2018). Compounds that are sensitive and moderate CYP2C19 substrates (e.g. omeprazole, diazepam, lansoprazole, rabeprazole, voriconazole) and CYP2D6 substrates (e.g. dextromethorphan, eliglustat, nebivolol, tolterodine, encaïnide, metoprolol, propranolol, tramadol) will have an increased total area under the concentration-time curve of ≥ 5-fold drug exposure when treated with fluoxetine (https://www.fda.gov/drugs/drug-interactions-labeling/drug-development-and-drug-interactions-table-substrates-inhibitors-and-inducers). Lastly, patients who have a pharmacogenomic profile of being a CYP2D6 Poor Metabolizer or CYP2D6 Intermediate Metabolizer should be closely monitored for potential fluoxetine side-effects; but they may also have a higher rate of achieving the target trough EC90 concentration at a 20 mg daily fluoxetine dose relative to CYP2D6 Normal (Extensive) Metabolizers.
Conclusions
This study investigated fluoxetine pharmacokinetics and human organ tissue distribution which confirmed that previously published median effective concentrations and specifically the EC90 fluoxetine value inhibiting SARS-CoV-2 in Calu-3 human lung cells are achievable using standard fluoxetine doses (20mg/day, 40mg/day, and 60mg/day) and also corroborates findings from a retrospective clinical study showing fluoxetine exposure was associated with reduced risk of intubation and death. Overall, assuming patients are not treated with medications that result in drug-drug interactions with fluoxetine, a dose of 40 mg per day of fluoxetine will likely be most effective with inhibiting the SARS-CoV-2 viral titers with 59% of the population achieving the trough EC90 target on day one, 92% by day seven, and 93% of patient population achieving the trough target EC90 concentration to inhibit the SARS-CoV-2 within 10 days.

Data availability
Underlying data
This project contains the following underlying data:

- Data File 1: fluoxetine_20mg_PO_QAM.xlsx
- Data File 2: fluoxetine_40mg_PO_QAM.xlsx
- Data File 3: fluoxetine_60mg_PO_QAM.xlsx

Software availability
This project contains the following software:

- popkp_fluoxetine_sars_cov2_inhibition.cpp
- Fluoxetine_pharmacokinetic_sars_cov2_simulation_script.R

Data are available under the terms of the Creative Commons Zero “No rights reserved” data waiver (CC0 1.0 Public domain dedication).

Acknowledgements
This author acknowledges the researchers who conducted the in vitro studies, as well as the retrospective clinical study that encouraged this population pharmacokinetic dosing study with fluoxetine to be realized.

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Eugene AR: Data for Fluoxetine pharmacokinetics and tissue distribution suggest a possible role in reducing SARS-CoV-2 titers.

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R Core Team: R: A Language and Environment for Statistical Computing. 2015.


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PubMed Abstract | Publisher Full Text | Free Full Text
Open Peer Review

Current Peer Review Status:  ?  ?

Reviewer Report 23 June 2021

https://doi.org/10.5256/f1000research.56641.r87690

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Eero Castren
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Andy Eugene has written an interesting paper on a very timely topic. Preclinical as well as limited clinical data support the idea that some antidepressants and related compounds may protect cells from SARS-CoV2 infection, but it has been unclear whether concentrations that are found in cell culture studies to be inhibitory are really achieved in clinical treatments. It is unfortunate that there is only limited amount of high-quality data on the tissue concentrations and distribution of fluoxetine in human tissues, but Eugene has taken advantage of the data that is available to model fluoxetine pharmacokinetics in human lung tissue. His results suggest that sufficiently high lung tissue concentrations are achieved within a 10 day treatment period in majority of patients with a standard 20 mg/day dosing, and with higher doses, the trough concentration is achieved within the first days of treatment. The simulations appear properly conducted and the manuscript is well written. I have a few issues that might improve the manuscript further.

1. The conclusions about how fast trough concentrations are achieved is based on the assumption that fluoxetine immediately distributes to lung tissue at the estimated 60 fold concentration compared to plasma levels. However, there is some (although clearly incomplete) information that fluoxetine may accumulate into tissues over time. Rasenick and colleagues have found that certain antidepressants accumulate in lipid rafts in vitro over several days of exposure (Erb et al. 2016). For human brain, Karson et al. have found that, using spectroscopy, fluoxetine/norfluoxetine is not detectable in human brain at about 1 week, but at about 2-3 weeks of treatment, concentrations of several micromolar can be assayed (Karson et al. (1993)) indicating that fluoxetine accumulates in brain over time. If the same is true for lung tissue, the 60-fold concentrations may not be reached in lung tissues as quickly as estimated here. Although paucity of information prevents any proper estimations, I think it would be important to discuss this issue in the discussion section.

2. The author should be complemented for indicating in many places in the manuscript fluoxetine concentrations in both ng/ml and in molar units, however, this is not done
systematically. It would be very helpful to, wherever possible, indicate both units. In the figures, it may not be feasible to have double set of units in the Y-axis, I would prefer molar units, but if the author prefers ng/ml, that is not a major problem.

3. The author may want to mention that a clinical phase 3 trial evaluating the effect of fluvoxamine vs. placebo is ongoing and should be completed by September this year.

References

Is the work clearly and accurately presented and does it cite the current literature?
Yes

Is the study design appropriate and is the work technically sound?
Yes

Are sufficient details of methods and analysis provided to allow replication by others?
Yes

If applicable, is the statistical analysis and its interpretation appropriate?
Yes

Are all the source data underlying the results available to ensure full reproducibility?
Yes

Are the conclusions drawn adequately supported by the results?
Partly

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Neuropharmacology

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Reviewer Report 21 June 2021

https://doi.org/10.5256/f1000research.56641.r87881
This is a review of the manuscript "Fluoxetine pharmacokinetics and tissue distribution suggest a possible role in reducing SARS-CoV-2 titers" submitted for publication in F1000Research. This is a very interesting and important study, which addresses a timely and important question – in a context of a growing number of preclinical and clinical studies showing a potential efficacy of certain antidepressants, and particularly fluoxetine, in COVID-19, what dose of fluoxetine should be optimally prescribed in clinical trials to ensure a maximum of patients achieve the effective concentration resulting in 90% inhibition (EC90) of SARS-CoV-2 as reported in Calu-3 human lung cells. On a more general, and potentially more innovative front, it is one of a small (but quickly increasing) number of studies contributing to this topic and the first study to the reviewer's knowledge addressing this specific pharmacokinetic issue, which is an important area of inquiry with major implications in this context of pandemic. There is much to like about this manuscript. The results are presented clearly, the methods are sound, the discussion follows well from it, and the manuscript is very well written. Below are several points that would strengthen the submission:

Title:
- A minor suggestion would be a modification of the title to increase the interest to this important study, such as "Fluoxetine pharmacokinetics and tissue distribution suggest a possible therapeutic role in COVID-19 at usual antidepressant dose".

Introduction:
- The introduction section should be updated and include a description of results from recent studies focused on the association between antidepressant use and the course of COVID-19, which would be very helpful to readers to understand the clinical context of this study and why this study (and its results) are important. In addition to the medRxiv one, the reference to the published article of Hoertel N et al. should be cited (Hoertel N et al. (2021¹)). This association/potential effect of certain antidepressants in COVID-19 has been confirmed in several other observational studies (Diez-Quevedo et al. (2021²), Hoertel et al. (2021³), Hoertel et al. (2021⁴)), two clinical trials (1 RCT and 1 open-label study) with the antidepressant fluvoxamine (which is, like fluoxetine, a functional inhibitor of sphingomyelinase acid and a S1R agonist) (Lenze et al. (2020⁵), Seftel & Boulware (2021⁶)), and other preclinical studies (Dechaumes et al. (2021⁷), Fred et al. (2021⁸)). Results from all those studies, increasing the interest in the present submission, should be recognized and briefly summarized in the Introduction.
- As evoked in the discussion, potential underlying mechanisms include antiviral effect of certain antidepressants, and particularly fluoxetine, through the inhibition of the acid sphingomyelinase/ceramide system, which may play a central role in COVID-19, as
suggested by preclinical studies (Carpinteiro et al. (2020)\(^9\), and Carpinteiro et al. (2021)\(^{10}\)) and observational studies (Hoertel N \textit{et al.} (2021)\(^3\), Darquennes G \textit{et al.} (2021)\(^{11}\), Hoertel \textit{et al.} (2021)\(^4\)), as well as anti-inflammatory properties through sigma-1-receptor agonist effect of certain antidepressants, and particularly fluoxetine and fluvoxamine (Köhler CA \textit{et al.} (2018)\(^{12}\), Rosen D \textit{et al.} (2019)\(^{13}\), Sukhatme VP \textit{et al.} (2021)\(^{14}\), Roumestan C \textit{et al.} (2007)\(^{15}\)). Moreover, a recent study (Marín-Corral \textit{et al.} (2021)\(^{16}\)) showed that ceramide plasma concentration is associated with inflammatory markers and clinical outcomes in COVID-19 patients. I think that results from all those studies should also be recognized and shortly summarized in the Introduction.

- The abstract might include some of these points listed above.

**Results:**

- Importantly, results from that study can help guide the choice of the right dose to use in COVID-19 clinical trials. I would strongly recommend the author to present in the result section as well as in the abstract (for main findings) for each following dose: 20 mg/d, 30 mg/d (which is also interesting for clinical tolerability purpose), 40 mg/d and 60 mg/d the percentage of patients reaching a) EC90 and b) EC50 at (i) 1 day, (ii) 2 days, and (ii) 3 days. In addition, given that 40 mg/d might lead to increased risk of mild side effects among older adults and that at the same time reaching the efficient dose as fast as possible is desirable given the potential antiviral properties of fluoxetine, would a loading dose of 30 mg or 40 mg the first or two first days, followed by a dose of 20 mg/day, perform better than only taking 20 mg/d?

- Finally, is there a way to ensure that at no point of the 10 day treatment course, no patient (typically older adults) will reach a toxic plasma dose with the different scenarios, as I think 40 mg/d is the maximum recommended dose in the elderly population? Even if the reviewer recognizes that it may represent a substantial additional amount of work, these detailed results will certainly be of great help to clinicians to balance the risks associated for each dose and the potential benefits, and help them design ideal clinical trial.

- With the different scenarios, at the end of the 10-day treatment, how long would it take to have a substantial decrease in the fluoxetine plasma concentration (and its metabolite norfluoxetine)? This information would give an idea of the duration of potential protection of the treatment against SARS-CoV-2 once stopped and of the required duration of medical follow-up.

**Conclusion:**

- Finally, with these different dose scenarios, what recommendation(s) of dose would do the author for a clinical trial including fluoxetine for COVID-19? Would it be desirable to do a loading dose then decrease to the standard 20 mg/d dose which is known to be very well tolerated in older adults? Or keeping 30 mg/d or 40 mg/d throughout the trial would be the best choice?

**References**


Is the work clearly and accurately presented and does it cite the current literature?
Yes

**Is the study design appropriate and is the work technically sound?**
Yes

**Are sufficient details of methods and analysis provided to allow replication by others?**
Yes

**If applicable, is the statistical analysis and its interpretation appropriate?**
Yes

**Are all the source data underlying the results available to ensure full reproducibility?**
Yes

**Are the conclusions drawn adequately supported by the results?**
Yes

**Competing Interests:** No competing interests were disclosed.

**Reviewer Expertise:** Psychiatry; antidepressants; methodology; biostatistics; epidemiology; sphingomyelinase; COVID-19.

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

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