ANTIBODY VALIDATION ARTICLE

Immunoblotting validation of research antibodies generated against HS1-associated protein X-1 in the human neutrophil model cell line PLB-985. [version 1; referees: 3 approved]

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Abstract
HS1-associated protein X-1 (Hax1) is a 32 kDa protein that is ubiquitously expressed. Hax1 is an anti-apoptotic protein with additional roles in cell motility, and autosomal recessive loss of Hax1 results in Kostmann syndrome, a form of severe congenital neutropenia. Because of the important role of Hax1 in neutrophils we demonstrate here validation of two research antibodies directed against human Hax1 in the human neutrophil model cell line PLB-985 cells. We show that both the mouse anti-Hax1 monoclonal IgG directed against amino acids 10-148 of Hax1 and a rabbit anti-Hax1 polyclonal IgG antibody directed against full-length Hax1 reliably and consistently detect Hax1 during immunoblotting of three different PLB-985 cell densities. Using shRNA mediated Hax1 knockdown, we demonstrate the specificity of both Hax1 antibodies. In addition, our results suggest that the rabbit anti-Hax1 polyclonal antibody is provides a stronger intensity in detecting Hax1 protein, with detection in as few as 0.1 x 10^6 cells in 6 total replicates we have performed.

Keywords
Hax1, neutrophil, PLB-985, tubulin

This article is included in the Antibody Validations gateway.
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**Competing interests:** No competing interests were disclosed.  

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Introduction

HS1-associated protein X-1 (Hax1) is a 32 kDa protein consisting of 279 amino acids that is ubiquitously expressed. Hax1 has been demonstrated to be a negative regulator of apoptosis in many immune cell types. Furthermore, Hax1 has been shown to have additional roles in regulating cell motility and adhesion, and is overexpressed in many types of cancer. Patients with autosomal recessive mutations in the HAX1 gene have a form of severe congenital neutropenia called Kostmann syndrome. Severe congenital neutropenia is characterized by early recurrent bacterial infections and decreased neutrophil counts in the bloodstream.

Because of the recent increase in Hax1 investigations, it is important to identify reliable antibodies directed against Hax1. Using the human neutrophil model cell line PLB-985 cells, which can be terminally differentiated into neutrophil-like cells after treatment with DMSO, we demonstrate the applicability and selectivity of two antibodies against Hax1. A mouse Hax1 monoclonal antibody (BD Biosciences) that is routinely used in publications investigating Hax1 directed against Hax1 amino acids 10-148, and a rabbit polyclonal antibody (Proteintech Group, Inc.) directed against the full length Hax1 protein.

Materials and methods

Reagent details

Details of all reagents used in the Western blotting procedures can be found in Table 1.

Antibody details

Anti-tubulin (beta-) is a mouse monoclonal IgG1 [E7 was deposited to the DSHB by Klymkowsky, Michael (DSHB Hybridoma Product E7)] and was used as a loading control for all Western blots at a dilution of 1:1000 resulting in a final concentration of 45 ng/mL. Rabbit anti-Hax1 (Proteintech Group, Inc, Table 2) is a polyclonal antibody generated to full length Homo sapiens Hax1. The lot number used was 1, and a dilution of 1:1000 was used for all Western blots resulting in a final concentration of rabbit anti-Hax1 of 230 ng/mL. Mouse anti-Hax1 (BD Biosciences) is a mouse monoclonal IgG1 raised against Homo sapiens Hax1 amino acids 10–148. The lot number used was 3266979, and a dilution of 1:1000 was used for all Western blots resulting in a final concentration of 250 ng/mL. Goat anti-rabbit IgG IRDye 680LT and Goat antimouse IgG IRDye 800CW (Li-Cor Biosciences, Table 2) were used at a dilution of 1:40,000 (25 ng/mL).

Cell culture

PLB-985 cells were maintained in RPMI 1640 (Mediatech, Inc.) supplemented with 10% fetal bovine serum, 60 μg/mL penicillin, and 100 μg/mL streptomycin (Mediatech, Inc.) at a concentration of 0.1–1 × 10^6 cells/mL. To differentiate PLB-985 cells into “neutrophil-like” cells 1.25% DMSO (Fisher Scientific) was added to 2 × 10^5 cells/mL for 6 days. Lentiviral Hax1 shRNA targets were purchased from Open Biosystems. Targets used; Hax1 shRNA (5’-ACAGACACTTCGGGACTCAAT-3’) and control shRNA (5’-TGTCTCCGAACGTGTCACGTT-3’). HEK293-FT cells were

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**Table 1. Details of reagents used for immunoblotting.**

<table>
<thead>
<tr>
<th>Process</th>
<th>Reagent Description</th>
<th>Manufacturer</th>
<th>Catalogue number</th>
<th>Concentration</th>
</tr>
</thead>
<tbody>
<tr>
<td>Protein blotting</td>
<td>0.45μm nitrocellulose pure transfer membrane</td>
<td>Santa Cruz Biotechnology</td>
<td>Sc-201705</td>
<td></td>
</tr>
<tr>
<td>Wash Buffer, blocking buffer, and Antibody Diluent</td>
<td>Tween-Tris/Saline (T-TS)</td>
<td>Fisher Scientific</td>
<td>Tris base BP152 NaCl S271 Tween-20 BP337</td>
<td>50mM Tris 150mM NaCl 1% Tween-20</td>
</tr>
<tr>
<td>Blocking</td>
<td>Bovine Serum Albumin, heat shock fraction</td>
<td>Sigma-Aldrich</td>
<td>A9647</td>
<td>5% in T-TS</td>
</tr>
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</table>

**Table 2. Details of Primary and Secondary Antibodies.**

<table>
<thead>
<tr>
<th>Antibody</th>
<th>Manufacturer</th>
<th>Catalogue number</th>
<th>RRID</th>
<th>Concentration</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tubulin (beta-)</td>
<td>Developmental Studies Hybridoma Bank</td>
<td>E7-s</td>
<td>RRID:AB_528499</td>
<td>45 ng/mL</td>
</tr>
<tr>
<td>Hax1</td>
<td>BD Biosciences</td>
<td>610824</td>
<td>RRID:AB_398143</td>
<td>250 ng/mL</td>
</tr>
<tr>
<td>Hax1</td>
<td>Proteintech Group, Inc.</td>
<td>11266-1-AP</td>
<td>RRID:AB_2263720</td>
<td>230 ng/mL</td>
</tr>
<tr>
<td>Goat anti-Rabbit IgG IRDye 680LT</td>
<td>Li-Cor Biosciences</td>
<td>926-32221</td>
<td>RRID:AB_621841</td>
<td>25 ng/mL</td>
</tr>
<tr>
<td>Goat anti-Mouse IgG IRDye 800CW</td>
<td>Li-Cor Biosciences</td>
<td>827-08364</td>
<td>RRID:AB_10793856</td>
<td>25 ng/mL</td>
</tr>
</tbody>
</table>
grown to 70% confluency in a 10cm tissue culture dish for each lentiviral target and transfected using 6μg Hax1, 0.6μg vesicular stomatitis virus (VSV)-G, and 5.4μg cytomegalovirus (CMV) 8.9.1. 72 hour viral supernatant was collected and concentrated using Lenti-X concentrator (Clontech, Inc.) following the manufacturer’s instructions. 1 × 10^6 PLB-985 cells were infected with viral supernatant for 3 days in the presence of polybrene (4 μg/mL, Santa Cruz Biotechnology). Stable cell lines were generated with puromycin (1 μg/mL, Sigma Aldrich) selection.

**Immunoblot analysis**

- Differentiated PLB-985 cells were counted and 0.1 × 10^6, 0.5 × 10^6, and 1 × 10^6 cells were pelleted by centrifugation.
- Cells were lysed in Triton X-100 lysis buffer with protease inhibitors (25 mM HEPES, pH 7.5, 150 mM NaCl, 1% TX-100, 10 mM MgCl_2, 1 mM EDTA, 10% glycerol, 1 μg/mL pepstatin A, 2 μg/mL aprotinin, 1 μg/mL leupeptin) on ice for 10 minutes and clarified by centrifugation.
- Cellular lysate was then removed and added to Laemmli sample buffer, boiled at 90°C for 5 minutes, and run on 10% SDS-PAGE gels.
- Proteins were then transferred to 0.45μm nitrocellulose membranes (Santa Cruz Biotechnology) at 400mA for 1 hour at 4°C.
- Following transfer, the membrane was blocked in 5% BSA in 1x T-TS for 1 hour at room temperature with gentle rocking.
- Membranes were then probed with mouse anti-tubulin (beta-) (45 ng/mL), and either mouse anti-Hax1 (BD Biosciences, 250 ng/mL) or rabbit anti-Hax1 (Proteintech Group, Inc., 230 ng/mL) at room temperature for 1 hour.
- After primary antibody incubation the membranes were washed 3 × 5 minutes with 1× Tris-HCL/NaCl saline buffer (1× T-TS), see Table 1.
- The membranes were incubated with goat anti-rabbit IgG IRDye 680LT and goat anti-mouse IgG IRDye 800CW (Li-Cor Biosciences, 25 ng/mL).
- After secondary antibody incubation the membranes were washed 3 × 5 minutes with 1× T-TS.
- Blots were imaged with an infrared imaging system (Odyssey Fc; Li-Cor Biosciences) using a 2-minute exposure time.

**Results**

To determine the reproducibility and sensitivity of the mouse and rabbit anti-Hax1 antibodies on the PLB-985 cells, we performed Western blot analysis using three separate cell densities, 0.1 × 10^6, 0.5 × 10^6, and 1 × 10^6 cells. In our research using the PLB-985 cell system, we routinely use 1 × 10^6 – 10 × 10^6 cells in a Western blot. Using beta-tubulin as a loading control our Western blots illustrate an increasing protein concentration in the three samples as would be expected with increasing cell densities. We found that the mouse anti-Hax1 antibody (BD Biosciences) is visible as low as 0.5 × 10^6 cells, binding to a protein band at the expected Hax1 size of 32 kDa (Figure 1). In six different experiments (Figure 1 and Figure 4) we found inconsistency in protein detection with the Ms anti-Hax1 antibody. In all blots Hax1 was visible, however with varying degrees of intensity. Conversely, when the rabbit anti-Hax1 antibody (Proteintech Group, Inc.) was used, the antibody gave consistent and robust detection (Figure 2 and Figure 4). In some cases, Hax1 can be detected in as low as 0.1 × 10^6 cells using the Rb anti-Hax1 antibody (Figure 2C). We do not believe the difference between the two antibodies is due to variations in the cell extract or imaging software because when the same cell extract is immunoblotted on two different blots and scanned simultaneously the difference in sensitivity can be observed (Figure 3A). Using the Odyssey imaging system (Li-Cor Biosciences) to measure the intensity of each band, we calculated the intensity ratio of Hax1 relative to the tubulin loading control (Figure 3B). In both blots the levels of tubulin are similar, however it is evident that the

**Figure 1.** Detection of Hax1 in differentiated PLB-985 cells using a mouse anti-Hax1 antibody. Western blot analysis of differentiated PLB-985 cell lysates from 0.1 × 10^6, 0.5 × 10^6, and 1 × 10^6 cells from three independent replicates (A–C). Mouse anti-tubulin (beta-) is used as a loading control and can be seen present at 55 kDa. Mouse anti-Hax1 detects a band at approximately 32 kDa as predicted. Hax1 can be detected in densities of 0.5 × 10^6 and 1 × 10^6 cells.
rabbit anti-Hax1 antibody exhibits a stronger signal compared to the mouse monoclonal antibody. However, it should be noted that both antibodies reliably detect Hax1 in differentiated PLB-985 cells.

To demonstrate the specificity of both Hax1 antibodies we generated stably-expressing control shRNA and Hax1 shRNA PLB-985 cells (Figure 4). As described previously using the mouse anti-Hax1 antibody the control shRNA cells show inconsistent staining intensity, however it is clear that in these samples it is more robust than in the wild-type PLB-985 cells. Both the mouse anti-Hax1 and rabbit anti-Hax1 antibodies show reduced detection in the Hax1-deficient PLB-985 cells, which demonstrates that the antibodies are highly specific for Hax1. In many of the experiments we observed additional background bands that could be attributed to the goat anti-rabbit IgG secondary antibody (Figure 5).

**Dataset 1. Raw data for Figure 3**

http://dx.doi.org/10.5256/f1000research.6516.d47582

Comparison of mouse and rabbit anti-Hax1 antibodies in differentiated PLB-985 cells. Quantification of the band intensities was measured and the ratios of Hax1 to tubulin were graphed.
Conclusion

Here we show validation and comparison results of two antibodies generated against HS1-associated protein X-1, an anti-apoptotic protein that has a multi-factorial role in regulating cell proliferation and differentiation, cell motility, and cancer. Homozygous loss-of-function of Hax1 results in severe congenital neutropenia, a life threatening loss of circulating neutrophils in the bloodstream. Thus studying the function of Hax1 in primary neutrophils and the neutrophil model cell line PLB-985 will help elucidate the disease pathogenesis of neutropenia syndromes. We demonstrate...
that mouse anti-Hax1 (BD Biosciences) and rabbit anti-Hax1 (Proteintech Group, Inc.) are both specific for Hax1. Furthermore, we show that as little as $0.5 \times 10^6$ differentiated PLB-985 cells can be used to reliably detect Hax1 expression with both of the antibodies. We have evidence that the rabbit anti-Hax1 (Proteintech Group Inc.) results in a more robust and consistent detection of Hax1, likely due to the polyclonal nature of the antibody. Finally, lentiviral knockdown of endogenous Hax1 expression results in loss of Hax1 detection by both mouse anti-Hax1 and rabbit anti-Hax1 demonstrating the specificity of each antibody.

In conclusion we recommend the use of either mouse or rabbit anti-Hax1 antibodies shown here for studies using the PLB-985 cells as a neutrophil model cell line. Furthermore, it is our conclusion that a minimum cell density of $0.5 \times 10^6$ neutrophils should be used as a starting point for immunoblotting of Hax1, with greater than or equal to $1 \times 10^6$ cells being optimal.

Data availability

_F1000Research_ Dataset 1. Raw data for Figure 3, 10.5256/f1000research.6516.d475821

References

Open Peer Review

Current Referee Status: ✔ ✔ ✔

Version 1

Referee Report 08 July 2015
https://doi.org/10.5256/f1000research.6992.r9247

Lawrence L. LeClaire III
Department of Biochemistry and Molecular Biology, University of South Alabama, Mobile, AL, USA

Kristina and Cavnar present data to validate two commercial antibodies for HSA-associated protein X-1 in a human neutrophil cell line. The authors describe the conditions for the experiments as well as provide sufficient information/protocols for reader about the conditions used in that assays. In addition, the authors present the entire blots and describe the secondary bands that were detected. Only minor changes in the text should be made (below).

1. The commercial source of the antibody should be listed in the abstract. It currently gives the impression that the antibodies used in the study were generated by the authors.

2. The electrophoretic transfer conditions should be listed. Transfer buffer contents (%MeOH) as well as the model of transfer equipment (dry, semi-dry) will cause variability in protein transfer/antibody detection.

3. Did the authors attempt to vary blocking conditions (milk, commercial blocking agents)?

4. Is purified HAX1 available to use as a positive control or to build a standard curve for quantifying total HAX1 in the samples? Not necessary for this publication but would be extremely useful. Figure 3B suggests that detection is close to linear (especially with LiCor technology).

5. Authors should denote the size of bands on an SDS-PAGE as "relative mobility." For example, a band with a relative mobility of 32 kDa was detected...

6. In Figure 3, change "After transfer, the membrane was cut." To "the membrane was divided"

7. Are the authors certain that the band detected by the secondary antibody is the lower of the two bands in the experiments? A single lane could be divided and one side probed with primary and the other with only secondary to confirm.

Competing Interests: No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.
Peter Cavnar, University of West Florida, USA

Thank you for your kind review. I have revised and submitted a new version of this manuscript. Per your questions I will address each one:

1. I have included the commercial availability of the antibodies in the abstract, introduction, and discussion.

2. The transfer buffer conditions have now been included in Table 1.

3. We have tried blocking with non-fat dry milk and do not find any differences in the selectivity of the Hax1 antibodies, however we did get considerable background from the milk in the goat anti-rabbit 680 channel. We have tested this with a blank membrane. Therefore we will traditionally use 5% BSA to block.

4. We do have bacterial constructs that could be used to purify Hax1. Naturally that was not the purpose of this study, but I concur that these antibodies do behave quite linearly.

5. Thank you for this suggestion, we have included all sizes as a relative mobility in our results and figure legends.

6. We have edited figure 3 legend to reflect this.

7. We have revised figure 5 which demonstrates that the background band that has a relative mobility of 32 kDa is from the secondary antibody.

Competing Interests: No competing interests were disclosed.

Referee Report 06 July 2015

https://doi.org/10.5256/f1000research.6992.r9249

Mautusi Mitra
Department of Biology, University of West Georgia, Carrollton, GA, USA

The paper titled "’Immunoblotting validation of research antibodies generated against HS1-associated protein X-1 in the human neutrophil model cell line PLB-985," by Kristina and Cavnar focuses on the validation of two commercial antibodies generated against the human Hax1 protein. The experiments are in most part well designed. The research approaches, methods and the reagents used for the experiments are well described. The researchers have shown the entire Western blot and have also used Hax1-deficient PLB-985 cell lines to demonstrate the specificity of the two Hax1 antibodies. The data presented in the article strongly indicate that both the antibodies, especially the rabbit anti-Hax1 polyclonal antibody, can be used for specific detection of the human Hax1 protein and also shows the minimum cell density of neutrophils that can be used for immunoblotting experiments.

The manuscript is suitable for indexing, provided the authors address few issues, which are described below:
1. It is not clear if the antibodies were purchased commercially or were generated by the research laboratory with the help of a commercial company. This should be clarified.

2. Hax1 interacts with the polycystic kidney disease protein PKD2, located in the ER membrane/plasma membrane protein. Have the authors checked the efficiency of protein solubilization in their experiments? Have the authors done western analyses on a blot which has proteins transferred from a SDS-PAGE gel that has the intact stacking gel? If the solubilization is incomplete, insoluble proteins will be detected by the respective antibody in the stacking wells in the blot. This experiment can help to confirm the minimum cell density of neutrophils to be used for immunoblotting and might help to improve the sensitivity of the Hax1 detection by these antibodies.

3. The authors should specify the detailed composition of the Laemmli sample buffer as the composition of this buffer varies slightly from lab to lab. Some research labs add 2M-4M urea in the Laemmli buffer for complete solubilization of membrane bound- hydrophobic proteins that have a tendency to precipitate in SDS when heated at 95°C. If urea plus SDS are used for sample solubilization, samples are incubated at room temperature for 30 minutes [as heating causes urea to break down].

4. Have the authors tried immunoblotting with the pre-immune serum to make sure that these antibodies do not have any non-specific interactions? The authors should make this clear in the manuscript.

5. The authors should clearly state the duration of the secondary antibody incubation which is missing in the manuscript.

6. The authors state that the background protein bands are due to the goat anti- rabbit IgG secondary antibody (Figure 5). But there are background bands with the goat anti-mouse IgG secondary antibody (Figure 5). Have the authors performed the same experiment with the goat-anti-mouse secondary antibody? Is the background due to the cell line used in the experiment or due to the specific batch of the antibody? It would be nice to discuss this issue in the discussion section of the manuscript.

7. No explanation was given for the observation why the control shRNA cells show more intense staining intensity than the wild type cells. This should be addressed in the discussion.

8. Figure 3B does not clearly state how many replicates were used for the quantification of the band intensities.

9. The authors should quantify the Hax1 knockdown in the Hax1 shRNA cell line (Figure 4). Without quantification, it is hard to accurately distinguish the specific Hax1 reduction from the background non-specific weak binding of antibodies.

**Competing Interests:** No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.
Thank you for your kind review of our manuscript. Below I have addressed your comments:

1. We have clarified in the abstract, introduction, and discussion that these antibodies are commercially available.

2. We have performed these experiments using gels that have not had the stacking gel removed and we do not observe any noticeable signal in the stacking gel. Furthermore, Gallagher et al. use a 1% TX-100 lysis buffer in their experiments similar to our lysis conditions. It would be interesting to purify the nuclear extract and compare to the cytosolic fractions to determine this further.

3. The composition of the Laemmli sample buffer has now been added to Table 1.

4. Thank you for this suggestion. Pre-immune sera experiments have been added to figure 5. In short we find that there is increased background using the pre-immune sera that we don't typically see. This demonstrates that the commercially available antibodies are relatively clean after the purification process.

5. The 1 hour incubation of the secondary antibody has been added to the manuscript.

6. The apparent background bands noticed in the mouse channel was a mistake on our part. For many of the figures the rabbit and mouse channels were overlays. This has been fully corrected and every figure now contains separate blots for the mouse and rabbit channels. As can be seen the background is only observed in the rabbit channel.

7. We have repeated these experiments and on average we do not see any significant increase in band intensities between the wild-type PLB-985 cells and the control shRNA cells (see Figure 5C). This could have been variation due to loading in the past figures. We hope the new figures are more clear in this regard.

8. Figure 3B is quantification from three independent replicates. This has been clarified in the figure legend, and the included dataset 1 contains the raw numerical information.

9. This has been completed. Quantifying the knockdown from the 0.1 x 10^6 and 0.5 x 10^6 cell samples was highly variable due to the low level of protein available, however at the 1 x 10^6 cell density quantification was reliable and both antibodies showed a relatively equal level of knockdown.

**Competing Interests:** No competing interests were disclosed.
The work presented by Kristina and Cavnar presents data validating two commercial antibodies raised against Hax1. Their experimental approach is well explained and there is a good description of the methods and reagents used. The data is also convincing and well presented, I particularly liked the fact that complete blots are shown and knockdown used to show specificity. Overall, it is a very strong example of an antibody validation paper.

I believe the manuscript is suitable for indexing subject to a few fairly minor changes, which are detailed below:

1. I would make it clear that the antibodies are commercially available in the abstract.

2. It would be worth double checking that details are provided for all of the western blotting reagents and methods. There is plenty of evidence that small changes can alter antibody behaviour and it really helps those trying to reproduce the work. One thing I noticed is that I don't think the contents of Laemmli loading buffer are spelt out, does it have reducing agent in it? Another example is how long were secondaries incubated for?

3. Figure 3- Is the quantification based on one or more experiments- this should be clear in the legend.

4. Dataset 1- should be “plotted” and not “graphed”.

5. I was interested in the background which appears to be caused by the secondary antibodies. The authors state it is due to the anti-rabbit antibody, but it seems there is at least some with the anti-mouse staining? I would be interested if this is common with this cell line or is it this set of secondaries- I think this would be worth briefly mentioning in the discussion.

6. The authors should present the quantification of the knockdown (figure 4). If both antibodies are specific then you would predict that they will show roughly similar levels of reduction in signal. This would suggest that the remaining signal is caused by a lack of knockdown rather than low levels of non-specific staining.

**Competing Interests:** No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Author Response 03 Aug 2015

**Peter Cavnar**, University of West Florida, USA

Thank you for the kind review of our manuscript. We have addressed your comments below:

1. Commercial availability of the antibodies has now been included in the abstract, introduction, and discussion.

2. We have now included the composition of the Laemmli loading buffer and also our transfer buffer composition to Table 1.

3. Figure 3 includes quantification from three independent experiments.
4. All references to datasets and graphs have been changed to "plotted".

5. This was a mistake on our part in our first submission the two channels were overlays. This has been corrected and all blots have been separated into their subsequent mouse and rabbit channels. We hope it is clear now that the background bands we see are due to the goat anti-rabbit 680 secondary antibody with no detectable background in the mouse channel.

6. This was a great suggestion and has been completed. The quantification is highly variable at low cell densities, however at 1 x 10^6 cells the level of the Hax1 KD is relatively constant and significant using the mouse or rabbit Hax1 antibodies. This demonstrates the specificity of the Hax1 antibodies used in this study.

**Competing Interests:** No competing interests were disclosed.

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