Structure and dynamics of the membrane attaching nitric oxide transporter nitrophorin 7 [version 2; referees: 2 approved, 1 approved with reservations]

Markus Knipp¹, Hideaki Ogata¹, Giancarlo Soavi⁶, Giulio Cerullo⁶, Alessandro Allegri², Stefania Abbuzzetti³, Stefano Bruno⁵, Cristiano Viappiani²,⁴, Axel Bidon-Chanal⁷, F. Javier Luque⁷

¹Max-Planck-Institut für Chemische Energiekonversion, Mülheim an der Ruhr, 45470, Germany
²Dipartimento di Fisica e Scienze della Terra, Università di Parma, Parma, 43124, Italy
³Dipartimento di Bioscienze, Università di Parma, Parma, 43124, Italy
⁴NEST, Istituto Nanoscienze, Consiglio Nazionale delle Ricerche, Pisa, 56127, Italy
⁵Dipartimento di Farmacia, Università di Parma, Parma, 43124, Italy
⁶Dipartimento di Fisica, Politecnico di Milano, Milano, 20133, Italy
⁷Departament de Fisicoquímica, Facultat de Farmàcia and Institute of Biomedicine, Universitat de Barcelona, Santa Coloma de Gramenet, E-08921, Spain

First published: 13 Feb 2015, 4:45 (doi: 10.12688/f1000research.6060.1)

Abstract
Nitrophorins represent a unique class of heme proteins that are able to perform the delicate transportation and release of the free-radical gaseous messenger nitric oxide (NO) in a pH-triggered manner. Besides its ability to bind to phospholipid membranes, the N-terminus of NP7, a member of the NO transporter nitrophorin family, contains an additional Leu-Pro-Gly stretch, which is a unique sequence trait, and the heme cavity is significantly altered with respect to other nitrophorins. These distinctive features encouraged us to solve the X-ray crystallographic structures of NP7 at low and high pH and bound with different heme ligands (nitric oxide, histamine, imidazole). The overall fold of the lipocalin motif is well preserved in the different X-ray structures and resembles the fold of other nitrophorins. However, a chain-like arrangement in the crystal lattice due to a number of head-to-tail electrostatic stabilizing interactions is found in NP7. Furthermore, the X-ray structures also reveal ligand-dependent changes in the orientation of the heme, as well as in specific interactions between the A-B and G-H loops, which are considered to be relevant for the biological function of nitrophorins. Fast and ultrafast laser triggered ligand rebinding experiments demonstrate the pH-dependent ligand migration within the cavities and the exit route. Finally, the topological distribution of pockets located around the heme as well as from inner cavities present at the rear of the protein provides a distinctive feature in NP7, so that while a loop gated exit mechanism to the solvent has been proposed for most nitrophorins, a more complex mechanism that involves several interconnected gas hosting cavities is proposed for NP7.

Open Peer Review

Referee Status: ✔ ✔ ☑

Invited Referees
1
2
3

REvised
version 2
published 18 Aug 2015

version 1
published 13 Feb 2015

Report ✔ ✔ ☑

1 Marten H. Vos, Ecole Polytechnique France
2 Suman Kundu, University of Delhi India
3 Antonio Cupane, University of Palermo Italy, Matteo Levantino, University of Palermo Italy

Discuss this article

Comments (0)
This article is included in the **Oxygen-binding and sensing proteins** collection.

**Corresponding author:** Cristiano Viappiani (cristiano.viappiani@unipr.it)

**Competing interests:** No competing interests were disclosed.

**How to cite this article:** Knipp M, Ogata H, Soavi G et al. Structure and dynamics of the membrane attaching nitric oxide transporter nitrophorin 7 [version 2; referees: 2 approved, 1 approved with reservations] *F1000Research* 2015, 4:45 (doi: 10.12688/f1000research.6060.2)

**Copyright:** © 2015 Knipp M et al. This is an open access article distributed under the terms of the Creative Commons Attribution Licence, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited. Data associated with the article are available under the terms of the Creative Commons Zero "No rights reserved" data waiver (CC0 1.0 Public domain dedication).

**Grant information:** This work was financially supported by the Deutsche Forschungsgemeinschaft (DFG) grant KN951/1-1, the Max Planck Society (both to M.K.), Spanish Ministerio de Economía y Competitividad (SAF2011-27642; FJL), Generalitat de Catalunya (2014SGR1189; FJL), Icrea Academia (FJL). The Consorci de Serveis Universitaris de Catalunya and the Barcelona Supercomputer Center are acknowledged for computational facilities.

*The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.*

**First published:** 13 Feb 2015, 4:45 (doi: 10.12688/f1000research.6060.1)
Amendments from Version 1

We have modified the manuscript by taking into account the suggestions and the issues raised by the reviewers.

- Specifically, we have rewritten the statement about the sequence identity, which seemed to be confusing in the original version of the manuscript. We have also expanded Table 2 to include the sequence identity with NP3.

- We believe that both crystal packing and electrostatic interactions impose a severe constraint on the AB loop, limiting the conformational flexibility of this region, and this likely explains why there seems to be no apparent major difference between the structures at pH 5.8 and 7.8. These comments have been introduced in the revised version of the manuscript.

- The section dealing with the data analysis was modified as follows. We have expanded the discussion of the inaccuracy in the determination of rate constants that may derive from the limited precision with which ultrafast and nanosecond kinetics are merged. A comment on the problem of the large number of fitting parameters was also introduced.

- We have explicitly indicated the hypothetical association of reaction intermediates $T_a$, $T_p$, and $T_f$ with the cavities identified by molecular modeling and indicated by colored isocontours in Figure 10.

- The discussion about rate constants has been expanded to include relevant cases from the literature.

- The last section with concluding remarks has been rewritten to better discuss the role of the protein as an NO reservoir.

See referee reports

Introduction

Nitrophorins (NPs) comprise a family of heme binding proteins that originate from the saliva of the *Rhodnius* species of Triatomid bugs [1,2]. These insects use the heme iron as an anchor to store nitric oxide (NO), which is released during feeding on a host while the saliva is continuously pumped into the host tissue [3]. NO assists the feeding process by inhibition of blood coagulation and increase of blood vessel diameter. The heme cofactor may derive from the limited precision with which ultrafast laser-flash photolysis experiments, which allowed to characterize the ligand migration and binding in NP7 from the 10^{-12} to the 10^{-3} time-scale.

Materials and methods

Expression of NP7

The protein was expressed, reconstituted, and purified as was described previously [4,7]. The extended purification by cation exchange chromatography using a Ca^{2+}-charged chelating Sepharose (GE Healthcare) [18] was essential for the successful crystallization. Protein preparations were judged by SDS-PAGE to be >95% pure. MALDI TOF MS (Voyager DE Pro, Applied Biosystems) confirms the correct molecular masses including an initial Met-0 residue and accounting for two Cys–Cys disulfides (calculated for [NP7 + H]^+: 20,969 Da, observed: 20,966 ± 20 Da). Proteins were kept at −20°C in 200 mM NaOAc/HOAc (Carl Roth), 10% (v/v) glycerol (Carl Roth) (pH 5.0) until use.

X-ray crystallography

Protein crystals were obtained from 10 mg/mL NP7 in 10 mM NaOAc/HOAc (pH 5.5) using the vapor-diffusion method [2,14] upon mixing with an equal volume of crystallization solution (Hampson research) as indicated in Table 1. The crystals were soaked for 10 min on ice in the respective crystallization solution plus 15% glycerol as a cryo-protectant. Where needed, heme ligands were added into the cryo-protectant solution. Afterwards, the crystals were immediately frozen in liquid nitrogen and kept there until the measurement. Diffraction data sets were collected at 100 K using the beamlines, BL14.2 at BESSYII (Helmholtz-Zentrum Berlin, Germany) and PXII at SLS (Villigen, Switzerland). The data set was processed with XDS [18] and CCP4 [19]. The molecular-replacement method was applied using M oL R [20] and an initial model from NP4 (PDB code 3MVF) [7]. Model building and refinement were carried out using W eN C oot [17] and P hENIX [22], respectively. Data collection and refinement statistics are summarized in Table 1. PHENIX was used to check the stereochemical properties.
Table 1. Refinement statistics of the crystal structures of NP7 obtained with various heme ligands and under various crystallization conditions. The highest resolution shell is shown in parenthesis.

<table>
<thead>
<tr>
<th>heme ligand</th>
<th>H2O a</th>
<th>H2O b</th>
<th>NO c</th>
<th>Hm d</th>
<th>ImH e</th>
<th>Gly–Gly–Gly c</th>
</tr>
</thead>
<tbody>
<tr>
<td>pH</td>
<td>5.8</td>
<td>7.8</td>
<td>7.8</td>
<td>7.8</td>
<td>7.8</td>
<td>7.8</td>
</tr>
<tr>
<td>PDB code</td>
<td>4XMC</td>
<td>4XMD</td>
<td>4XME</td>
<td>4XMF</td>
<td>4XMG</td>
<td>4XMH</td>
</tr>
</tbody>
</table>

**Data collection**

| X-ray source | Wavelength (Å) | Space group | Unit-cell parameters | Resolution (Å) | No. of observed reflections | No. of unique reflections | Rmerge (%) | Completeness (%) | <|I|/|σ|I|> |
|--------------|----------------|-------------|----------------------|----------------|----------------------------|--------------------------|------------|-----------------|-----------------|
| SLS          | 0.9999         | P2 1        | 38.42, 67.00, 39.00, 116.6 | 34.88-1.42     | 106086                     | 59987                    | 0.033 (0.646) | 91.3 (71.4)    | 11.0 (1.4)     |
| PXII         | 0.91841        | P2 1        | 38.31, 66.81, 39.03, 116.9 | 30.42-1.60     | 86820                      | 23143                    | 0.040 (0.486) | 99.4 (99.8)    | 20.5 (3.0)     |
| BESSYII      | 0.9999         | P2 1        | 38.89, 67.05, 39.08, 117.3 | 34.71-1.29     | 140029                     | 83829                    | 0.032 (0.925) | 94.0 (84.0)    | 9.5 (1.0)      |
| BL14.2       | 0.91841        | P2 1        | 38.34, 66.72, 39.83, 116.9 | 30.74-1.60     | 86063                      | 22417                    | 0.067 (0.780) | 96.9 (95.5)    | 15.3 (1.9)     |
| SLS          | 0.800010       | P2 1        | 38.26, 66.75, 38.56, 116.5 | 34.25-1.80     | 52612                      | 15836                    | 0.083 (0.439) | 97.6 (96.8)    | 9.2 (3.0)      |
| PXII         | 0.99999        | P2 1        | 38.37, 66.93, 38.89, 116.8 | 34.71-1.32     | 126311                     | 41095                    | 0.024 (0.290) | 92.8 (64.7)    | 21.4 (2.3)     |

The crystallization conditions (all reagents from Hampton Research) were:

a) 25%(w/v) PEG 3350, 0.1M MES monohydrate pH 5.8, 0.25%(w/v) Gly-Gly, 0.25%(w/v) Gly-Gly-Gly, 0.25%(w/v) pentaglycine, 0.02 M HEPES sodium pH 6.8.

b) 25%(w/v) PEG 3350, 0.1M bis-Tris propane pH 7.8, 0.02 M HEPES pH 6.8, 0.25%(w/v) naphthalene-1,3,6-trisulfonic acid trisodium salt hydrate, 0.25%(w/v) 2,6-naphthalenedisulfonic acid disodium salt, 0.25%(w/v) 4-aminobenzoic acid, 0.25%(w/v) 5-sulfosalicylic acid dehydrate.

c) 25%(w/v) PEG 3350, 0.1M bis-Tris propane pH 7.8, 0.25%(w/v) Gly-Gly, 0.25%(w/v) Gly-Gly-Gly, 0.25%(w/v) hexamminecobalt(III) chloride, 0.25%(w/v) salicylamide, 0.25%(w/v) salicylamide, 0.25%(w/v) vanilic acid, 0.02 M HEPES sodium pH 6.8.
Ultrafast spectroscopy
Samples for kinetic investigation were prepared by equilibrating 80 µM NP solutions in a sealed 0.2×1 cm-quartz cuvette connected to a tonometer with 0.1 or 1 atm CO. Na₂S₂O₃ was then titrated anaerobically into the solution while formation of the CO adduct was monitored by absorption spectroscopy. The solubility of CO in water was such that its concentration was 1.05 mM at 10°C when the solution was equilibrated with 1 atm CO. The temperature dependence of CO solubility was taken into account in the numerical analysis. The numerical analysis of the rebinding kinetics was described in detail previously21,22.

The experimental setup used for the pump-probe studies has been described23. The sample was excited with a 70-fs, -10 nm bandwidth pump pulse centered at 530 nm obtained by a home-made optical parametric amplifier, driven by a regeneratively amplified Ti:sapphire laser at 1 kHz repetition rate. The probe pulse was a broadband single-filament white-light continuum generated in CaF₂ covering a spectral range from 350 nm to 700 nm approximately. The pump pulse passed through a delay line and was then overlapped with the probe beam on the sample. The transmitted probe light was dispersed on an optical multichannel analyzer equipped with fast electronics, allowing single-shot recording of the probe spectrum at the full 1 kHz repetition rate. By changing the pump-probe delay we recorded 2D maps of the differential absorption (ΔA), as a function of probe wavelength and delay. The temporal resolution was determined to be approximately 150 fs and the sensitivity, for each probe wavelength, was better than 10⁴.

Nanosecond laser flash photolysis
The experimental setup was described in detail elsewhere23. Photolysis of CO complexes was obtained using the second harmonic (532 nm) of a Q-switched Nd:YAG laser (Surelite-Continuum II-10) at 10 Hz repetition rate. We used a cw output of a 75 W Xe arc lamp as probe beam, a 5-stages photomultiplier (Applied Photophysics) for detection and a digital oscilloscope (LeCroy LT374, 500 MHz, 4 GS s⁻¹) for digitizing the voltage signal. A spectrograph (MS257 Lot-Oriel) was used to select the monitoring wavelength (436 nm) and to remove the stray light from the pump laser. The overall temporal resolution was determined to be about 10 ns and the sensitivity was better than 10⁴. The sample holder was accurately temperature-controlled with a Peltier element, allowing a temperature stability of better than 0.1°C. Experiments were performed at 20°C.

Data analysis
The analysis of the entire CO rebinding kinetics required the use of a detailed kinetic scheme (see section Ligand rebinding kinetics for details) in order to estimate the microscopic rate constants. The differential equations associated with the kinetic schemes were solved numerically, and the rate constants were optimized to describe simultaneously the experimental data at two different CO concentrations. Numerical solutions to set coupled differential equations associated with reaction schemes were determined by using the function ODE15s within Matlab 7.0 (The MathWorks, Inc., Natick, MA). Fitting of the numerical solution to experimental data was obtained with a Matlab version of the optimization package Minuit (CERN).

Spectra acquired in the fs-ps time scale were analyzed by singular value decomposition (SVD), performed using the software Matlab. Our data matrix D, consists of differential absorption measured as a function of two variables: the wavelength of the probe beam and the time delay between the pump and the probe pulses. The singular value decomposition of D can be written as:

\[ D = USV^T \]  
(1)

The meaning of the symbols is the following: columns of matrix U are a set of linearly independent, orthonormal basis spectra; columns of V represent the amplitudes of these basis spectra as a function of time; V^T is the transpose of matrix V; matrix S is a diagonal matrix of non-negative singular values that give the extent of the contributions of each of the products of the i-th column vectors U_iV_i to the data matrix D. The selected components can be further screened by evaluating the autocorrelation of the corresponding columns of U and V, rejecting the component if the autocorrelation falls below 0.8²⁴.

MD simulations
MD simulations were used to explore the conformational space of wild type NP7 and its NP7(Δ1–3) variant, specifically regarding the structural flexibility of the N-terminus and the loops that shape the mouth of the heme cavity. The molecular systems were modeled from the X-ray structures of NO-bound NP4 at pH 5.6 and 7.4 (PDB ID: 1X80, resolution of 1.0 Å; 1X8N; resolution of 1.1 Å)³⁵ as representative systems at low and high pH. Comparison with the X-ray structure showed a remarkable agreement for the protein backbone (RMSD close to 1.7 Å), especially after exclusion of the residues located in A-B and G-H loops, which can be ascribed to the contacts due to the head-to-tail arrangement found in the crystal lattice (see below). Following previous studies³⁶–³⁸, standard ionization states were assigned to all the residues with the only exception of Asp32, which was protonated at low pH and ionized at high pH. The two native disulfide bridges were also imposed between residues Cys5 and Cys124 and between Cys42 and Cys173³⁹. Finally, the heme was modeled in the A orientation, which was shown to be the thermodynamically favored orientation for both NP7 and Met-NP7(Δ1–3)³⁹,⁴⁰.

Simulations were run using the Charmm21 force field and the NAMD program⁴¹. The protein was immersed in a pre-equilibrated cubic box (~70 Å per side) of TIP3P water molecules. Bonds involving hydrogen atoms were constrained at their equilibrium length using SHAKE and SETTLE algorithms, in conjunction with a 2 fs time step for the integration of the Newton’s equations. Trajectories were collected in the NPT (1 atm, 300 K) ensemble using periodic boundary conditions and Ewald sums (grid spacing of 1 Å) for long-range electrostatic interactions. A multistep protocol was used to minimize and equilibrate the system. Thus, the energy minimization was first performed for the hydrogen atoms, then water molecules, and finally the whole system. The equilibration was performed by heating from 100 to 298 K in four 200-ps steps at constant volume. Then, a 200 ps MD at constant pressure and temperature was run. The final structure was used for the production MD runs, which covered 100 ns. Frames were collected at 1 ps intervals, which were subsequently used to analyze the trajectories.
Results and discussion
X-Ray crystallography

Conditions for the crystallization of NP7 at pH 7.8 were previously reported. The addition of di- and polyanionic substances to compensate for the positively charged surface of NP7 that is responsible for the binding to negatively charged phospholipid membranes was crucial for the crystal formation. The previously reported crystals diffracted to a resolution of 1.8 Å. Further optimization of the crystallization conditions resulted in crystals that diffract to even higher resolution down to 1.29 Å (Table 1). It was also possible to crystallize the protein at low pH conditions, i.e., pH 5.8. Since charge compensation for the Lys surface patch was crucial, we also tried to add the amine coordinating bisphosphonate (“lysine tweezer”) 1 (Scheme 1), which is known to be a Lys-specific binder, resulting in high resolution crystals. Another additive that turned out to be successful was the Gly–Gly–Gly tripeptide.

To obtain different iron-ligated structures, the crystals were soaked with imidazole (ImH), histamine (Hm), or the NO donating compound DEA/NONOate for 10 min at room temperature. Upon incubation with the mother liquor containing 15% of glycerol as a cryo-protectant, crystals were frozen in liquid nitrogen. Diffraction experiments were carried out at two different synchrotrons at 100 K. All crystals occupied the space group $P2_1$. Refinement of the crystal structures was obtained through the molecular replacement method using the structure of NP4 (PDB code 3MVF) as a template. The refinement statistics are summarized in Table 1. It should be mentioned that no electron density of the additives (see Table 1 footnotes) except for the Gly–Gly–Gly tripeptide was observed.

Protein fold

The overall fold of NP7 resembles that of other NPs. Eight anti-parallel $\beta$-strands (A to H) form a barrel that hosts the heme cofactor including the ligation by the proximal His60 residue. The structural identity reflected by the RMSD values obtained from the comparison of the backbone atoms of two of the isoforms correlates with the amino acid sequence identities (Table 2). Figure 1 displays the overall fold compared to those of NP2 and NP4. The core of the lipocalin fold is well superposed in all cases. The position of the two disulfide bridges, which is a common trait of the NPs, is very similar among all the NPs. The largest differences are found in the A-B, B-C and G-H loops. In detail, the bending of the $\beta$-strands ($\beta_8$ and $\beta_9$) is significantly more similar in the case of NP7 and NP2 compared to NP4. This is also true for the AB-loop. On the other hand, significant differences are found in the spatial arrangement of the G-H loop, which is markedly bent in NP7 compared to NP2 and NP4 (Figure 1).

![Scheme 1](image)

Table 2. Comparison of the RMSD values (Å; upper triangle) and amino acid sequence identities (%) of NP1, 2, 3, 4, and 7.

<table>
<thead>
<tr>
<th></th>
<th>NP1</th>
<th>NP2</th>
<th>NP3</th>
<th>NP4</th>
<th>7°</th>
</tr>
</thead>
<tbody>
<tr>
<td>NP1</td>
<td></td>
<td></td>
<td>0.79</td>
<td>1.54</td>
<td></td>
</tr>
<tr>
<td>NP2</td>
<td>45</td>
<td></td>
<td>1.51</td>
<td>1.26</td>
<td></td>
</tr>
<tr>
<td>NP3</td>
<td>44</td>
<td>81</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>NP4</td>
<td>88</td>
<td>44</td>
<td>42</td>
<td>1.63</td>
<td></td>
</tr>
<tr>
<td>NP7</td>
<td>44</td>
<td>60</td>
<td>60</td>
<td>41</td>
<td></td>
</tr>
</tbody>
</table>

In the crystal structure of NP7[ImH] the electron density map clearly reflects the presence of heme A orientation, as expected for NP7 (see above). However, to our surprise, the NP7 structure that crystallized with Gly–Gly–Gly has heme B orientation (Table 1 and Figure 4). Thus, it may be concluded that the insertion of this ligand leads to a widening of the heme pocket, which may allow the heme to turn. Since the structures of NP7[NO], NP7[Hm] and unliganded NP7 at pH 5.8 were derived from crystal soaking and displacement of the Gly–Gly–Gly ligand, the heme consequently was found in the heme B orientation as the crystal lattice does not allow the cofactor to turn. However for NP7[NO], A orientation was demonstrated by circular dichroism spectroscopy. Caution is needed regarding the presence of B orientation in unliganded NP7 at pH 7.8, since it could not be properly assigned due to the loosely occupied heme (Table 1).

Protein surface

One of the most interesting features of NP7 is its ability to bind to negatively charged membranes. The crystal structures demonstrate the extensive clustering of Lys side-chains at the protein surface opposite the heme pocket that accomplishes the strong protein-membrane interaction. On the other hand, the side of the heme mouth has a negative charge potential leading to a total bipolar charge distribution. It was previously noticed that NP7 tends to aggregate at elevated concentrations, which can be explained by a charge stabilized aggregation process. X-ray crystallography indeed supports such a head-to-tail interaction, which involves a variety of salt bridges (Figure S1), leading to chain-like arrangement in the crystal lattice (Figure 2a).
Figure 1. Representation of the backbone structure of X-ray NP7 structures. (a) Overall fold of NP7. (b) Overall fold of NP7 (green) in comparison to NP2 (blue) (PDB code, 2A3F) and NP4 (orange) (PDB code, 1YWD).

Figure 2. Head-to-tail arrangement of NP7 in the crystal lattice. (a) Arrangement of molecules in the crystal lattice of NP7. (b) Crystal contact between two neighboring molecules in the NP7 crystal. (c) The electrostatic potential of NP7. The red and blue colors show the negative and positive potentials, respectively.

Figure 3. Heme b nomenclature and heme orientation inside the pocket of NP7. The heme pocket of NP7 is indicated by the three green circles corresponding to the position of the three aliphatic side-chains pointing from the top of the distal pocket onto the heme plane. The two possible heme orientations are indicated as A and B.
In the electron density map derived from the diffraction pattern of the unliganded NP7 at pH 7.8 the heme was very poorly defined, while the rest of the structure was very well defined (Figure 4b). In contrast, the electron density of the cofactor bound to ImH, Hm and NO was well defined (Figures 4c–e). Since the ferriheme iron undergoes rapid photo reduction during the recording of diffraction patterns\(^{40–43}\), the crystal structures display the Fe\(^{II}\) state. We had previously reported that the Fe\(^{II}\)–N\(_{His60}\) bond in NP7 is surprisingly weak, so that the reduction of NP7 leads to the equilibrium noted in Equation 2 at neutral pH (pK\(_a\) = 7.8)\(^{44}\).

\[
\text{H}_2\text{O} + \text{Fe}^{II}(\text{ppIX})-\text{N}_{\text{His60}} \rightleftharpoons \text{H}_2\text{O} - \text{Fe}^{II}(\text{ppIX}) + \text{N}_{\text{His60}} \quad (2)
\]

Overall, these findings suggest that the ill-defined heme density of the unliganded NP7 reflects the movement of the cofactor inside the heme pocket according to the two coordination states. The binding of the π donors ImH, Hm or CO has a positive \textit{trans} effect, which stabilizes the Fe\(^{II}\)–N\(_{His60}\) bond, so that the heme resides in the pocket. Interestingly, NO has a negative \textit{trans} effect due to the overlap of its anti-bonding \(\pi^*\) orbital with the iron \(d_z^2\) orbital, thus weakening the Fe–N\(_{His}\) σ bond\(^{45–47}\).

The crystal structures allow detailed examination of Glu27 (Figure 5a). The side-chain is folded away from the hydrophobic heme side toward the interior of the structure, where it is involved in a network of H-bonding contacts. This includes a single water that is further coordinated to Tyr175 near the protein surface. It was previously found that the mutation Glu27→Gln has a remarkable destabilizing effect on the NP7 fold\(^{30}\). It can now be understood that the negative charge of Glu27 attracts the Tyr175 hydroxyl group, which forms an important interaction. On the other hand, the dense packing of side-chains next to His60 was identified as another destabilizing factor of the Fe\(^{II}\)–N\(_{His60}\) bond, where Phe43 plays a crucial role\(^{44}\). Figure 5b shows the arrangement in comparison to...
the crystal structures of NP2 and NP4. Phe43 is oriented parallel to the heme plane with a distance of 3.5 Å leading to π-stacking between the two aromatic rings. Moreover, the phenyl ring is perpendicularly oriented toward the His60 plane with a distance of 3.6 Å. The distance between Glu27:Cβ and Phe43:Cβ is 4.0 Å, while Glu27 also H-bonds to Phe43:NH.

**Binding of NO**

In the crystal structure, NO (occupancy 0.44) is bound with \( \angle(\text{Fe–N–O}) = 124^\circ \), which corresponds to a reduced iron, i.e. Fe(II)(NO). In the case of NP1[NO] a similar angle of 123° to 135° was reported43. For NP4[NO] two orientations of 110° and 177° were assumed because the electron density could not be sufficiently fit with a single conformation49. The data were interpreted by the parallel co-existence of photoreduced Fe(II)(NO) and some residual Fe(III)(NO) in the frozen crystal. However, according to our41 and other10,43 work, photoreduction occurs even in frozen crystals within the first seconds of data collection, thus the presence of significant amounts of NP4[Fe(III)NO] seems very unlikely. Also in the case of NP7[NO] some of the electron density could not be sufficiently fit with the presence of a single NO configuration (Figure 4c). However, this may reflect an incomplete occupancy of the axial Fe site with NO and displacement with water since the protein crystals were prepared and transported in the Fe(III)(NO) form to the synchrotron, whereby loss of NO gas is possible.

**Binding of imidazole and histamine**

Similar to other NPs, the binding of Hm is accomplished not only through the coordination of heme iron, but also through the salt bridge of its ethylamine group with the Asp32 carboxylate (Figure 6a). When ImH is bound, the missing ethylamine is compensated by H-bonding to a water molecule, which is then coordinated by Asp32 (Figure 6b). However, in marked contrast to other NPs, the affinity constant for ImH and Hm was markedly decreased. While for NP2 the equilibrium constant \( (K_{eq}) \) was found to be \( 2.5 \times 10^7 \) M\(^{-1}\), for NP7 \( K_{eq} \) was \( 1.0 \times 10^6 \) M\(^{-1}\). This difference is even more pronounced for Hm, as the equilibrium constants are \( 1.0 \times 10^8 \) M\(^{-1}\) and \( 1.0 \times 10^4 \) M\(^{-1}\) for NP2 and NP7 [8], respectively. The remarkable difference observed between the two ligands in NP7 as well as between NP2[Hm] and NP7[Hm] is not explained by the structures. However, taking into account that the N-terminus is expected to move into the space between the A-B and G-H loops, the disruption of the Asp32-Hm interaction is feasible. This is further supported by the finding that deletion of the N-terminal residues increases the affinity for Hm \( (K_{eq}(NP7(\Delta 1-3)) = 1.3 \times 10^7 \) M\(^{-1}\))\(^{50}\). This trend is also found in the enhanced affinity for ImH \( (K_{eq}(NP7(\Delta 1-3)) = 3.2 \times 10^7 \) M\(^{-1}\))\(^{6}\), which compares with the value reported for NP2 (see above).

**pH dependence and N-terminus**

It was previously suggested that the pH can have significant influence on the A-B and G-H loop conformations in NP4 due to the change in the protonation state of Asp30 (part of the A-B loop)\(^{32,37,31-34}\). While in the protonated state Asp30 binds to the backbone C=O of Leu130 (part of the G-H loop), it is detached upon deprotonation leading to the opening of the heme pocket\(^1\). Based on this mechanism, a model for the pH-dependent NO release under biological conditions was developed. However, very limited pH dependences of the ligand release rates and equilibrium constants leave considerable doubts about this likely oversimplified model\(^{55}\).

In contrast to NP1-4, where the pH dependence of the equilibrium constant is less than an order of magnitude, the difference for NP7 spans three orders of magnitude\(^{5}\). However, no significant conformational differences were found between the X-ray structures collected at two different pHs (5.8 and 7.8; RMSD = 0.22 Å). This finding likely reflects the reduced conformational flexibility of the AB loop imposed by both crystal packing and especially by the electrostatic interactions favored by the head-to-tail arrangement of the proteins in the crystallographic structure. On the other hand, in all NP7 structures Asp32 is bound to a water that is held by

---

**Figure 6. Local structural details of the heme pocket.** (a) Hm, (b) ImH, (c) pH 5.8 and (d) pH 7.8. The numbers represent the bond distances (Å).
the backbone NH and C=O of Asp134, that is, the Asp32-Ile132 H-bond that corresponds to the Asp30-Leu130 H-bond in NP4 is not observed (Figures 6c and 6d). Thus, there are significant differences in the key structural features during the transition between closed and open forms of the protein. However, it is unclear whether these differences may be attributed exclusively to the head-to-tail arrangement (see above and Figure 2), or alternatively may also be influenced to some extent by the distinct N-terminus stretch of NP7, which fills the region between A-B and G-H loops (Figure S2).

Upon maturation, a major difference of NP7 compared to all other NPs is its extended N-terminus (Figure S3), which is characterized by the extension of the tripeptide Leu1-Pro2-Gly3. It should be mentioned that the N-terminus of the recombinant NP7 contains an additional Met0 residue originating from the start codon of the expression system. In the X-ray structures, the residues Met0 and Leu1 were not seen. As noted above, the crystal lattice involves a crystal contact between the A-B and G-H loops of one molecule and the positive patch of the back side of another molecule (Figure 2b), where the sharp E-F loop Lys116 plays a major role, contributing to the head-to-tail arrangement via salt bridges with residues Asp34 and Asp32 (distances of 3.4 and 5.0 Å, respectively). As a consequence, the N-terminus is rather floppy sitting on top of the structure (Figure S3).

A series of MD simulations were run to examine the conformational flexibility and potential interactions formed by the N-terminus stretch of NP7 in aqueous solution. To this end, MD simulations were carried out for models of the wild type protein representative of the closed and open states, and the results were compared with those obtained for the Δ(1-3) variant to ascertain the structural impact of the three extra residues. Furthermore, in order to avoid an artifactual bias due to packing effects, models of wild type and Δ(1-3) NP7 proteins were built up using the X-ray crystallographic structures of NO bound NP4 at pH 5.6 and 7.4, because the conformations of the AB and GH loops are characteristic of the closed and open states, respectively. All the simulations led to stable trajectories, as noted in RMSD values comprised between 0.9 and 1.3 Å (data not shown).

The behavior observed for the open forms of wild type NP7 and its Δ(1-3) variant is highly similar, mainly reflecting significant fluctuations of the A-B loop irrespective of the length of the N-terminus (Figure 7a), which for NP7 fills the region between A-B and G-H loops, thus resembling the arrangement found in the X-ray structures of NP7 (Figure S2). This structural resemblance is reduced in the closed state, because the A-B and G-H loops exhibit larger fluctuations in the wild type protein compared to the Δ(1-3) species (Figure 7b), and this effect is accompanied by the enhanced conformational flexibility of the Leu1-Pro2-Gly3 stretch in NP7, which is in contrast with the more rigid structure adopted by the N-terminus in the Δ(1-3) variant. Interestingly, superposition of the snapshots sampled for wild type and Δ(1-3) proteins reveals a widening of the heme cavity mouth in the wild type protein (Figure 7b), which would suggest a higher probability for the formation of transient pathways that connect the heme cavity with the bulk solvent. This is noted, for instance, in the larger area estimated for the heme cavity mouth for the closed states of the wild type NP7 and its Δ(1-3) variant (Figure S4).

Overall, these findings could explain why deletion of the N-terminal residues increases the affinity for ImH and Hm, an effect that can presumably arise from the more compact nature of the heme cavity and lower fluctuations of the A-B and G-H loops in the Δ(1-3) protein. Finally, the enhanced flexibility of the closed form of wild type NP7 may also explain the larger sensitivity to pH dependence, since the access of water molecules to the heme cavity could facilitate breaking of the H-bond between Asp32 and Ile132, thus favoring deprotonation of Asp32 and hence the transition to the open state.

**Time resolved differential absorption spectra**

The availability of the NP7 structures now allows the combination with the studies of fast dynamics of the ligand-protein interaction. The FeII–CO complex was studied as a model for the isoelectronic FeII–NO because of its slower geminate rebinding rates. The change in absorption spectra of CO bound and unbound heme proteins (Figure S5) allows the determination of time resolved differential absorption spectra after photolysis of the Fe–CO bond. In this study, NP7–CO was pumped with 70-fs laser pulses at 532 nm. Previous

---

*Figure 7. Structural analysis of MD sampled snapshots.* Superposition of the snapshots taken every 10 ns from the MD trajectories sampled for the (a) open and (b) closed states of wild type NP7 and its Δ(1-3) variant. The backbone of the extra Leu-Pro-Gly stretch found in NP7 is shown in magenta, and the backbone of the A-B and G-H loops for wild type and Δ(1-3) NP7 is shown in green and yellow, respectively.
Ligand rebinding kinetics

We have previously reported the CO rebinding kinetics to NP7 after nanosecond laser photoysis, showing that the binding reaction can be followed by the absorbance change at 436 nm\(^{-1}\). In this work we now aim at reconstructing the full time course of CO rebinding, from a few picoseconds to reaction completion, occurring on the millisecond time scale. We have accomplished this by merging the CO rebinding kinetics determined in the femtosecond pump-probe and in the nanosecond laser flash photoysis experiments.

Due to inherent limitations of the methods employed to collect the kinetics in the two time regimes, the kinetics in the time interval between ~2 ns (the longest available delay in the pump-probe experiment) and 20 ns (the shortest time accessible without distortion due to the instrumental response function of the nanosecond laser flash photoysis setup) is not available. Thus, a connection of the two data sets is not straightforward. In order to obtain a single kinetic trace, we have first normalized the amplitude \(V_t\) retrieved from SVD analysis of the pump and probe experiment, taking advantage of the fact that the fraction of photoproduct with respect to that initially generated at time \(t_p\), i.e., \(N(t) = \Delta A(t)/\Delta A(t_p)\), (with \(t > t_p\)), decreases with time from unity. Unfortunately, it is not possible to extend this procedure to the nanosecond photolysis data since multiple photolysis events occur during the laser pulse under our experimental conditions, thus impairing a correct determination of the fraction of unliganded species surviving at 20 ns. We have therefore estimated the concentration of the unliganded molecules at the end of the temporal window of pump and probe experiments by fitting the kinetics in this time window with a double exponential decay function, and extrapolating the transient signal beyond 2 ns. The flash photolysis data were then scaled in order to match the extrapolated fraction of unliganded molecules. This procedure may admittedly slightly bias the kinetics since the offset extrapolated beyond 2 ns is retrieved with a limited precision, but the effect on microscopic rates is expected to be minor. Simulations demonstrate that an over-/under-estimate of the extrapolated signal from the subnanosecond kinetics by approximately 10%, results in changes of less than 15% in \(k_i\) and much smaller changes in other microscopic rates. The impact on \(k_{ON}\) is less than 5%. We should say that, unlike the method proposed by Champion and coworkers\(^{36}\), the current procedure for merging the two different time ranges has some degree of uncertainty since there is about one log time unit of missing data which may contain kinetic information. Nevertheless, given the shape of the signals, it is expected that this is not a major contribution for the kinetics reported in this work.

Figure 9 shows the overall rebinding curve for NP7 obtained with this method. From a simple inspection of the progress curves, it is easy to notice that for NP7 the sub-ns kinetics becomes faster and larger when the pH is lowered to 5.5. It is comprised of two kinetic phases, which can be well described by a double exponential decay, whose amplitudes and apparent rates increase at pH 5.5 (not shown). The sub-ns rebinding phase is very similar to the previously described CO rebinding to the related NP4\(^{38}\). On the longer time scales, the progress curve for NP7 shows the features previously reported, with a heterogeneous bimolecular rebinding which becomes faster as the pH is lowered to 5.5.
Recently we proposed that the reaction progress for NP7 can be described by a microscopic model which takes into account rebinding of photodissociated ligands from internal cavities that are accessible at room temperature. The current data indicate that in addition to more remote internal cavities, capable of hosting the ligand for a relatively long time, at least one additional temporary docking site exists in the vicinity of the reaction site at the heme, which modulates sub-nanosecond geminate rebinding. From a structural point of view, this assumption is supported by the topological analysis of the inner cavities present in the snapshots sampled along the MD simulation of the closed form of NP7. Thus, Figure 10a shows the three major cavities that shape the tunnel leading from the heme pocket to the back of the protein (shown as magenta isocontour) determined by using the MDpocket tool, and also confirmed by Implicit Ligand Sampling calculations. Besides the inner tunnel, Figure 10a shows the presence of two additional pockets around the heme. Residues Ile121, Ile123, Leu135 and Ser137 shape the first pocket (orange isocontour), and the side chains of Glu27, Phe43, Phe45 and Leu139 contribute to the second pocket (blue isocontour). Hence, it can be expected that rebinding of photolyzed CO would occur from a variety of transient docking sites, reflecting the topological distribution of pockets located close to the heme (these are represented as the orange and magenta isocontours at around ~9 Å from the heme iron in Figure 10) as well as from inner cavities present at the rear of the protein (at around ~22 Å from the heme iron), which can be visited via the inner tunnel present as a distinctive feature in NP7 compared to other NPs.

On the basis of the previous findings, the reaction scheme previously proposed by us is then expanded as shown in Figure 11 to accommodate for transient population (likely located at the orange and magenta isocontours in Figure 10) and T₄ (blue isocontour in Figure 10) of the ligand in docking sites located nearby the distal pocket (DP), and is found to describe well the rebinding kinetics under the investigated experimental conditions. The model was fit to the observed kinetics using a global analysis of progress curves measured at 1 and 0.1 atm CO, in order to increase the reliability of retrieved parameters. Sample fits on CO rebinding kinetics to NP7 are reported in Figure 12.

The microscopic rates reported in Table 3 show that exit to the solvent occurs with a remarkably high rate (9 × 10⁸ s⁻¹ and 8.2 × 10⁸ s⁻¹ at pH 7.5 and 5.5, respectively), which competes, although to a minor extent, with internal rebinding and migration processes. This suggests that the connection between the distal pocket and the solvent must be quite efficient, much more than, for instance, in the case of other proteins with well recognized rather open connections. These include, for instance, myoglobin, for which CO escapes through the His gate at a rate 2.7 × 10⁶ s⁻¹, and type 1 non symbiotic hemoglobin AHb1 for which exit through the His gate...
gate occurs at $5 \times 10^3 \text{s}^{-1}$ [12,63]. This nicely parallels the finding of two large and accessible pathways connecting the distal pocket and the solvent (denoted cage-to-top and cage-to-front in Figure 10b), which can be ascribed to the larger fluctuations of the A-B and G-H loop and widening of the heme cavity in the closed form of wild type NP7 (see Figure 7b).

Inspection of the microscopic rates in Table 3 also reveals that there are some substantial differences with respect to the values previously determined on the sole basis of nanosecond laser photolysis data [33]. Inclusion of the ps rebinding phase results in a major increase in rebinding rates, a fact that is not surprising and was anticipated in our previous analysis. The value of the rebinding rate $k_{j}$ is remarkably large compared to, for instance, rebinding to the highly reactive type II truncated Hb from *Thermobifida fusca* (TrHbO), for which rebinding occurs with a rate of $3 \times 10^9 \text{s}^{-1}$ [148], a value which is 20 fold smaller than the one for NP7. Mutation of distal cavity residues capable of stabilizing water molecules nearby the binding site lowers the barrier for rebinding and increases the rate to $2 \times 10^9 \text{s}^{-1}$, a value which becomes comparable to the value for NP7 at neutral pH, but is still one order of magnitude smaller than observed for NP7 at pH 5.5.

Similarly, CO rebinding to TrHbO from *Mycobacterium tuberculosis* occurs on the picosecond time scale with biexponential kinetics, whose apparent lifetimes are on the same order of magnitude as those observed from *Thermobifida fusca* TrHbO [33]. Molecular dynamics simulations suggest that the presence of water molecules influences the rebinding kinetics. Mutations at G8 and CD1 positions were found to affect lifetimes, and in some cases lead to the appearance of third kinetic phase. A direct comparison of rate constants is not possible since only apparent rates were reported.

Our results compare well with the average rate constant for CO rebinding to NP4, $4.7 \times 10^9 \text{s}^{-1}$ at pH 7.5 and increasing to $2.1 \times 10^{10} \text{s}^{-1}$ at pH 5.5, retrieved by a single distribution model [33]. In their analysis, Champion and coworkers adopted a statistical model to describe the effect of the strong distortion of the heme in NP4, to explain the observed non-exponential rebinding. In the case of NP7 the ps phase is clearly described by a double exponential decay, and therefore the use of a kinetic model that introduces an additional discrete reaction step appears justified.

The ultrafast dynamics of ligands within heme proteins have been the subject of several studies which were recently reviewed [88,99]. While it has long been known that ligands like O$_2$ and NO rebind on subnanosecond time scale, more recently a few heme-proteins were reported to rebind CO on the picoseconds time scale, some of them through multiple exponential kinetics. Besides TrHbO from *Mycobacterium tuberculosis* [61], ultrafast CO rebinding was observed for the oxygen sensor Dos from *Escherichia coli* [75], the heme-based GAF sensor domains of the histidine kinases DosS and DosT from *Mycobacterium tuberculosis* [71], and the CO-sensing transcriptional activator CooA [72]. A direct comparison with microscopic rates in Table 3 is difficult, because rate constants from fully microscopic kinetic models were not obtained in those studies.

Finally, it is important to stress that the $k_{j}$ values derived from the current determination of microscopic rate constants reproduces...
values that well agree with our previous estimates\(^7\). The combination of microscopic rates to obtain \(k_{\text{on}}\) is such that the resulting parameter is mostly sensitive to \(k_{\text{on}}\), whereas it is relatively insensitive to the actual values of \(k_{\text{1}}\) and \(k_{\text{2}}\) as long as their ratio remains comparable.

Dataset 1. Data of membrane attaching nitric oxide transporter nitrophorin 7
http://dx.doi.org/10.5256/f1000research.6060.d42567

Detailed legends describing each data file can be found in the text file provided.

**Concluding remarks and future directions**
The analysis of the X-ray crystal structures of NP7 at low and high pH and with different heme ligands show the existence of a large resemblance in the overall fold, which is also similar to the spatial arrangement found in other NPs. In particular, the large structural resemblance observed in the heme pocket for liganded and unliganded forms of NP7 support the functional role as a reservoir for NO storage, as it is widely accepted for this family of proteins. A peculiar feature of NP7, nevertheless, is the extensive clustering of Lys side-chains at the protein surface opposite the heme pocket, which is implicated in the charge-stabilized head-to-tail interaction observed in the crystal lattice. Furthermore, it provides a basis to realize the ability of NP7 to bind to negatively charged membranes. These features, together with the presence of tunnels and cavities in the interior of the lipocalin fold, suggest that NP7 could have specific pathways for direct exchange of NO with the membrane. However, packing effects also explain the large structural similarity observed for the different ligand-bound and unbound structures, although such a structural similarity does not permit to rationalize the effect of deletion of the N-terminal residues on the binding affinities of certain ligands nor the pH sensitivity of NP7.

The peculiar structural and dynamical properties of NP7 suggest that the mechanism at the pH sensitivity of this protein exhibits differential features with regard to the one that is operative for the other characterized NP isoforms. The extremely high reactivity of the binding site has the consequence that much of the kinetics resembles for liganded and unliganded forms of NP7 support the functional role as a reservoir for NO storage, as it is widely accepted for this family of proteins. A peculiar feature of NP7, nevertheless, is the extensive clustering of Lys side-chains at the protein surface opposite the heme pocket, which is implicated in the charge-stabilized head-to-tail interaction observed in the crystal lattice. Furthermore, it provides a basis to realize the ability of NP7 to bind to negatively charged membranes. These features, together with the presence of tunnels and cavities in the interior of the lipocalin fold, suggest that NP7 could have specific pathways for direct exchange of NO with the membrane. However, packing effects also explain the large structural similarity observed for the different ligand-bound and unbound structures, although such a structural similarity does not permit to rationalize the effect of deletion of the N-terminal residues on the binding affinities of certain ligands nor the pH sensitivity of NP7.

**Abbreviations**

\(\alpha,\alpha',\alpha\)-microglobulin; DEA/NO, sodium (Z)-1-(N,N-diethylamino)diazene-1-ium-1,2-diolate; Hb, hemoglobin; Hm, histamine; ImH, imidazole; IPTG, isopropyl \(\beta\)-\(d\)-1-thiogalactopyranosid; L, distal ligand on heme iron; LFP, laser flash photolysis; MALDI, matrix assisted laser desorption ionization; Mb, myoglobin; MD, molecular dynamics; MOPS, 3-(N-morpholinopropanesulfonic acid; NOS, nitric oxide synthase; NP, nitrophorin; PDB, Protein Databank of the Research Collaboratory for Structural Bioinformatics (RSCB) at http://www.pdb.org/pdb/home/home.do; RMSD, residual mean square deviation; TOF, time-of-flight; \(wt\), wild-type.

**Author contributions**

HO and MK conducted the X-ray analysis of the NP7 structures. GS, GC, SA, SB and CV performed the spectroscopic studies and the flash photolysis assays. AA and AB-C contributed to the molecular dynamics simulations. MK, CV and FJL contributed to the experimental design and preparation of the manuscript. All authors were involved in the revision of the draft manuscript and have agreed to the final content.

**Competing interests**

No competing interests were disclosed.

**Grant information**

This work was financially supported by the Deutsche Forschungsgemeinschaft (DFG) grant KN951/1-1, the Max Planck Society (both to M.K.), Spanish Ministerio de Economía y Competitividad (SAF2011-27642; FJL), Generalitat de Catalunya (2014SGR1189; FJL), Icrea Academia (FJL). The Consorci de Serveis Universitarios de Catalunya and the Barcelona Supercomputer Center are acknowledged for computational facilities.

I confirm that the funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.
Supporting information

Figure S1. Distribution of positively and negatively charged residues in the head-to-tail interface between NP7 molecules.

Figure S2. Representation of the backbone in the X-ray structure of NP7 solved at low pH (5.8). Residues Pro and Gly of the extra N-terminus stretch (Leu-Pro-Gly) found in NP7 are shown as green-coloured spheres. The spatial position of Leu, and the additional Met0 residue originating from the start codon of the expression system, were not seen in the X-ray crystal.
<table>
<thead>
<tr>
<th>0</th>
<th>10</th>
<th>20</th>
<th>30</th>
<th>40</th>
<th>50</th>
<th>60</th>
</tr>
</thead>
<tbody>
<tr>
<td>NP1:</td>
<td>M</td>
<td>KCTKNAL</td>
<td>AQTGFNKDKY</td>
<td>FNGDVWYVT</td>
<td>YLDLEPDDVP</td>
<td>KRYC</td>
</tr>
<tr>
<td>NP2:</td>
<td>M</td>
<td>DCSTNIS</td>
<td>PKGLDKAKY</td>
<td>FSG–KWYVT</td>
<td>FLDKDP–QVT</td>
<td>DQYCSSTP</td>
</tr>
<tr>
<td>NP3:</td>
<td>M</td>
<td>DCSTNIS</td>
<td>PKGLDKAKY</td>
<td>FSG–TWYVT</td>
<td>YLDKDP–QVT</td>
<td>DPYCSSSTP</td>
</tr>
<tr>
<td>NP4:</td>
<td>M</td>
<td>ACTKNAI</td>
<td>AQTGFNKDKY</td>
<td>FNGDVWYVT</td>
<td>YLDLEPDDVP</td>
<td>KRYC</td>
</tr>
<tr>
<td>NP7:</td>
<td>M</td>
<td>LPGECSVNVI</td>
<td>PKNLDAKAF</td>
<td>FSG–TWYETH</td>
<td>YLDMDP–QAT</td>
<td>EKFC</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>70</th>
<th>80</th>
<th>90</th>
<th>100</th>
<th>110</th>
<th>120</th>
</tr>
</thead>
<tbody>
<tr>
<td>NP1:</td>
<td>YHDPPKQDTO</td>
<td>FYDVSELQEE</td>
<td>SPG–KYTANF</td>
<td>KKVKEKNVQK</td>
<td>VDVTSGNYYT</td>
</tr>
<tr>
<td>NP2:</td>
<td>YYNANKTTS</td>
<td>FYNIGEGLKE</td>
<td>SSSLQYTAKY</td>
<td>KTVDKKKAIVL</td>
<td>KEADEKNSYT</td>
</tr>
<tr>
<td>NP3:</td>
<td>YHFNSSKKTTS</td>
<td>FYNIGEGLKLG</td>
<td>SSVQVYTAKY</td>
<td>NTVDKKRKEI</td>
<td>EPADPKDSTY</td>
</tr>
<tr>
<td>NP4:</td>
<td>YHDPPKQDTO</td>
<td>FYDVSELQVE</td>
<td>SLG–KYTANF</td>
<td>KKVDKNNGVK</td>
<td>VAVTAGNYYT</td>
</tr>
<tr>
<td>NP7:</td>
<td>YHFNSDSKVS</td>
<td>FYNTGTQGLE</td>
<td>SNGAKYTAKF</td>
<td>NTVDKKGKEI</td>
<td>KPADEKYSYT</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>130</th>
<th>140</th>
<th>150</th>
<th>160</th>
<th>170</th>
<th>180</th>
</tr>
</thead>
<tbody>
<tr>
<td>NP1:</td>
<td>ALIHTCLHKG</td>
<td>NKDGLDLAYV</td>
<td>LNNNKNVTAG</td>
<td>DVKKGAVTAA</td>
<td>SLKFSDFIST</td>
</tr>
<tr>
<td>NP2:</td>
<td>ALVHICLREG</td>
<td>SKDGLDLYTV</td>
<td>LTHQKDAEPS</td>
<td>AKVKSATQTA</td>
<td>GLQLGSQFVGT</td>
</tr>
<tr>
<td>NP3:</td>
<td>ALVHICLREG</td>
<td>PKDLGLDLYTV</td>
<td>LSHQKTEEPS</td>
<td>ATVKNAVAGA</td>
<td>GLKLNDVVDT</td>
</tr>
<tr>
<td>NP4:</td>
<td>ALIHTCLHKG</td>
<td>NKDGLDLAYV</td>
<td>LNNNKNVTAG</td>
<td>DVKGSVSAAA</td>
<td>TLEFKSKFIST</td>
</tr>
<tr>
<td>NP7:</td>
<td>ALIHCICLQED</td>
<td>GKDGLDLYSV</td>
<td>LNNKNKALPN</td>
<td>KKKKALNKV</td>
<td>SLVLTKFVVD</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>184</th>
</tr>
</thead>
<tbody>
<tr>
<td>NP1:</td>
</tr>
<tr>
<td>NP2:</td>
</tr>
<tr>
<td>NP3:</td>
</tr>
<tr>
<td>NP4:</td>
</tr>
<tr>
<td>NP7:</td>
</tr>
</tbody>
</table>

**Figure S3.** Amino acid sequence alignment of *R. prolixus* NP1 (Swiss-Port entry Q26239), NP2 (Swiss-Port entry Q26241), NP3 (Swiss-Port entry Q94733), NP4 (Swiss-Port entry Q94734), and NP7 (Swiss-Prot entry Q6PQK2) (1). The total sequence identity among all five protein sequences amount to 33% (indicated by ‘*’). The proximal His is indicated by ‘#’. An initial Met residue (in grey) results in case of the recombinantly expressed proteins of NP1, NP2, NP3, and NP7 is not present in the mature proteins in vivo (2, 3). Residues of relevance to this study are highlighted in color.
Figure S4. Time (ns) evolution of the estimated area (Å²) of the heme cavity mouth determined from the triangles defined by the Cα atoms of residues His60, Lys66, Phe88 and Ile132. Wild type NP7 in open and closed states is shown as red and black lines; Δ(1-3) variant in open and closed states is shown as blue and green lines.

Figure S5. Absorption spectra of ferric and ferrous NP7 at pH 7.5. Black: NP7[Fe³⁺]; red: NP7[Fe²⁺]; blue: NP7[Fe¹−CO].


44. He C, Noya S, Knipp M: Breaking the proximal Fe3+-N4 bond in heme proteins through local structural tension: lessons from the heme b proteins nitrophorin 4, nitrophorin 7, and related site-directed mutant proteins. Biochemistry. 2011; 50(1): 6559–6575. Published Abstract | Publisher Full Text


Open Peer Review

Current Referee Status: ✓ ✓ ?

Version 1

Referee Report 27 July 2015
doi:10.5256/f1000research.6488.r9066

Antonio Cupane, Matteo Levantino
Department of Physics and Chemistry, University of Palermo, Palermo, Italy

The article by Knipp et al. reports the results of an extensive investigation of the nitrophorin isoform NP7. The authors have resolved the X-ray crystallographic structure of NP7 at different pH values and ligation states, measured the ligand rebinding kinetics after laser photolysis of CO bound NP7 at different pH values and performed MD simulations on different NP7 conformations to assess the degree of flexibility of different protein domains and the topology of inner protein cavities that are shown to play a significant role in the binding of ligands to the protein.

The presented results are new and significant to our understanding of the structure-function-dynamics relationship in the NP7 nitrophorin. However, the paper would benefit from a more clear discussion of an important aspect that emerges from the results presented by authors: the charge distribution across NP7, which peculiarly determines the ability of this particular nitrophorin isoform to bind negatively charged membranes, is likely to significantly affect both the way NP7 molecules are arranged in the crystalline state and their overall 3D structure in the crystal. As stated by the authors, the surprisingly small differences between the crystal structure of NP7-H2O at pH 5.8 and 7.8 could be due to the overwhelming contribution of electrostatic interactions between neighboring proteins within the crystal. The authors refrained from using their NP7 X-ray structures as starting points for their MD simulations since they admittedly do not account for the conformational change expected to occur upon increase of the pH from 5.8 to 7.8. Overall, these findings seem to indicate that, in the case of NP7 nitrophorin, the structural models retrieved from X-ray crystallography fail to accurately describe the 3D structure of the protein in solution.
It would be worth to better clarify and discuss this point in the paper.

A further important result of the paper comes from the analysis of ligand rebinding kinetics after photolysis of NP7-CO (presumably at a much lower protein concentration, at which interparticle interactions are negligible). A large fraction of CO ligands rebind geminately in the picosecond time-scale with a biphasic time course. Moreover, nanosecond flash-photolysis experiments clearly show that ligand migration through protein cavities modulates the rebinding process at longer time-scales. Although the main findings are clear already from inspection of the raw data, the procedure used to match the kinetics measured in the sub-nanosecond time-scale with those measured at longer time-scales seems problematic (as also noted by Marten H. Vos in his review of the paper). The detailed procedure used to match the two datasets should be added to the paper. In particular, a supplementary figure showing the two-exponentials fit of the picosecond data and its extrapolation beyond 2 ns, would be helpful in this respect. It would be also useful to have an idea of how much a different choice of the relative scaling between the two datasets would affect the estimation of microscopic rates.
Minor points:
- as noted by Suman Kundu in his review of the paper there are sentences that need to be corrected in the abstract; apart from the second sentence (line 3), also the last sentence (line 17) “Finally, the topological distribution of pockets located around the heme as well as from inner cavities …” clearly needs to be corrected.
- page 3, last paragraph of the Introduction: “Here we report on seven crystal structures of NP7 …”. Only 6 different crystal structures are described in the paper and in Table 1.
- the caption of Figure 12 should be corrected in comparison to Figure 11: T1 should be T2.

**Competing Interests:** No competing interests were disclosed.

We have read this submission. We believe that we have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however we have significant reservations, as outlined above.

---

**Author Response 05 Aug 2015**

**Cristiano Viappiani,** Università degli Studi di Parma, Italy

The article by Knipp et al. reports the results of an extensive investigation of the nitrophorin isoform NP7. The authors have resolved the X-ray crystallographic structure of NP7 at different pH values and ligation states, measured the ligand rebinding kinetics after laser photolysis of CO bound NP7 at different pH values and performed MD simulations on different NP7 conformations to assess the degree of flexibility of different protein domains and the topology of inner protein cavities that are shown to play a significant role in the binding of ligands to the protein. The presented results are new and significant to our understanding of the structure-function-dynamics relationship in the NP7 nitrophorin. However, the paper would benefit from a more clear discussion of an important aspect that emerges from the results presented by authors: the charge distribution across NP7, which peculiarly determines the ability of this particular nitrophorin isoform to bind negatively charged membranes, is likely to significantly affect both the way NP7 molecules are arranged in the crystalline state and their overall 3D structure in the crystal. As stated by the authors, the surprisingly small differences between the crystal structure of NP7-H2O at pH 5.8 and 7.8 could be due to the overwhelming contribution of electrostatic interactions between neighboring proteins within the crystal. The authors refrained from using their NP7 X-ray structures as starting points for their MD simulations since they admittedly do not account for the conformational change expected to occur upon increase of the pH from 5.8 to 7.8. Overall, these findings seem to indicate that, in the case of NP7 nitrophorin, the structural models retrieved from X-ray crystallography fail to accurately describe the 3D structure of the protein in solution.

It would be worth to better clarify and discuss this point in the paper.

In agreement with the reviewer, we believe that both crystal packing and electrostatic interactions impose a severe constraint on the AB loop, limiting the conformational flexibility of this region, and this likely explains why there seems to be no apparent major difference between the structures at pH 5.8 and 7.8. This has been explicitly stated in the revised version of the manuscript.

A further important result of the paper comes from the analysis of ligand rebinding kinetics after
photolysis of NP7-CO (presumably at a much lower protein concentration, at which interparticle interactions are negligible). A large fraction of CO ligands rebind geminately in the picosecond time-scale with a biphasic time course. Moreover, nanosecond flash-photolysis experiments clearly show that ligand migration through protein cavities modulates the rebinding process at longer time-scales. Although the main findings are clear already from inspection of the raw data, the procedure used to match the kinetics measured in the sub-nanosecond time-scale with those measured at longer time-scales seems problematic (as also noted by Marten H. Vos in his review of the paper). The detailed procedure used to match the two datasets should be added to the paper. In particular, a supplementary figure showing the two-exponentials fit of the picosecond data and its extrapolation beyond 2 ns, would be helpful in this respect. It would be also useful to have an idea of how much a different choice of the relative scaling between the two datasets would affect the estimation of microscopic rates.

We have addressed this issue in response to the similar argument raised by Marten Vos (Reviewer #1). Within the “Ligand rebinding kinetics” section we have added a statement about the effect of a wrong estimate of the extrapolated value of the signal from the subnanosecond kinetics. We provide an estimate of the effect on the rates constants for over/under-estimating the residual absorption after the subnanosecond rebinding phase. As detailed in the text, even a relatively large 10% error results in modest effects on rate constants.

Minor points:

as noted by Suman Kundu in his review of the paper there are sentences that need to be corrected in the abstract; apart from the second sentence (line 3), also the last sentence (line 17) “Finally, the topological distribution of pockets located around the heme as well as from inner cavities …” clearly needs to be corrected.

We have revised the sentence as suggested by Suman Kundu

page 3, last paragraph of the Introduction: “Here we report on seven crystal structures of NP7 …”. Only 6 different crystal structures are described in the paper and in Table 1.

This wrong statement was revised. We thank this reviewer for pointing that out.

the caption of Figure 12 should be corrected in comparison to Figure 11: T1 should be T2.

As noted above, we have revised the caption.

**Competing Interests:** No competing interests were disclosed.

---

doi:10.5256/f1000research.6488.r9064

Suman Kundu
Department of Biochemistry, University of Delhi, New Delhi, Delhi, India

Markus Knipp et al applied a repertoire of biophysical and computational techniques to provide insight into the unique structural features of NP7, a transporter nitrophorin. The study also highlighted the
differences with other nitrophorins and unraveled a likely mechanism for NO migration and exit in NP7, which represents advancement in the understanding of nitrophorins. This is a nice, elaborate piece of work that triggers the mind in many ways.

The article will read better and assume greater significance if the following issues can be addressed:

- The second sentence (line 3) in the Abstract should be modified as “Besides its ability to bind to phospholipid membranes, the N-terminus of NP7, a member of the NO transporter nitrophorin family, contains …………..”; otherwise the “other” in that sentence does not make any sense. Moreover, placing NP7 in this sentence also introduces the target protein to the reader better, than mentioning it later. Without such an introduction of the subject of the study, the beginning of the abstract reads pretty abrupt and can be difficult for readers to grasp, unless they come back to it after reading the Introduction.

- How does one ensure that NO stays bound to heme iron in the crystal structures of NPs, given the photoreduction and the fact that Fe(II) might have altered affinity for NO than Fe(III)? The fact that the electron density could not be fit with a single configuration of NO indicates that NO is only partially bound, which means that the crystals contain a mixture of liganded and unliganded states? Does such mixed population influence conclusions about structural features? A sentence in this regard could be added in the relevant section in the manuscript to dispel doubts.

- Since the authors solved the crystal structures of NP7 at low (5.8) and high pH (7.8) and in NO-bound form, for MD simulations these structures could be used to model the molecular systems rather than using NP4 structures. Alternatively, the models obtained from NP4 could be compared with the crystal structures. Clarification would help readers.

- Since the liganded and unliganded structures are similar, is it safe to assume that the structure of NP7 is not adapted to regulate ligand binding, but rather acts as a reservoir for NO storage suitably endowed with tunnels and cavities for the NO release as and when required, with dependence on pH only? A line of discussion would help.

- Finally, as a future perspective, a crystal structure of the mutant of NP7 (with extra N-terminal residues deleted), both in unliganded and liganded forms, might help to validate the rational hypothesis laid down in the paper.

I congratulate the authors on the wonderful manuscript.

**Competing Interests:** No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

---

**Author Response 05 Aug 2015**

**Cristiano Viappiani**, Università degli Studi di Parma, Italy

*Markus Knipp et al applied a repertoire of biophysical and computational techniques to provide insight into the unique structural features of NP7, a transporter nitrophorin. The study also highlighted the differences with other nitrophorins and unraveled a likely mechanism for NO migration and exit in NP7, which represents advancement in the understanding of nitrophorins. This is a nice, elaborate piece of work that triggers the mind in many ways.*

We thank this reviewer for his appreciation of our work.

*The article will read better and assume greater significance if the following issues can be addressed:*
The second sentence (line 3) in the Abstract should be modified as “Besides its ability to bind to phospholipid membranes, the N-terminus of NP7, a member of the NO transporter nitrophorin family, contains…………….”; otherwise the “other” in that sentence does not make any sense. Moreover, placing NP7 in this sentence also introduces the target protein to the reader better, than mentioning it later. Without such an introduction of the subject of the study, the beginning of the abstract reads pretty abrupt and can be difficult for readers to grasp, unless they come back to it after reading the Introduction. The underlined words can be added to improve the readability of the sentence.

We thank reviewer #2 for pointing this out, we completely agree that the sentence was missing a reference to the specific protein, which is the subject of the present study.

How does one ensure that NO stays bound to heme iron in the crystal structures of NPs, given the photoreduction and the fact that Fe(II) might have altered affinity for NO than Fe(III)? The fact that the electron density could not be fit with a single configuration of NO indicates that NO is only partially bound, which means that the crystals contain a mixture of liganded and unliganded states? Does such mixed population influence conclusions about structural features? A sentence in this regard could be added in the relevant section in the manuscript to dispel doubts.

The occupancy of NO ligand was refined to 0.44. The crystal structure in this case contains a mixture of the NO-bound and NO-unbound states, however, this does not influence conclusions. The occupancy of NO has been included in the revised manuscript.

Since the authors solved the crystal structures of NP7 at low (5.8) and high pH (7.8) and in NO-bound form, for MD simulations these structures could be used to model the molecular systems rather than using NP4 structures. Alternatively, the models obtained from NP4 could be compared with the crystal structures. Clarification would help readers.

We have already addressed this point which was also raised by reviewer #1. As stated above, the structural similarity of the protein backbones of the homology and X-ray model is now mentioned in the revised version (Materials and Methods. MD simulations).

Since the liganded and unliganded structures are similar, is it safe to assume that the structure of NP7 is not adapted to regulate ligand binding, but rather acts as a reservoir for NO storage suitably endowed with tunnels and cavities for the NO release as and when required, with dependence on pH only? A line of discussion would help.

We have modified the first paragraph of the “Concluding remarks and Future Directions” section to discuss this aspect.

Finally, as a future perspective, a crystal structure of the mutant of NP7 (with extra N-terminal residues deleted), both in unliganded and liganded forms, might help to validate the rational hypothesis laid down in the paper.

It has been mentioned in the last paragraph of the manuscript that availability of the crystal structure of the protein missing the three amino acids at the N-terminus may help understanding the role of this short sequence.

**Competing Interests:** No competing interests were disclosed.
Marten H. Vos
Laboratory of Optics and Biosciences, CNRS, Ecole Polytechnique, Palaiseau, France

This work combines crystallography, time-resolved spectroscopy and MD simulations to investigate structure and CO dynamics in a member of the NO transporter NP family, NP7. Whereas the found properties are similar to other NPs, in particular the well-studied NP4, this work is useful as substantial differences had been suggested from previous studies.

A few suggestions for improvements:

1. Intro, 2nd par, 3rd phrase suggests that NP1 and NP2 share more identity with NP4 than NP7, which appears not the case for NP2 (Table 2).

2. Please specify sample conditions (path length, concentration) for both types of spectroscopic measurements.

3. MD simulations of NP7 are based on homology modelling from NP4 structures. A comparison of the homology and X-ray models, other than the flipped heme, would be interesting.

4. Admittedly it will not change the order of magnitude of the results, but the problematic gluing of the gap between 1 ns and 20 ns by extrapolating a non-finished double exponential (fit parameters?) can be avoided by determining the effective CO dissociation yield at ~20 ns in the linear excitation regime using an established procedure (Brunori et al., 1973).

5. If I understand well the kinetics in Fig. 9 can be fitted by four exponentials (two <10 ns and two μs-ms), and thus 7 independent free parameters. The model has as many as 11 rate parameters (Table 3, $k_{o}$ is strictly not a microscopic rate constant). This would suggest the model is underdetermined. Please discuss.

6. Do the authors associate T1, T2 and T3 with specific modeled cavities?

7. It is unclear why the rebinding rate is specifically or only compared with TrHbO. In a number of sensor proteins, including CooA, CO rebinding phases on the order of tens and hundreds of picoseconds have been observed.

Competing Interests: No competing interests were disclosed.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Cristiano Viappiani, Università degli Studi di Parma, Italy

This work combines crystallography, time-resolved spectroscopy and MD simulations to investigate structure and CO dynamics in a member of the NO transporter NP family, NP7.
Whereas the found properties are similar to other NPs, in particular the well-studied NP4, this work is useful as substantial differences had been suggested from previous studies.

A few suggestions for improvements:

Intro, 2nd par, 3rd phrase suggests that NP1 and NP2 share more identity with NP4 than NP7, which appears not the case for NP2 (Table 2).

The sentence has been misinterpreted. The sequence identity is determined for NP1 versus NP4 (88%), and for NP2 versus NP3 (80%). The sentence has been rewritten in the revised version. Likewise, Table 2 has been modified to include the sequence identity with NP3, even though the lack of an X-ray structure impedes to include the data for the structural resemblance.

*Please specify sample conditions (path length, concentration) for both types of spectroscopic measurements.*

This relevant information was left out and has been included in the revised manuscript.

*MD simulations of NP7 are based on homology modelling from NP4 structures. A comparison of the homology and X-ray models, other than the flipped heme, would be interesting.*

The structural similarity of the protein backbones of the homology and X-ray model has been mentioned in the revised version (Materials and Methods. MD simulations).

*Admittedly it will not change the order of magnitude of the results, but the problematic gluing of the gap between 1 ns and 20 ns by extrapolating a non-finished double exponential (fit parameters?) can be avoided by determining the effective CO dissociation yield at ~20 ns in the linear excitation regime using an established procedure (Brunori et al., 1973).*

We agree that this would be the best way to handle the data. Indeed, this procedure was applied to merge the two time regimes in our previous works on trHb from *Thermobifida fusca* (Marcelli et al., 2012; Bustamante et al., 2014).

However, unlike that case, in the current nanosecond experiments we were unable to reach a low enough laser pulse energy range where multiple photolysis becomes completely negligible, yet with a reasonable signal to noise ratio. This impairs the possibility of using the above mentioned procedure. For this reason we resorted to the approach we adopted for the present experiments. We are aware that the present data normalization procedure may bias to some extent the kinetics, but this effect is expected to be minor, as already mentioned in the original submission. A further statement about this has been added in the “Ligand rebinding kinetics” section.

*If I understand well the kinetics in Fig. 9 can be fitted by four exponentials (two <10 ns and two μs-ms), and thus 7 independent free parameters. The model has as many as 11 rate parameters (Table 3, k_on is strictly not a microscopic rate constant). This would suggest the model is underdetermined. Please discuss.*

Overall kinetics can be approximated by 4-5 exponential decays. Apparent rates and amplitudes are combinations of microscopic rates, some of which (in the bimolecular phase) are dependent on the reactant concentration. By performing a global analysis of the rebinding kinetics at two different CO concentrations it is possible to remove almost entirely the problem of under-determination of parameters due to their cross correlation.
Do the authors associate T1, T2 and T3 with specific modeled cavities?

Based on their location, it appears reasonable to associate transient population of the cavities T2 and T3 close to the distal pocket with fast reaction intermediates, whereas the longer lived reaction intermediate T4 is interpreted as rebinding from the cavity in the back tunnel. In the revised manuscript we have indicated putative locations of the cavities with reference to isocontours in Figure 10.

Incidentally, we note that the caption to Figure 12 had a typo, where the cavities were erroneously labeled T1, T2 and T3 instead of T2, T3 and T4. This was noted also by Reviewer #3. The scheme in Figure 11 also contained a typo, which is now corrected in the revised version. We apologize for that.

It is unclear why the rebinding rate is specifically or only compared with TrHbO. In a number of sensor proteins, including CooA, CO rebinding phases on the order of tens and hundreds of picoseconds have been observed.

This remark is certainly relevant. The comparison of the rate constants proposed in the final paragraphs refers to trHbO only because this is the only available set of microscopic rates to compare with.

Indeed, several excellent papers have appeared in the literature, reporting apparent lifetimes/rate constants for CO rebinding. We have expanded the discussion in the revised manuscript to include some recent works.

**Competing Interests:** No competing interests were disclosed.