REVIEW

Individuality, phenotypic differentiation, dormancy and ‘persistence’ in culturable bacterial systems: commonalities shared by environmental, laboratory, and clinical microbiology [version 2; referees: 2 approved, 1 approved with reservations]

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Abstract

For bacteria, replication mainly involves growth by binary fission. However, in a very great many natural environments there are examples of phenotypically dormant, non-growing cells that do not replicate immediately and that are phenotypically ‘nonculturable’ on media that normally admit their growth. They thereby evade detection by conventional culture-based methods. Such dormant cells may also be observed in laboratory cultures and in clinical microbiology. They are usually more tolerant to stresses such as antibiotics, and in clinical microbiology they are typically referred to as ‘persisters’. Bacterial cultures necessarily share a great deal of relatedness, and inclusive fitness theory implies that there are conceptual evolutionary advantages in trading a variation in growth rate against its mean, equivalent to hedging one’s bets. There is much evidence that bacteria exploit this strategy widely. We here bring together data that show the commonality of these phenomena across environmental, laboratory and clinical microbiology. Considerable evidence, using methods similar to those common in environmental microbiology, now suggests that many supposedly non-communicable, chronic and inflammatory diseases are exacerbated (if not indeed largely caused) by the presence of dormant or persistent bacteria (the ability of whose components to cause inflammation is well known). This dormancy (and resuscitation therefrom) often reflects the extent of the availability of free iron. Together, these phenomena can provide a ready explanation for the continuing inflammation common to such chronic diseases and its correlation with iron dysregulation. This implies that measures designed to assess and to inhibit or remove such organisms (or their access to iron) might be of much therapeutic benefit.

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Invited Referees

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Amendments

This summary of the differences between versions 2 and 1 is very short, since we simply made modifications as described in our response to the referees’ very helpful comments, particularly around recognising the semantic issues (persistence vs dormancy). We rehearsed a little more some of our areas of ignorance of the detailed physiological states that these high-level, replicatorily observable phenotypes represent. We clarified the meaning of “reversibility” (of growth/non-growth) in terms of states vs mechanisms. We added references to some work that we had missed, e.g. that of McKinney. We amplified slightly the points about how a ‘standing crop’ of mainly non-growing bacteria (else it would be sepsis) must reflect a balance between resuscitation, growth and clearance, and how these and related questions (e.g. how cells evade the innate and adaptive immune systems) represent a future ‘to do’ list. We stressed further that the observation of bacterial sequences in the absence of immediately culturable microbes always implies their potential for resuscitation/regrowth, although it cannot, of course, discriminate dormant from moribund, injured or reversibly nonculturable (“dead”) cells. We added a paragraph on the more philosophical reasoning behind our approach, which takes the idea that a self-consistent narrative is more persuasive intellectually than one lacking elements of joint-up, a principle known in Philosophy of Science circles as ‘coherence’. Finally, we entirely redid Table 3 to make its layout much more logical, and streamlined it so as to add more emphasis on the nature of the evidence of bacterial involvement in the various classes of diseases.

See referee reports

Introduction

“It is now well established that some micro-organisms can, under certain conditions, be deprived of all visible signs of life and yet these organisms are not dead, for, when their original conditions are restored, they can return to normal life and activity.”

“Bacterial populations in both batch and continuous culture are much more heterogeneous than is normally assumed, and such cultures may consist of several types of subpopulations simultaneously differing in viability, activity and integrity of the cells”. Consider a typical axenic flask or broth culture of bacteria (Figure 1), arguably the staple of modern laboratory microbiology. We seed a suitable growth medium with an appropriate inoculum of cells known to be capable of replicating in that growth medium. After a lag phase the number of culturable cells (the ‘viable count’) as judged by plate counts of the number of colony-forming units observable on the same medium solidified by agar or a similar material is observed to increase, typically exponentially, for a number of generations (the growth phase or exponential phase). Apart from the changes in nutrient concentration, and for non-synchronised cultures, it is generally taken that cells pass smoothly through their cell cycles en route to doubling their numbers by binary fission. The population distribution of organisms in different parts of their cell cycle during the exponential phase is thereby unchanged and thus in a steady state (from which the cell cycle parameters can even be inferred). In time this increase in cell numbers ceases, usually because of the exhaustion of a nutrient in a closed system, or sometimes in part or whole because of the build-up of toxins. Again, after a further period, the viable or colony count decreases (often to quite low levels if such starvation is carried out for extended periods). Inoculation of a new broth culture with a similar number of viable cells from this culture usually provides a simple repeat of the previous culture, and in the absence of mutation may reasonably be anticipated, for organisms proliferating asexually, to be played out indefinitely.

The development of continuous, nutrient-limited (‘chemostat’) or feedback-controlled (‘turbidostat’) cultures was and is entirely consistent with this view of steady-state microbial doubling via homogeneous cell cycles that are common, within statistical fluctuations, to each cell. The same is true for cultures undergoing serial transfer (where there is slightly more of a focus on selection for genotypic variants that grow faster – see e.g. 12–14).

There should be nothing controversial in the above passage, but in fact it hides a variety of assumptions that themselves conceal a considerable feast of very interesting physiology. The chief one here is that – given that all cells in the culture are genetically homogeneous and see the same ‘environment’, and modulo where they are in their cell cycles – all such cells are indeed supposed to represent a single population (as per Figure 2). If they do not, and as we shall see they never do15–18, we are dealing with differentiated systems. It turns out that a particular subset of typical cell cultures – a phenotypically dormant or non-growing sub-population, occurring even in non-sporulating bacteria – is widespread to the point of ubiquity. This leads to an exceptionally important biology with significant consequences both for our understanding of microorganisms and our ability to harness and domesticate them. Although the relevant literatures rarely cite each other or overlap, it is clear that similar phenomena are common to bacterial behaviour in the natural environment, the laboratory, and in a variety of samples of clinical interest. This theory or hypothesis that we develop here comes about from the synthesis15 of a large amount of data, and is summarised in Figure 3 and Figure 4.

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**Figure 1. A typical laboratory bacterial culture.** After the end of stationary phase the viable count decreases over time, but very rarely to precisely zero. Some authors recognise an extended “period of prolonged decrease” during which some of the survivors undergo significant dynamics, and in which mutants are selected. Our interest here is largely in cells that have not mutated.

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**Figure 2. Log (plate count) vs Time**

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**Figure 3.**

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**Figure 4.**
Phenotypic differentiation to dormancy or persistence – some early indications
While dormancy and resuscitation of rotifers had been observed by Leeuwenhoek himself in 1702, some of the earliest modern indications for a physiologically significant ‘phenotypic heterogeneity’ or differentiation of microbial cultures came in the 1940s. In a conceptually simple experiment (illustrated in Figure 5), Bigger exposed staphylococcal cultures to concentrations of penicillin that would normally be sufficient to kill them completely (and they did kill all but 1 in a million). However, these (10⁻⁶) survivors, that Bigger and McDermott (and many modern commentators have) referred to as ‘persisters’, were not genetic mutations selected for resistance to penicillin, since when they were inoculated into fresh broth they were just as susceptible as were those in the first culture. Bigger recognised (correctly) that the only explanation that made any kind of sense was that despite being exposed to nominally the same conditions, these cells were operationally dormant in the sense of not replicating in a medium that, apart from the penicillin, would normally admit their growth (even if they were metabolically active, and thus phenotypically resistant to the penicillin (that anyway kills only dividing cells)). Similarly, Luria and Latarjet

Figure 2. To clarify the general concept of a population as used here, a population of individuals involves those who share certain properties (between stated values). One main population is shown. A second, smaller population is also shown; these might represent dormant cells.

Figure 3. Infographic summary of the review. (1) A bacterial system contains distinct subpopulations, that we classify as culturable, dormant and non-culturable (2). Specific attention is given to persister cells (3), and the inter-relationship (4) between the subpopulations. Subpopulations within environmental biology are discussed (5), followed by subpopulations within laboratory cultures (6). Particular emphasis is placed on phenotypic switching between the culturable and dormant subpopulation of laboratory cultures (7). Generalized detection techniques typically fail to detect dormant cells, and we review the various reasons for this failure and discuss alternatives (8). Resuscitation of and endotoxin production by such dormant cells underpins many diseases not normally seen as having a microbial component.
noted that approximately 1% of the cells in a culture of *Escherichia coli* displayed a phenotypic resistance to normally sterilising doses of ultraviolet irradiation. Many similar experiments since (e.g. 29–32), discussed in more detail below, have recapitulated this basic phenomenon. (We note here that high-frequency antigenic ‘phase’ variation can occur due e.g. to changes in microsatellite DNA; detailed discussions of such genotypic changes, including those that can affect the extent of dormancy in persistent bacteria, are outwith the scope of the present, purely phenotypic analyses.)

**Dormancy as an operational property, and semantic issues**

For the avoidance of doubt, and in accordance with Keilin’s description with which we opened, we shall define dormancy as:

“a reversible state of [often] low metabolic activity, in which cells can persist for extended periods without division; we shall see that this often corresponds to a state in which cells are not ‘alive’ in the sense of being able to form a colony when plated on a suitable solid medium, but one in which they are not ‘dead’ in that when conditions are more favourable they can revert to a state of ‘aliveness’ as so defined”.

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**Figure 4. Summary of the review in the form of a ‘mind map’ of the article.**

**Figure 5. Assessment of phenotypic differentiation of a dormant subpopulation via antibiotic challenge.** This kind of protocol can be used to determine if the resistant subpopulation has accumulated genetic mutations that encoded resistance or whether, as focused on here, the resistance is purely phenotypic. A detailed analysis of the shape of the time-survivor curves may also be informative.
We thus stress\(^{36}\) the recognition that dormancy is not solely an innate property of a bacterial cell; it is a property assessed by one or more experiments, so whether a cell appears to be dormant depends on both the cell and the experiment used to assess that dormancy. (This principle shares a similar philosophical foundation to the independence from any specific experiment, or otherwise, of the perceived state of objects within the quantum theory\(^{36–39}\)). As do Postgate\(^{34,39}\) and Barer\(^{40–44}\), we take the hallmark of a viable or living bacterial cell to be its ability to replicate or its ‘culturability’. This means that we cannot tell via culturability that a cell is alive, only (after a cell division) that it was alive\(^{36,45}\). Dormant cells – even if ‘not immediately culturable’ – must by definition be resuscitable to form culturable cells. We also recognise (as does Michael Barer\(^{489}\)) that it may be hard to discriminate the resuscitation of dormant cells from the recovery of injured cells. Although the term ‘nonculturable’ is quite commonly used to describe not-immediately-culturable cells, it is best avoided, as we cannot try every possible combination\(^{36}\) of incubation conditions that might serve to resuscitate a cell in a sample. ‘Non-cultured’, ‘as-yet-uncultured’ or ‘operationally nonculturable’ are better terms. Culturable, (operationally) non-culturable and (operationally) dormant bacteria in the differentiated bacterial (cellular) system can therefore be seen as distinct subpopulations of the system, and cultivable and dormant bacteria as reversible states of the same population. A culture containing several subpopulations, whether distinct (as in Figure 2, or part of a single population characterised by a particular value from a range of an extensive variable) may be said to be differentiated (and of course may de-differentiate) in terms of physiological macrostates, that may or may not be able to interconvert. However, we recognise (thanks to Michael Barer\(^{489}\)) that such interconversion does not imply a mechanistic reversibility. The same kinds of issues attach to cells described as having any other physiological property with regard to the ability to replicate. We note (with thanks again to Michael Barer\(^{489}\)) that it is easy to confuse dormancy and ‘persistence’, since they do share some similarities (e.g. such cells are not immediately replicable); however, there is not much in the way of evidence as to how different say their expression profiles are, since it would require, for instance, single cell omics measurements, that are only just becoming available (e.g. \(^{47,48}\)), more typically\(^{49}\) for the much larger eukaryotic cells. Certainly there can be extensive changes in gross biochemical composition as cultures are starved\(^{46}\). One strategy would be to separate sub-populations\(^{31,32}\), acquire ‘averaged’ values of say their transcriptome, proteome or metabolome, and see how much they differed. In a similar vein, whether states such as dormancy are adaptive is a matter for experiment.

The general relationships between various subpopulations of the bacteria within a differentiated cellular system are shown in Figure 6.

**Figure 6.** The relationships between culturable, dormant and non-culturable bacteria within a differentiated cellular system.

**Table 1.** Some bacterial infections for which an intracellular, reversibly non-replicating, persistent or dormant state is well established as part of the cells’ lifestyle. Examples are given for both low- and high-GC Gram positives, as well as a number of Gram-negative organisms.

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<th>Comments</th>
<th>Selected references</th>
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<td>Bartonella spp.</td>
<td>Persists inside erythrocytes</td>
<td>347–350</td>
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<tr>
<td>Brucella spp.</td>
<td>Environmental and intracellular persistence and immune evasion</td>
<td>351–354</td>
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<td>Listeria monocytogenes</td>
<td>Well-established low-GC Gram-positive intracellular saprophyte and non-sporulating persister</td>
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<td>Mycobacterium tuberculosis</td>
<td>Often seen as the ‘classical’ dormant bacterium, a high-GC Gram-positive; probably one third of humans carry it in a latent or potentially dormant state; other forms may be metabolically active</td>
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<tr>
<td>Salmonella typhimurium</td>
<td>Gram-negative; non-replicating forms common in macrophages and elsewhere</td>
<td>367–370</td>
</tr>
<tr>
<td>Staphylococcus aureus</td>
<td>Low-GC Gram-positive; can escape antibiotics by hiding inside various phagocytes</td>
<td>371–374</td>
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culturability, only presence or activity. Similarly, macromolecular sequencing methods such as those based on rDNA and its amplification (e.g. 53–58), or that of other housekeeping genes (e.g. 59–61), almost certainly reflect mainly dormant cells plus any actively dividing ones (in that ‘naked’ DNA is usually degraded fairly rapidly in serum or the environment). The difference between culturable counts and total sequence-based counts probably provides one of the best methods for detecting and enumerating potentially dormant cells when they cannot yet be brought back into culture, although (as recognised by referee 1) such differences may also reflect dead, injured or moribund cells. It is particularly noteworthy (and see also 62 and below) that the amount of prokaryotic DNA in whole blood exceeds by 10–100-fold that detectable in serum36, implying adsorption onto or sequestration within blood cells.

We shall return to clinical and laboratory microbiology later, but it is to environmental microbiology that we now turn to discuss the culturability of typical microbes. While the same general truths undoubtedly pertain in viruses (e.g. 64,65), and in yeasts, fungi, archaea, mycoplasmas and other unicellular organisms, our focus will be on prokaryotes.

Bacterial culturability and dormancy in environmental microbiology

It has long been known that the number of bacteria observable microscopically exceeds, typically 100-fold, those that can readily be grown axenically in standard isolation media (i.e. to proliferate in liquid culture or to form colonies on solid media). The latter has been referred to as ‘the great plate count anomaly’66, and has been amply confirmed by more modern, culture-independent sequencing methods. A selection of papers and reviews serve to document both the numerical anomaly and the much greater biodiversity detectable by sequencing (e.g. 67–86). It is thus useful to discriminate (1) bacteria that have been cultured, that are typically available in culture collections, and whose growth requirements are known, from (2) bacteria that may be recognised as novel via macromolecular sequencing (typically of ribosomal DNA30,171–179) but that have not yet been cultured and whose growth requirements may not yet even be known. Much (sequencing) evidence indicates that the bulk of the ‘missing microbes’ or ‘dark matter’170–173 in natural ecosystems falls into this second category18, and that ‘single cell’ methods may be required to culture them19.

There are at least four general reasons of principle why these organisms have not yet been cultured. We consider each in turn (although more than one may contribute in individual cases).

Not-yet-cultured bacteria may have more-or-less fastidious growth requirements

It is an elementary observation in microbiology, and the basis for selective isolation media, that not all bacteria grow on all media and in all conditions. Leaving aside truly syntrophic bacteria (that for thermodynamic or unknown nutritional reasons require another organism for growth (e.g. 96–102)), some organisms may have quite fastidious growth requirements. A number of bacteria determined as causative of disease, whose role had originally been inferred only through microscopic observation, were later cultured and could be shown to fulfill Koch’s postulates. These include *Helicobacter pylori*20,103,104 (with an unusually high requirement for urea to fuel its alkalogenic urease activity20) and *Legionella pneumophila*106–109 (with an unusually high requirement for cysteine). Note that even the supposedly rich LB medium10 (Lysogeny Broth, often erroneously called Luria-Bertani medium, see http://schaechter.asmblog.org/schaechter/2009/11/the-limitations-of-lb-medium.html) is not in fact a particularly rich medium110–112. An especially nice example113,114 is provided by *Tropheryma whippelii*, the causative organism of Whipple’s disease115,116. It resisted attempts (over many decades) to bring it into axenic culture until systematic genome sequencing117,118 showed its requirements for a variety of common amino acids that it was unable to synthesise itself, the provision of which permitted its growth. The MetaGrowth database119 is now available for similar purposes. Another good example is *Coxiella burnetii*, the causative agent of Q fever, for which a genome-derived growth medium (‘acidified citrate cysteine medium’) permitting axenic culture has now been developed120,121. Other examples are given by Stewart122 and by Singh and colleagues123, and include marine bacteria of the highly common SAR11 clade124,125,126. Of course these kinds of phenomena are not absolute; much evidence indicates that host stress hormones may act as growth or virulence factors for a variety of Gram-negative organisms, representing a kind of ‘microbial endocrinology’ (e.g. 126–128).

Not-yet-cultured bacteria may even be killed by our isolation media

Organisms in nature are often living in low-nutrient conditions129–133. It is thus reasonable (and unsurprising) that the isolation of microbes from starved, oligotrophic environments benefits from the use of low-nutrient conditions36,123,134–136, some manifest this ‘starvation’ through their size, as ‘ultramicrobacteria’ (see e.g. 137–143). In a similar vein, taking cells from low-nutrient natural environments directly onto, say, a highly aerobic agar plate may produce stresses that effectively kill them, so that afterwards they would not even grow on the kinds of media (as in the previous section) that would support their growth. Thus, Tanaka and colleagues138 showed interactions between phosphate and agar when autoclaved together that led to the production of compounds inimical to bacterial growth. Gellan may be a better solidifying agent here18. However, we recognise that it may be hard to discriminate cells that we kill in the act of trying to isolate and grow them from ‘already dead’ bacteria.

Not-yet-cultured bacteria may simply be dead and thus incapable of resuscitation

While this possibility certainly exists, and is included for completeness, it is actually the least likely for a number of conceptual and empirical reasons. The first is that if an organism is present in a particular environment it must have been able to grow and divide in it at some point in the more or less recent past, even if the result of such growth was its utilisation of a finite amount of necessary nutrients or growth factors whose exhaustion caused replication to cease. (Interestingly, in soil it seems that sequestration, rather than nutrients or growth factors whose exhaustion caused replication to cease. (Interestingly, in soil it seems that sequestration, rather than complete exhaustion, of nutrients is the more significant phenomenon145–147.) Secondly, it is highly unlikely that evolution could select for unicellular organisms that cannot replicate. Thirdly, environmental organisms can be shown to metabolise even when they cannot be shown to divide (e.g. in the ‘Direct Viable Count’ method148 and in any number of other tests that detect metabolic activity136,149). And finally, as we shall see in the next section, careful methods of resuscitation/cultivation do indeed allow a very significant fraction
of organisms that can be isolated from a variety of environments (e.g. the gut) to be resuscitated and to grow very effectively.

**Not-yet-cultured bacteria are mainly dormant and thus resuscitatable**

As indicated in the introduction, it is now well established that even laboratory cultures, that from a macroscopic point of view are growing exponentially, contain subpopulations of non-growing cells. These cells are dormant by definition, because they may later be resuscitated and grow. It is easy to ascribe an evolutionary advantage of this culture differentiation from the perspective of the benefits of having a sub-population that by not growing is more resistant to environmental stresses (e.g. 154–156). Indeed, this general kind of phenotypic differentiation strategy, in which the variance in reproductive rate is traded off at the expense of the mean, has been referred to as bet hedging and is actually adaptive. An important point here is that in many natural environments, asexually reproducing organisms such as bacteria are likely to be (spatially) close to their ancestors and descendants, such that inclusive fitness theory implies that it is entirely reasonable for them to behave altruistically, e.g. by ‘bet hedging’. This is also discussed further below.

It is also reasonable that in isolated (closed) natural environments, nutrients and thus sources of energy must be exhausted at some point, and thus for simple energetic reasons multiplication becomes impossible and a dormant state likely (if later resuscitation proves it to be so). Similarly, it is likely that in the absence of energy, nutrients and/or signalling molecules, and based on more ecological or community considerations (e.g. 172–175), it is necessary to add any or each of them to ‘prime’ bacteria to resuscitate. This has indeed been shown, including for sources of energy, iron-acquiring compounds ( siderophores), cell wall muramides, and various signalling molecules (especially pheromones) that exist in natural environments. We note too that ‘kick starting’ dormant cells may require the synthesis of transporters (a neglected clade) necessary for the uptake of all kinds of molecules. Overall, the idea that most bacteria that may be observed in the natural environment are ‘unculturable’ is incorrect.

Finally here, and though this is obvious it is well worth rehearsing, the simple fact that we can store non-growing microbes under desiccated or frozen conditions or as agar ‘stabs’ in culture collections for extended periods means that most microbes are certainly well adapted to entering and leaving dormancy.

**Pheromonal proteins**

A related and unexpected discovery came from analyses of starved laboratory cultures of the actinobacterium *Micrococcus luteus*, in which almost all cells lost culturability. However, they were not dead but dormant, as they could be resuscitated by using a combination of weak nutrient media and a signalling molecule found in spent culture supernatants. The original studies used flow cytometry to discriminate the physiological state of individual cells (see also 211,212). By using another ‘single cell’ assay based on dilution to extinction (that avoids artefacts connected with the regrowth of ‘initially viable’ bacteria), we were able to purify the signalling molecule. It turned out to be a protein, named Rpf (for ‘resuscitation-promoting factor’). In *M. luteus* there is only one homologue, and the gene (product) is essential for both resuscitation and multiplication. Rpf contains a highly conserved 70 amino acid ‘Rpf domain’ and is widely (and probably ubiquitously) distributed throughout the actinobacteria, but with examples elsewhere. Most organisms that have a homologue have more than one. Thus *M. tuberculosis* has five homologues. Rpf can have peptidoglycanase and muralytic activity and known crystal structures are consistent with this. These activities can certainly account for at least some of the resuscitation-promoting properties. As an extracellular protein that may be required for growth, and with a high level of immunogenicity, it is obviously an excellent candidate target for inclusion in appropriate vaccines against pathogenic actinobacteria. It is also more directly of potential utility in stimulating bacterial communication and resuscitation in a variety of cultures in both samples taken from Nature and in the laboratory.

**Culturability, dormancy and persistence in laboratory cultures of non-fastidious bacteria**

Having established the frequency of occurrence of microbial dormancy in the natural environment, it is of interest to understand better the mechanisms by which microbes might effect this dormancy and potential resuscitation. Unsurprisingly, microbiologists have turned to *E. coli*, and considerable progress has been made.

The starting position is as in Figure 1 and Figure 6, to the effect that at any given moment in a typical culture a small fraction of the population is non-growing, and thus potentially dormant. Since clearly the same fraction cannot (or is wise not to) remain in dormancy indefinitely in the presence of suitable nutrients that permit the growth of its siblings, we must invoke at least one mechanism that can cause the bacteria to ‘oscillate’ between growing and dormant states. Many simple gene expression network topologies admit this behaviour, including a simple feedback loop with delay, and we note that even whole cultures can exhibit oscillations and deterministic chaos. While flow cytometric observations show that even ‘homogeneous’ laboratory cultures show highly heterogeneous distributions in cellular volume (not just between X and 2X) and expression profiles (and see 289), our particular focus will be on ‘binary’ or ‘ bistable’ systems in which individual cells either are or are not operationally cultivable.

Experimentally, it is also common to assess the phenotypic ability of subpopulations of cells to tolerate normally inhibitory concentrations of bactericidal drugs, this being a marker for that fraction of cells that is ‘persistent’ (and maybe dormant) at the stage in question. Note that the persistence phenotype is not induced by the drugs. Changes or transitions in the state of a particular cell in a population between the various phenotypic states is a phenomenon that may be (and is commonly) referred to as ‘phenotypic switching’.
out potentli by a second gene product that thereof undoes the inhibition of proliferation, can have the effect of phenotypically switching cells between dormancy and growth. This seems to be precisely what is going on, and such pairs of gene products have been referred to (somewhat misleadingly) as toxin-antitoxin (TA) pairs. One such involves the well-known ppGpp metabolic system that can serve to inhibit DNA gyrase, and points to the fact that in these circumstances, persisters may be quite metabolically active, even if transiently incapable of reproduction. Another phenotype switching mechanism, underlying colony phenotype switching, comes from metabolic bifurcations driven by the levels of a particular metabolite.

Any mechanisms that permit cells to communicate with each other can amplify switching effects by cell synchronisation, and by definition such ‘social’ signals act as pheromones, whose apparent ‘altruisms’ can be explained on the basis of kin selection theory. There is considerable interest, largely outwith our scope here, in ‘altruism’ can be explained on the basis of kin selection theory. Such ‘social’ signals act as pheromones, whose apparent ‘altruisms’ can be explained on the basis of kin selection theory.

The existence of bacterial DNA in even ‘healthy’ blood has long been known, and since naked DNA would be degraded and living cells would soon kill the host, the (seemingly) obvious conclusion that the prokaryotic DNA must reflect occult, and potentially dormant bacteria can represent as much as 99% of the organisms that may be present. As noted above for environmental microbiology, dormant bacteria can represent as much as 99% of the organisms that may be observed microscopically or by macromolecular sequencing, but they too have special adaptations to an intracellular lifestyle (e.g. .)

### Classical clinical microbiology of culturable organisms

Until relatively recently, almost all of clinical microbiology was based on rather classical methods of plate counting, coupled to assessment of antibiotic sensitivity. Various means of automated blood culture that assess metabolism exist (although they require typically 48–72h to show a ‘positive’). Positive tests, often implicitly involving culture (and not just metabolism) within the assay, would be followed by other tests seeking to identify the organisms detected, nowadays typically by nucleic acid sequence-based methods. However, these and other tests for the presence of antigens or even antibodies cannot speak to the question of cultivability (and of course antigens such as lipopolysaccharide (LPS) are shed by dying cells). This said, it makes little sense to try to culture microbes from samples that molecular sequencing methods indicate lack them, so the molecular methods always provide a useful starting point for seeking to resuscitate any resuscitable (hence operationally dormant) microbes that might be present.

The existence of bacterial DNA in even ‘healthy’ blood has long been known, and since naked DNA would be degraded and living cells would soon kill the host, the (seemingly) obvious conclusion that the prokaryotic DNA must reflect occult, and potentially dormant, cells seems neither to have been drawn nor acted upon.

### Some well-established cases of dormancy in clinical microbiology

The idea that (typically intracellular) dormancy is a major component in infectious diseases (including in the absence of antibiotics that may serve to light up ‘persisters’) is of course well-established, and the main purpose of this brief section is simply to remind readers of this. Such a reminder serves as a prelude to a longer discussion of the very many clinical circumstances where we consider that the role of dormant microbes is not widely appreciated, and where they are not really considered to involve a communicable or microbial component at all. Thus Table 1 shows a few organisms (and references) for which we consider that most readers would regard the idea of and evidence for dormancy as more or less uncontroversial. We do not include disease-causing infectious agents where they are better known for their ability to persist in the natural environment. Organisms such as Legionella pneumophila that represent significant public health issues, fall into this category, and Legionella and other persisters (in environments such as water system biofilms) are indeed well known (e.g. ), although they too have special adaptations to an intracellular lifestyle (e.g. )

### Generalised failure of classical techniques to detect dormant bacteria in clinical microbiology

As noted above for environmental microbiology, dormant bacteria can represent as much as 99% of the organisms that may be observed microscopically or by macromolecular sequencing, but classically (and by definition) they are not enumerated by culture-based methods that determine ‘immediate culturability’. Such culture-based methods are also widely used in clinical microbiology. However, if we were to plate out 100 µL of a culture containing 200 bacteria mL⁻¹, of which 99% were dormant at any instant, we would expect (based on a Poisson distribution) to see fewer than 1 propagule or colony-forming unit per sample. We have noted above that it can be determined by sequencing that many of the non-cultured environmental organisms largely differ from those in standard culture collections. Certainly the examples given above in clinical microbiology, such as Tropheryma whippelii, were both observed microscopically and were sequenced prior to being brought into axenic culture.

The PCR method is exquisitely sensitive (down to one cell or propagule per sample), and we note that contamination artefacts from the PCR reagents represent a real issue that must always be checked (e.g. ). We have rehearsed elsewhere five classes of argument that collectively make it implausible that these are all contamination artefacts; probably the most persuasive is simply the sheer number of prokaryotic DNA molecules that can be measured in blood and serum (e.g. ). While some of the most recent nucleic acid sequencing methods (e.g. ) do operate on single molecules, and genome-wide sequencing may soon be routine (e.g. ), the analysis of prokaryotes usually used a broad-range PCR step to amplify small-subunit rDNA to assess their presence, whether in environmental or clinical samples. Using this, and while these methods alone cannot tell whether they were operationally dormant or dead, a very considerable number of studies have been performed in which ‘culture-negative’ clinical samples showed the presence of prokaryotes (at least as judged by sequence-based methods). This has some profound consequences.

We note that in a steady state such cells must be supplied at a rate equal to that of their clearance, and that the fact that clearance is lower than probably expected implies a significant ability of such cells to evade the innate and adaptive immune systems. We also take it that at least for common organisms (not very slow growers such as certain mycobacteria) the former rates must be much lower than those typically attainable in laboratory cultures, else we would have classical sepsis, and we do not. Most likely the observable facts are best accounted for by a combination of a periodic resupply of resuscitating cells, coupled to physiological changes in
non-growing cells (especially including of cell wall antigens) that help them evade natural clearance mechanisms.

**Broad-range PCR methods indicate the widespread presence of prokaryotic DNA in culture-negative clinical samples**

While PCR-based methods have long been used to assess the species involved in culture-positive samples\(^{406}\), e.g. from blood, our interest here is in samples that are culture-negative\(^{407}\) that may yet (and indeed likely do) contain dormant cells. Among the first such indications of this was the study by Relman’s group\(^{380}\), who showed that the blood of even healthy controls contained significant amounts of prokaryotic DNA. Table 2 lists some studies in which broad-range PCR has been used to amplify and detect prokaryotic rDNA in culture-negative samples.

In environmental microbiology, as mentioned above, there were many early indications (as observed microscopically or flow cytometrically) for the presence of bacteria that did not (or not easily) prove resuscitable or culturable. In a similar vein, many studies have shown microscopically observable organisms in culture-negative but disease-positive samples. This is true both for diseases considered to be due to microbial pathogens and, in fact, for many others normally considered non-communicable\(^{62}\).

**Microscopically observable and potentially dormant bacteria in clinical disease**

Microscopic observations in tissues have been a major part of the discovery process by which certain bacteria were indeed identified as the cause of various diseases. Billings\(^{331}\), Price\(^{332}\), Domingue\(^{413,433-435}\), Mattman\(^{436}\), Ewald\(^{437}\) and Onwuamaegbu and colleagues\(^{438}\) review the extensive and largely forgotten early literature. Domingue and Schlegel\(^{439}\) also mentioned that they could recover culturable bacteria, probably mainly from L forms (see 62,436,440), from lysates of normal and diseased blood. It was to be assumed that these cells were not replicating at significant rates in the blood itself. However, we can find no evidence that this was ever followed up. Our own work\(^\) summarised in 62, showed that both bacillary and coccoid cells could be found attached to and within the erythrocytes of patients with Parkinson’s disease and Alzheimer’s disease, at rather greater concentrations than in samples taken from nominally healthy controls.

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**Table 2. Some examples of blood culture-negative but PCR-positive systems, implying the presence of dormant bacteria.** Note that we have sought to exclude examples where anaerobic bacteria could be detected by PCR but not cultured simply because cultures were not anaerobic, and also cases (e.g. 408,409) where high antibiotic concentrations might have prevented culture.

<table>
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<tr>
<th>Aims</th>
<th>Culture-negative but PCR-positive</th>
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<td>Assessment of endocarditis</td>
<td>6 out of 29</td>
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<tr>
<td>Development of broad-range PCR</td>
<td>71 out of 382</td>
<td>406</td>
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<tr>
<td>Development of broad-range PCR; limit of detection 5000 cfu/mL(^{1})</td>
<td>10 out of 103</td>
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<tr>
<td>Improved broad-range PCR method</td>
<td>20 out of 24</td>
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<tr>
<td>Review</td>
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<tr>
<td>Interstitial cystitis</td>
<td>14 out of 14</td>
<td>413</td>
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<tr>
<td>Endocarditis</td>
<td>270 (36.5%) of 740</td>
<td>414 (and see 415)</td>
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<td>Endophthalmitis</td>
<td>116 out of 116 (selected)</td>
<td>416</td>
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<td>General study</td>
<td>18 out of 394 (271 also culture-positive, PCR-positive)</td>
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<td>Bacteraemia in intensive care</td>
<td>48 out of 197</td>
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<td>45 out of 94</td>
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<td>Sepsis/SIRS</td>
<td>29 out of 59</td>
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<td></td>
<td>38 out of 72 culture-positive</td>
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<tr>
<td></td>
<td>14.6% vs 10.3% (no antibiotics)</td>
<td>422</td>
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<td>123 vs 95</td>
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<td>Osteoarticular samples</td>
<td>141 out of 1667</td>
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<td>Review</td>
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<td>Various, including antibiotic-treated</td>
<td>34 out of 240</td>
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<td>Meningitis</td>
<td>26 out of 274</td>
<td>427</td>
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<td></td>
<td>19 out of 21</td>
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<td>Orthopaedic samples</td>
<td>9% out of 125</td>
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<td>Thoracic empyema</td>
<td>14 out of 22</td>
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<td>Trauma</td>
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In a similar way, our preliminary data show that bacteria are visible in plasma, as well as in whole blood smears in various inflammatory conditions. Here we show bacteria in platelet-rich plasma (PRP) taken from a patient with systemic lupus erythematosus and smeared onto a glass cover slip (Figure 7A and Figure 7B). We also show the same from patients with hereditary hemochromatosis (Figure 7C) and type 2 diabetes (Figure 7D). We also noted microbiota associated with erythrocytes in thromboembolic ischemic stroke (Figure 8A and Figure 8B). (Our microscopy methods are as published previously (e.g. 442–451), but fuller publications will appear elsewhere). The ultramicroscopic evidence that these are indeed small bacteria and not say, cellular debris or microparticles (see 452) is presently mainly morphological, though we note the considerable evidence for the presence of bacterial DNA in blood (see previous sections and e.g. 63,340,453).

It is worth rehearsing the very great significance of this. With erythrocytes being present at some 5x10⁹.mL⁻¹ in human blood, even if only one erythrocyte in a thousand harboured just a single dormant bacterium (that would be hard to detect microscopically, but see 453–457), the dormant bacterial load would still be 5,10⁶.mL⁻¹. This is both far from negligible, and serves to exclude the (always potentially worrisome) claim that ‘it is all contaminants’.

### A culturable blood microbiome

A recent and highly significant paper by Damgaard and colleagues bears discussion. These workers note that while bacterial growth can normally be elicited during sterility testing in vitro from fewer than 1 in a 1000 blood units (459–461), transfusion-transmitted infections occur with a very much higher frequency (more like 10–12% (462), or even more (464)), and are responsible for a high fraction of transfusion-associated deaths (465–467). Although it was acknowledged that venepuncture-associated contamination or an effect of transfusion in suppressing the immune system might contribute, it was also recognised that one means by which to account for this would be that ‘normal blood’, and in particular its erythrocyte components, might

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**Figure 7.** A and B) Platelet rich plasma (PRP) from a patient with systemic lupus erythematosus (SLE). A) Platelet with bacteria visible in the surrounding smear (pink arrows); B) areas in smear with bacteria (pink arrows); C) Erythrocyte with associated bacteria from patient with confirmed hereditary hemochromatosis D) Erythrocytes with bacteria from patients with diagnosed type II diabetes. A–C Scale bar: 1 μm and D 400 nm.

**Figure 8.** Bacteria in whole blood from a patient with thromboembolic ischemic stroke A) Microbiota in whole blood; scale bar: 200 nm. B) Erythrocyte with bacteria; scale bar: 1 μm.
also contain infectious agents that might be able to grow post-transfusion. Indeed, these authors found that under anaerobic conditions a small number of colony-forming units (ca 4–5.10^{6}) could be recovered by direct plating from fully 62% of blood units, with ‘controls’ producing an average of just 1 cfu.mL^{-1}. More of the bacteria were associated with red blood cells than with plasma, and rDNA was used to identify them. These data are entirely consistent with the idea that dormant bacteria are present in the blood of even ‘normal’ individuals (note that periodontitis was not a criterion for donor exclusion here), that they are probably lurking in or on erythrocytes, and that they can be resuscitated and grow under the correct conditions.

### Evidence for a microbial component in a very large variety of ‘non-communicable’ diseases

We have surveyed the literature for evidence in which a microbial component has indeed been observed to be an accompaniment of, and probably a major contributory factor to, a variety of (typically inflammatory) diseases that are normally considered ‘non-communicable’. Rarely has the physiological state of these microbes been considered, but since it would be obvious if they were growing, it is most likely that they are indeed dormant. Table 3 summarises these highly extensive associations. While some are just associations, and we could have extended this table considerably, some studies (e.g. [470]) contain very detailed aetiological arguments that

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<td>17 patients with relapsing-remitting MS, 20 patients with progressive MS, and 27 patients with other neurological conditions. Bacterial present in the cerebrospinal fluid.</td>
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<td>Chlamydia (Chlamydophila) pneumoniae</td>
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<td>PCR, Serology Many patients studied: cerebrospinal fluid</td>
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<td>Blood &amp; tissue culture, patient records Hypocomplementaemia and infection with encapsulated bacteria</td>
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<td>Vasculitis</td>
<td>Possibly mainly viral, but bacteria include Staphylococcus aureus, Treponema pallidum, Rickettsiaceae, Borrelia burgdorferi, M. tuberculosis</td>
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### CARDIOVASCULAR DISEASES

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<td>Antigens, PCR and treatment of patients with antibiotics with good results</td>
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**OTHER INFLAMMATORY CONDITIONS**

<table>
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<th>Disease</th>
<th>Class of bacteria</th>
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<tr>
<td>Preeclampsia</td>
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<td><em>Hafnia alvei</em>, <em>Pseudomonas aeruginosa</em>, <em>Morganella morganii</em>, <em>Proteus mirabilis</em>, <em>Pseudomonas putida</em>, <em>Citrobacter koseri</em>, <em>Klebsiella pneumoniae</em></td>
<td>Serum IgA and IgM against LPS Serology</td>
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<td></td>
<td>Mycoplasmal infections (M. pneumonia, M. fermentans, M. hominis, M. penetrans), Chlamydia pneumonia, Human herpes virus-6</td>
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<td>IgG is patient blood</td>
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<td>Vitamin D receptor</td>
<td>Cell wall deficient bacteria</td>
<td>Evade immune destruction by invading nucleated cells where they persist in the cytoplasm. From here they down-regulated the vitamin D receptor</td>
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<td>(VDR) dysregulation</td>
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<td></td>
<td>Multiple organisms, including mycobacteria, <em>Borrelia</em></td>
<td>Paper discusses a model describing how multiple species-bacterial, viral, and fungal-can cumulatively dysregulate expression by the VDR nuclear receptor</td>
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<td>Antiphospholipid</td>
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<td>General review paper reviewing co-infections</td>
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<td>Toxoplasma</td>
<td>Anti-Toxoplasma antibody screening in 98 patients with antiphospholipid syndrome</td>
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leave little room for doubt. Overall, the sheer size of the Table does strongly indicate the commonality of many of the microbially based mechanisms underpinning or accompanying various autoimmune and inflammatory diseases. In conditions such as atherosclerosis, transient ischemic attacks (TIAs), and stroke, it is very easy to conceive how resuscitating bacteria might serve to block the flow of blood, for instance. At all events, our main point here is that the evidence for a microbial contribution to many diseases supposedly lacking a microbial component is both multi-factorial and very considerable. Indeed, the purpose of a synthetic review such as this is to provide such pointers for more detailed studies in individual cases. Our specific interest is with the chief mechanisms by which these supposedly dormant bacteria might resuscitate and act as triggers of disease.

**Relation between iron dysregulation, sepsis and other comorbidities**

Many of the diseases in Table 3 are precisely those inflammatory diseases that we have listed before as coupled to iron dysregulation183,184,452,733. A consequence of our analysis is that iron dysregulation and sepsis (as judged either by genuine infection by culturable bacteria or their inflammatory products such as LPS) should be associated causally with these various other diseases.

This leads to a variety of predictions and postdictions that we rehearse. A purposely simple (and simplistic) indication of a plausible chain of events (for which each step is underpinned by substantial evidence) is given in Figure 9, both in general terms (for unspecified diseases) and for a couple of steps to type 2 diabetes. Figure 9 aims specifically to highlight the relationship between the ability of available iron to stimulate bacterial growth and the potential disease sequelae thereof.

**Iron and sepsis**

First of all, it is well established that free iron may be raised in sepsis and related conditions183–184, as may serum ferritin183–184 (that has mainly dumped its iron185). We have here argued that this is likely to be a significant contributor to the relationship between overt or cryptic infection and the many iron-related inflammatory diseases discussed here and elsewhere183,184,452,733. Note that patients suffering from iron overload diseases such as hereditary haemochromatosis are especially susceptible to infection (see e.g. 748–750 and Table 3). Certainly the idea that iron-related metabolism and siderophores are virulence factors (e.g. 751–763) is established unequivocally. In many diseases (e.g. lupus764,765 or type 1 diabetes766) it is considered that patients with the disease are more prone to sepsis, but we suggest here that (as with stroke761,763,765,766) it may more likely be the converse that is truer: patients suffering from latent infections are in fact more prone to acquiring, having, or exacerbating the state of these other conditions, in a vicious cycle (see Figure 9).

**Role of iron chelation in preventing sepsis**

This was discussed at considerable length previously184, and that discussion is not repeated here (though a few more recent and pertinent references include776–779). However, while (perhaps surprisingly, given what we see as the evidence) it does not even appear in the guidelines750, there is considerable evidence98 that appropriate iron chelation slows, inhibits or overcomes sepsis. We note, however, that some chelators are in fact known iron siderophores, and such molecules may assist the pathogen (e.g. 781–783) and are to be avoided. On this basis, iron chelation may be a suitable alternative to antibiotics in preventing multiple inflammatory diseases (and such chelation may be nutritional rather than pharmacological in nature, e.g. 183). However, it is clear that we also need to learn to kill ‘dormant’ bacteria, and this usually requires that they are growing.

**Utility of antibiotics in treating non-communicable diseases**

It is well established that the re-use of protein motifs in natural (and directed98) evolution means that most drugs, especially the more lipophilic ones, are promiscuous in the sense that they bind to multiple targets183,785 (on average six known ones for marketed drugs786). This said (and while we are very far from wishing to encourage the unnecessary use of antibiotics), the prediction here is that appropriate antibiotics will prove to have clinical benefit in diseases commonly seen as non-communicable. This is certainly known to be the

<table>
<thead>
<tr>
<th>Disease</th>
<th>Class of bacteria</th>
<th>Nature of the evidence of involvement</th>
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<tbody>
<tr>
<td>Sudden Infant Death Syndrome</td>
<td><em>S. aureus</em></td>
<td>Review papers: seasonality, bacteriology; Papers discuss markers of infection and inflammation are often found on autopsy along with microbial isolates</td>
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<td>Migraine</td>
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<td>A randomized, double blind, controlled trial</td>
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<td>A meta-analysis of research between 2000 and 2013</td>
<td>732</td>
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</table>
A generalised scheme for microbial/iron-driven inflammatory disease, illustrated with Type 2 diabetes

Figure 9. An elementary systems biology model of how iron dysregulation can stimulate dormant bacterial growth that can in turn lead to antigen production (e.g. of LPS) that can then trigger inflammation leading to cell death and to a variety of diseases. While it is recognised that this simple diagram is very far from capturing the richness of these phenomena, there is abundant evidence for each of these steps, but sample references for the numbered interactions are (1) 184–186, (2) 285, 473, 475, 482–485, (3) 473, 475, 477–479, (4) 473, 475, 477–479, (5) 473, 475, 477–479, (6) 473, 475, 477–479, (7) 473, 475, 477–479, (8) 473, 475, 477–479, (9) 473, 475, 477–479, (10) 473, 475, 477–479.

Concluding comments: on the systems properties of dormancy and virulence

We have here brought together some of the relevant elements of environmental, laboratory, and clinical microbiology. We have argued that while their languages may differ (e.g. ‘dormancy’ vs ‘persistence’), very similar phenomena have been observed in each of these spheres (plausibly underlying a commonality of mechanism). Certainly the ability to culture microbes, and not merely to observe them (whether microscopically or via their macromolecular sequences or chemical products), remains an important goal of basic microbiology. This is likely to have significant payoffs in bioprospecting (e.g. 179, 806). However, we are sure that improved methods of detecting and identifying these dormant bacteria, whether this is done via chemical imaging, through macromolecular amplification and/or sequencing, or through resuscitation and culturing, will have a major role to play in increasing the awareness of their existence and importance.

Clearly dormant and/or persistent bacteria are likely to be relatively avirulent when they are in such dormant states, and able to bypass the attentions of the innate immune system (albeit the production of superantigens by at least some microorganisms may be what triggers autoimmune diseases). This ‘stealth’ antigenicity is probably why they have been largely unnoticed by us too, and their routine estimation via molecular methods seems highly desirable. Indeed, virulence varies widely between individual strains (e.g. 811, 812). Modern molecular microbiology places much emphasis on the virulence of the pathogen, with concepts such as ‘pathogenicity islands’, ‘virulence genes’, and the ‘virulome’ being commonplace. However, if dormant microbes resuscitate (or are to be resuscitated) in vivo we shall need to pay much more attention to the environmental triggers that can cause this to happen than we probably have so far (given that the pathogen genotype is fixed). In other words, virulence, like dormancy, is a phenotypic as well as a genotypic property. We remain largely ignorant of the means by which an optimal immune system has been selected for (or against) by longer-term evolution on the basis of microbial exposures in early life, and how this may have changed with more recent changes in human lifestyle. Nor do we understand
how such microbes might enter and exit blood cells (and see 62,347, 829–833) (albeit the known endosymbiotic origins of eukaryotic organelles must have presaged such mechanisms). Similarly, we do not yet know what may cause these dormant microbes to resuscitate (and/or to exit their intracellular niches). However, the potential for iron-associated replication and (e.g.) LPS production and shedding does provide a very straightforward explanation for the continuing low- or medium-grade inflammation characteristic of many inflammatory diseases we have considered here and elsewhere (83,184,449,452,733,890) (Figure 9).

Recognising that correlation does not at all equate to causality (e.g. 195,836), one approach to Science is based on varying independently something considered a cause and observing its predicted effects (e.g. 195,837,838). Temporal covariation of measurands can also be performed. The levels of free iron are clearly one such possibility. To assess causality in microbiology it is usual (e.g. 815, 839–841) to invoke what are (variously) referred to the Henle-Koch or Koch’s postulates. These are based on the nature and presence, but not the physiological state, of an agent that might be believed to ‘cause’ (or at least contribute to) an infectious disease. Consequently, dormancy poses something of a challenge to the full completion of the required tests. Indeed a number of authors (847,313,42–444) have recognised that these tests may need revision in the light of the ability to identify disease-causing microbes by sequence alone. We suspect that a key element here will be the ability to resuscitate dormant organisms in vivo and to see the effects of that on clinical disease.

From a ‘philosophy of science’ point of view (e.g. 841), one strategy taken to develop the evidence for a particular point of view hinges on the idea that if a series of ostensibly unrelated findings are brought together into a self-consistent narrative, that narrative is thereby strengthened. This is the strategy pursued here, and it is known as ‘coherence’ (846–446).

As phrased by Silvers (649), “Several of our contributors showed how discoveries and insights could emerge with what seemed great promise, and yet be pushed aside, discarded, and forgotten – only to re-emerge once again, sometimes many years later, and become, in their new formulation, accepted as important”. In this sense, and as presaged in the opening quotation, it seems that ideas, as well as bacteria, can remain dormant for extended periods (850,851).

Author contributions
This review originated as part of a discussion between the corresponding authors, who have a funded collaboration as outlined under ‘grant information’, and was partly written during a visit of EP and MP to Manchester. All authors contributed to the writing of the manuscript.

Competing interests
No competing interests were disclosed.

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This review paper is an important modern perspective on bacterial persistence and expression of disease. The role of ‘stressed’, atypical, cell wall-defective, cryptic, pleomorphic forms in chronic inflammatory diseases in clinical medicine and especially in diagnostic pathology and clinical microbiology is grossly neglected and overlooked. This paper warrants publication for its timely approach to a vastly important overlooked topic in science and medicine and its relevant, useful 890 cited references. I see no point in further elaboration on semantics: persistence vs. dormancy. In my opinion, regardless of terminology preferred (or debated) the relevant and important fact is identity of atypical forms in tissues, their basic biology and their relationship to disease.

Although there is provocative circumstantial evidence linking pleomorphic forms suspected of being bacterial in origin in a wide array of chronic diseases, many were categorized as autoimmune, unrelated to microbes (unambiguous proof lacking). While the persistence of stainable forms (various structures) are often seen in tissue specimens utilizing histopathologic and bacteriologic stains, most are discarded as insignificant staining artifacts and debris, and especially in the absence of non-cultivable bacteria from accompanying specimens. In my opinion, this is a primary reason the significance of such stainable findings has been ignored in clinical medicine and has stymied their identity as causative agents of disease. Furthermore, even when there may be growth of atypical bacterial forms on artificial culture media, bizarre (non-standard) morphologic, biochemical, physiological characteristics of the isolated organism \textit{in vitro}, the findings are most often disregarded as “contaminants”.

Permit me to digress and cite such an example of an unidentified pleomorphic form isolated from patients with interstitial cystitis (a chronic, debilitating disease of unknown origin). These atypical isolates were subjected to elaborate, microbiological, immunological, biochemical, physiologic and electron microscopic characterizations. The findings (data) were presented at the American Urological Association annual meeting, only to have a well-known academic urologist and interstitial cystitis specialist congratulate the researchers for the elaborate experimental description of an ‘artifact’. Obviously with that type of unexpected and disappointing comment, there was nothing more to be said, other than, thank you! So there you have it: The regrettable dismissal of a potentially important finding in a disease of unknown origin by an individual who could not see the forest through the trees and without evidence to substantiate the claim that the finding was an ‘artifact’.

The 14 topics and subsections outlined in Figure 4 of the review set the stage for the “dramatics” – ‘mind map’ – that follows. In itself this graphic may have been enough when accompanied by germane references instead of lengthy written discussions for each topic since much is \textit{déjà vu}, gleaned over a
period of many decades from published findings. On the other hand, it is often useful to repeat, for emphasis, and especially to call attention to neglected topics, which I suspect was the intent. Although the tables and graphics are worthwhile, it does take time to digest it all, meaning it may have been possible to shorten the paper.

Two important publications (not cited in list of 890 references): companion papers by Green *et al* in *Infection and Immunity*, 1974, Oct; 10 (4): 889-914 and 915-927; demonstrated the phenomena of microbial persistence and reversion with *Streptococcus faecalis* L-forms in human embryonic kidney cells, followed by a proposed reproductive cycle for a relatively stable L-phase variant of *Streptococcus faecalis*. I call these publications to the attention of the authors because of their possible application to the fundamental basis of persistence by ‘stressed’, atypical bacteria in chronically diseased human subjects. Essential to the thesis of Green *et al* is that small, electron dense, non-vesiculated L-forms were shown to be the central (core) element in bacterial persistence in these experimental studies. The researchers concluded that depending on the stimulus received, these dense forms might be considered as undifferentiated cells, with the capacity to develop along several different routes. *In vitro*, the dense form was observed to divide and bud rapidly. In addition, the dense forms appeared to be capable of growth and development within vesicles of mature mother forms. When these forms were released from the vesicles into the surrounding fluid medium, further growth occurred, resulting in the development of immature and ultimately mature mother forms. Under conditions unfavorable for L-form growth, these dense forms developed first into transitional forms and then into the bacterial form. These dense forms might therefore be considered as undifferentiated ‘stem cells’ with the capacity to develop along several different routes, depending upon the stimulus received. Hence, in applying these findings to altered forms created *in vivo* (humans) these may take up intracellular and/or extracellular residence; possibly establishing a sort of immune protected parasitic relationship persisting/surviving phagocytic action, and creating subtle pathologic changes in the host during a prolonged period of tissue persistence. This might translate into an etiology for chronic inflammatory diseases, when the ‘stressed’ bacteria increase in numbers and overwhelm the normal biological functions of the host. I further propose that *in vivo* persistence of these bacterial elements escape immune surveillance partially, completely, or may integrate with host cell organelles to create bacteria-host cell-antigen complexes which could provoke immunopathologic consequences. Highly relevant, recently published data on modifications of gene expression, modes of division for stressed bacteria, and the paradoxical finding of peptidoglycan in L-forms are pertinent to the hypothesis that atypical, pleomorphic bacteria are the organisms responsible for persistence and expression of disease. Finally, it is hoped that the Kell, Potgieter, Pretorius timely, interesting and provocative review will call attention to this highly significant, too often overlooked subject. In my opinion, this review calls for a scientific/medical challenge: 1) to motivate visionary scientists and clinicians to investigate the fundamental origins of bacterial persistence in chronic diseases; 2) to unambiguously identify tissue persisting forms utilizing modern molecular technology, and 3) to design elegant experiments to provide convincing scientific proof (or disprove) that extracellular and or intracellular stainable bodies observed in histopathologic specimens and dense bodies at the electron microscopic level (culture negative) are bacteria existing as ‘stressed’ altered forms in tissues and not tissue or staining artifacts. Proof of the above hypothesis would open new arenas in clinical diagnosis, management and treatment of numerous chronic inflammatory human diseases of unknown etiology and might even extend to a bacterial cause for certain malignancies (as previously proposed many decades ago).

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

**Competing Interests:** No competing interests were disclosed.
I review Kell et al’s review relating to individuality, phenotypic differentiation, dormancy and “persistence” as a clinical microbiologist, infectious diseases doctor, with an interest in developing and assessing the impact of rapid sequence-based molecular blood and lung diagnostics in the critically ill.

This review reminded me of Mussorsky’s *Pictures at an Exhibition*, a collection of hastily composed pieces whose theme was to take an interested individual through an art gallery, and to tarry awhile in front of 10 *Tableaux*, interspersed with musical elements referring to the “Promenade” through the gallery.

And so it is with Kell et al’s review. After an introductory *Promenade* relating to matters of bacterial dormancy and its relationship with just about any other conceivable physical state between life and death, exhaustively referenced together with the thought provoking Postgate-ian concept of the difficulties inherent in differentiating bacterial life from death if you only have an instant in time to measure it – we are then presented with several pictures, garlanded for us in extensively referenced detail by the authors. Were mindmaps not enough to capture the reader’s curiosity as to this *magnum opus* of a kind, we are invited to walk through Kell et al’s gallery of mental pictures depicting scenes of the Yet to be Cultured, Those bacteria that aren’t culturable yet but are certainly not dead, the biological importance of bacterial pheromones, the evils of Iron - thence to the Clinical Microbiology Room of Pictures with a liberal helping of systems biology throughout.

I am a proponent of, and believer in, the present and future potential of Nucleic Acid Technology (NAT) for pathogen detection in Clinical microbiology and I use such techniques on a daily basis. When appropriately deployed, it allows me to find those “unculturable” pathogens as drivers for individual clinical cases of infection. Perhaps strangely, this is a relatively new paradigm for most practising clinicians, and one which likely will generate fundamental discoveries highly relevant to human disease, and for all we know as equally important as *Helicobacter*. That such sequences should be found in blood is hardly surprising, given that human beings have between 10 and 100 times more bacterial cells than their own, living (or persisting, or dormant) on and in them. This groups’ demonstration of bacteria adhering to red cells (also in red cells) is certainly very intriguing, and such suggested “atopobiosis” is more expansively dealt with in another publication and prompts far more questions than it answers – in a good way. Another obvious question relates to how these adherent bacteria may remain undetected and intact in the presence of numerous moieties central to both innate and acquired immunity (complement and antibody to name but two) as well as escaping phagocytosis in the liver and spleen. It would certainly be interesting to look at red cells in the grave condition of erythrophagocytosis, a condition whose mechanism is in most cases obscure –it might even be that adherent bacteria “opsonize” the red cells in these cases. This reader, however, does baulk at the very serious work to be done as regards untangling the mechanistic nature of an “association” with several diseases, and certainly at this stage it would be very unwise to suggest it’s anything more than that. Further work of this nature should be approached and undertaken
with extreme caution and rigor in view of the myriad possible explanations other than causative ones; the Measles vaccine/autism saga comes to mind here.

It is likely therefore that such technologies will perforce "lift the lid" on what might lie beyond the Culturable, and its relationship to human disease. This is explored in Table 3, which represents a tour de force as concerns the sheer volume of references relating to all that appears to associate human Disease and organisms, mostly bacteria.

Unfortunately, this Table doesn't work for me. Whilst it will serve me as a unique and accessible resource of information in this space, it is anarchic. Correctly described as “Evidence for agents in non-communicable diseases”, it lists, in no particular order, and with no apparently critical eye, references 470 to 712 as relevant to the Table subject stated above. This list's breadth as concerns both organisms and clinical diseases is extraordinary; and the literature quoted in a table described as "effect of bacterial involvement" ranges from unusual cases, to mechanistic assumptions of what LPS might do, to the concept of “dysbiosis” amongst many others. I was left rather dizzy from the mental exercise needed to constantly adjust to the sheer scale and variation of why a particular organism, or something it produces, might either directly causally relate to a particular disease, or perhaps through the individuals’ immune response to it; especially now we know how outbred we are as concerns immune responsiveness.

This review finishes with an impressive and lyrical chiding for Scientists, whereby those who research this field should wake up from their intellectual slumber, as might and indeed do bacteria.

This review is additionally peppered with tantalizing if perhaps sometimes unfounded assumptions, some arguable and some bordering on plain unreasonable. Certainly my eyebrow raising went into overdrive when considering Kell's conviction as concerns a Catholic Grand Unifying Theory based around the Evils of Iron, the subject of a previous equally grand Magnum Opus.

This review has to be one of the most undisciplined I have read in a long time, on occasions associating seemingly disparate observations and conflating “scientifically” determined facts with clinical issues.

Having said this, I should finish by applauding Kell et al’s review as a thumping good read. It's fast paced, edgy, a real treasure trove of papers for me to read at leisure, and goes way outside the usual, expected and conventional boundaries of style of prose and rigor we "normally expect" of such scientific publications. And (warts and all, and there are many) it left this reader thinking that there indeed is Life beyond dormancy within the review’s style itself, beyond the doubtless very important but less imaginative run-of-the-mill, tightly written yet dreary “Scientific Publication”. It is almost as if this review in all its unconventionality were particularly well aligned to the current state of the Art for the Uncultured in Clinical Medicine (bacteria, not Doctors) and its potential to release significant Paradigm shifts. No doubt this reviews’ readers are made up of those who have the capacity to appreciate Kells' latest brand of emergent, imaginative systems biology style of thinking underneath what some might consider a publication of inadequate scientific rigor.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

**Competing Interests:** No competing interests were disclosed.
"I review Kell et al’s review relating to individuality, phenotypic differentiation, dormancy and “persistence” as a clinical microbiologist, infectious diseases doctor, with an interest in developing and assessing the impact of rapid sequence-based molecular blood and lung diagnostics in the critically ill.

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This is a lovely analogy, which we shall let readers enjoy in the open referee’s report; we are probably not capable of recasting the review in Mussorgskian style anyway! In this regard, readers might also enjoy a little known and whimsical piece on bioinformatics that takes just such an approach: Goble C, Wroe C: The Montagues and the Capulets. Comp Func Genomics 2004; 5:623-632.

"I am a proponent of, and believer in, the present and future potential of Nucleic Acid Technology (NAT) for pathogen detection in Clinical microbiology and I use such techniques on a daily basis. When appropriately deployed, it allows me to find those “unculturable” pathogens as drivers for individual clinical cases of infection. Perhaps strangely, this is a relatively new paradigm for most practising clinicians, and one which likely will generate fundamental discoveries highly relevant to human disease, and for all we know as equally important as Helicobacter. That such sequences should be found in blood is hardly surprising, given that human beings have between 10 and 100 times more bacterial cells than their own, living (or persisting, or dormant) on and in them. This groups’ demonstration of bacteria adhering to red cells (also in red cells) is certainly very intriguing, and such suggested “atopobiosis” is more expansively dealt with in another publication and prompts far more questions than it answers – in a good way. Another obvious question relates to how these adherent bacteria may remain undetected and intact in the presence of numerous moieties central to both innate and acquired immunity (complement and antibody to name but two) as well as escaping phagocytosis in the liver and spleen. It would certainly be interesting to look at red cells in the grave condition of erythrophagocytosis, a condition whose mechanism is in most cases obscure –it might even be that adherent bacteria “opsonize” the red cells in these cases. This reader, however, does baulk at the very serious work to be done as regards untangling the mechanistic nature of an “association” with several diseases, and certainly at this stage it would be very unwise to suggest it’s anything more than that. Further work of this nature should be approached and undertaken with
extraordinary caution and rigor in view of the myriad possible explanations other than causative ones; the Measles vaccine/autism saga comes to mind here.”

These are excellent points, and we have covered some of them in the forward-looking concluding section. While they might be seen as ‘premature’ (in the sense that it requires acceptance of the basic ‘dormancy’ hypothesis in the first place) they do point to important areas where we would seek a mechanistic understanding of what is going on.

- “It is likely therefore that such technologies will perforce “lift the lid” on what might lie beyond the Culturable, and its relationship to human disease. This is explored in Table 3, which represents a tour de force as concerns the sheer volume of references relating to all that appears to associate human Disease and organisms, mostly bacteria.

Unfortunately, this Table doesn’t work for me. Whilst it will serve me as a unique and accessible resource of information in this space, it is anarchic. Correctly described as “Evidence for agents in non-communicable diseases”, it lists, in no particular order, and with no apparently critical eye, references 470 to 712 as relevant to the Table subject stated above. This list’s breadth as concerns both organisms and clinical diseases is extraordinary; and the literature quoted in a table described as “effect of bacterial involvement” ranges from unusual cases, to mechanistic assumptions of what LPS might do, to the concept of “dysbiosis” amongst many others. I was left rather dizzy from the mental exercise needed to constantly adjust to the sheer scale and variation of why a particular organism, or something it produces, might either directly causally relate to a particular disease, or perhaps through the individuals’ immune response to it; especially now we know how outbred we are as concerns immune responsiveness.”

We very much accept the point that the table could be improved with regard to ordering, and we have done so accordingly. However, we think that readers will recognise it for what it is (as does the referee), viz. as a useful resource and/or pointer to a large literature in which specialists in disease X may wish to read at least those papers we suggest as relevant to ‘their’ disease, while others will simply see it as a recognition of the widespread evidence for our more general claims.

- “This review finishes with an impressive and lyrical chiding for Scientists, whereby those who research this field should wake up from their intellectual slumber, as might and indeed do bacteria.

This review is additionally peppered with tantalizing if perhaps sometimes unfounded assumptions, some arguable and some bordering on plain unreasonable. Certainly my eyebrow raising went into overdrive when considering Kell’s conviction as concerns a Catholic Grand Unifying Theory based around the Evils of Iron, the subject of a previous equally grand Magnum Opus.”

As mentioned in the comments on the review of referee 1, the basis for this is the desire to produce a coherent story (in the sense used by Philosophers of Science), and (as referee 1 also states) it is well known that microbial growth in vivo is normally limited by iron availability. That iron dysregulation is also a hallmark of just those chronic inflammatory diseases that we highlight here is consistent with this view, and indeed serves to provide a simple explanation for this. Of course, as the referee indicates (and referee 1 does too),
further demonstrations will benefit from varying iron levels as an independent variable.

- “This review has to be one of the most undisciplined I have read in a long time, on occasions associating seemingly disparate observations and conflating “scientifically” determined facts with clinical issues.

Having said this, I should finish by applauding Kell et al’s review as a thumping good read. It’s fast paced, edgy, a real treasure trove of papers for me to read at leisure, and goes way outside the usual, expected and conventional boundaries of style of prose and rigor we “normally expect” of such scientific publications. And (warts and all, and there are many) it left this reader thinking that there indeed is Life beyond dormancy within the review’s style itself, beyond the doubtless very important but less imaginative run-of-the-mill, tightly written yet dreary “Scientific Publication”. It is almost as if this review in all its unconventionality were particularly well aligned to the current state of the Art for the Uncultured in Clinical Medicine (bacteria, not Doctors) and its potential to release significant Paradigm shifts. No doubt this reviews’ readers are made up of those who have the capacity to appreciate Kells’ latest brand of emergent, imaginative systems biology style of thinking underneath what some might consider a publication of inadequate scientific rigor.”

Many thanks for these last comments; we have nothing further to add here.

**Competing Interests:** No competing interests were disclosed.
In considering the specific points presented I have multiple concerns, the most significant of which I will indulge in outlining below.

Semantics present a central problem in considering bacterial viability and physiology and I broadly support the approach taken here. The authors do try to define their terms but some problems remain. In particular I take issue with the very broad application of term “Persisters” which should be reserved for cells that survive (have the potential to replicate) after exposure to an antimicrobial stress to which kills most cells in an actively growing culture of the organism concerned. Conflation of this term with “Dormancy” implies on the one hand that the persisting cells must have been dormant and on the other that dormancy and persistence represent the same physiological state in bacteria. This difficulty resurfaces later when they define dormancy but other problems emerge before then.

I was next concerned by the extensive use of the term “Differentiation”. I completely agree that what we used to think of as uniform bacterial populations are probably never so but the degree to which subpopulations may be considered differentiated rather than reflecting a range of adaptive responses or indeed, some degree of injury, is not considered here and again I think this leads to problems in considering their hypotheses under a unitary banner downstream. I consider differentiation to require phenotypic changes that are not directly reversible, as in the case of sporulation, whereas adaptation can involve expression of a single gene that can be reversed by its subsequent repression. I do agree that cell cycle contributes to the range of phenotypes in a pure bacterial culture and that this is not the only reason for their diversity (but was not enlightened by use of the term “modulo” in this regard).

The operational definition of dormancy given deliberately leaves open the possibility of metabolic activity and seems only to require that the cell so defined should not divide; this did not allow me to recognise which operational tests might be applied to enumerate or detect dormant cells. Subsequently the detection of molecular signals indicative of bacterial presence in samples from which they were not isolated in culture is taken as evidence of dormancy. In the first case do we accept any non-dividing cell as dormant and in the second I can (and will) offer multiple alternate explanations other than dormancy. Moreover, returning briefly to the conflation between dormancy and persisters, the recent work of John McKinney and colleagues shows that antibiotic exposed persisting cells are not necessarily non-dividing cells in the mycobacterial system he studied.

Alternative interpretations of the presence of bacterial 16SrDNA sequences in blood when culture fails to detect the organisms from which they derive, include the presence of dead, injured or moribund cells. If they are shown to be repeatedly present then they must either be able to persist in the face of clearance mechanisms or be supplied at a rate equal to their clearance; both seem equally plausible to the dormancy explanation to me. Moreover, why the first three explanations offered for “Not-yet-cultured” should apply to environmental bacteriology but not to clinical samples escapes me.

I am led to the conclusion that the authors have chosen to label evidence for discrepancies between culture and nucleic acid detection of bacteria in blood to give their hypotheses a simple headline. I have no problem with the proposal that human blood and tissues classically considered sterile in the absence of overt symptoms of infection are frequently exposed to bacteria and bacterial products that in many cases contribute to serious chronic disease. However, I consider the burden of available evidence currently provides many potential explanations within the field of microbiomics/metagenomics in contrast to the dormancy hypothesis offered here. Further, I feel this broad application of dormancy to bacterial phenotypes which, even in the case of Rpf dependency, have not been shown to result from a programme of gene expression that could be considered as differentiation, diminishes the value of the term. Indeed there remains no direct proof that dormancy of Mycobacterium tuberculosis underpins what
we call latent tuberculosis infection and it is not essential to the observed clinical or pathological pattern, notwithstanding the widespread acceptance of this view by most researches, including me.

I am not fundamentally opposed to the ideas presented by Kell and colleagues but I do not think they are assisted by lack of attention to the contradictions I have identified above.

Finally I come to the iron dysregulation hypothesis and its pro-inflammatory consequences. It is beyond my expertise to comment on the plausibility of the inorganic chemistry deployed here or to review the evidence relating to more than a fraction of the conditions listed. The importance of the struggle between pathogens and host for access to iron is beyond question. When I entered the medical field of infectious disease it was fully recognised that depriving bacteria from iron was a potential therapeutic angle and indeed iron chelation was studied. Desferioxamine, a widely used agent in iron overload, was investigated and found to effectively deliver iron to the pathogen and the approach was set aside. More recently this agent has been identified as a major risk factor in serious fungal infection and guidance specifically recommends its avoidance. Newer agents seem not to suffer from this problem and the approach deserves renewed attention. However, I would not underestimate the ability of pathogens to outwit our pharmaceutical industry in the battle to sequester iron. While there are reasons beyond the host–pathogen tug-of war for iron to consider chelation as a therapeutic option, the potential for adverse effects is significant and I think the suggestion that omission of iron chelation from recent guidance on sepsis management is “shocking” is not justified.

Focussing briefly on the specific diseases cited and their relation to bacterial exposure in one form or another, I find that evidence cited frequently rests on what can be considered “fringe” hypotheses that have little currency in their respective fields. This is not to discourage their continued pursuit but it does weaken the strength of the authors’ argument when investigation of the supporting literature frequently leads to papers that are given little credence in the specialist field. Of course “cave Helicobacter” must remain on the table. But there, an accidental technical breakthrough led to an avalanche of convincing laboratory and clinical data.

In summary Kell, Potgieter and Pretorius have produced an interesting read which bring many important ideas to our attention. I am not convinced of the breadth of conditions to which they argue their ideas are applicable and I await with interest, demonstration of of how they may be practically pursued and some selected definitive proofs that iron-driven inflammatory disease is as important as they claim.

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

**Competing Interests:** No competing interests were disclosed.
samples often reflects dormancy and 2) that such dormant bacteria interact with host iron regulation to contribute to or directly cause a panoply of chronic diseases largely labelled as non-communicable.

At a general level I support the provocative stance taken by the authors. With 861 cited references, at the very least they provide a valuable resource for anyone wishing to consider the potential microbial contribution to diseases traditionally considered free of this aetiological component. Of course Helicobacter infection stands as a monument to the stupidity of dismissing this possibility in the face of carefully assembled evidence. Indeed this reviewer, who many years ago, was presented with a case of duodenal ulcer in his final medical exams, would probably have experienced quite a different career had he claimed a role for infection in causing his patient’s pathology.

In considering the specific points presented I have multiple concerns, the most significant of which I will indulge in outlining below."

Many thanks for the above; it is perfectly accurate and we have nothing to add here.

- "Semantics present a central problem in considering bacterial viability and physiology and I broadly support the approach taken here. The authors do try to define their terms but some problems remain. In particular I take issue with the very broad application of term “Persisters” which should be reserved for cells that survive (have the potential to replicate) after exposure to an antimicrobial stress to which kills most cells in an actively growing culture of the organism concerned. Conflation of this term with “Dormancy” implies on the one hand that the persisting cells must have been dormant and on the other that dormancy and persistence represent the same physiological state in bacteria. This difficulty resurfaces later when they define dormancy but other problems emerge before then."

This is entirely fair; we see that we occasionally elided the terms ‘dormancy’ and ‘persistence’ to imply synonymy, when either there is none or at least there is no evidence for it. We think the best solution is to add a little section pointing out the semantic difficulties, repeating the operational nature of the definitions, and specifying that in very few cases do we actually know the true physiological state of individual cells – which is what matters with regard to replicatory potential. This material mainly appears in the section defining dormancy, and its title has been extended to note the semantic issues.

- "I was next concerned by the extensive use of the term “Differentiation”. I completely agree that what we used to think of as uniform bacterial populations are probably never so but the degree to which subpopulations may be considered differentiated rather than reflecting a range of adaptive responses or indeed, some degree of injury, is not considered here and again I think this leads to problems in considering their hypotheses under a unitary banner downstream. I consider differentiation to require phenotypic changes that are not directly reversible, as in the case of sporulation, whereas adaptation can involve expression of a single gene that can be reversed by its subsequent repression. I do agree that cell cycle contributes to the range of phenotypes in a pure bacterial culture and that this is not the only reason for their diversity (but was not enlightened by use of the term "modulo" in this regard)."

We mainly agree, and suggest what we think is a useful clarification or extension. We note again that “reversibility” is established post hoc, but there are at least two meanings
involved. At one level we are discussing a reversibility of states. Let us take a spore and a vegetative cell, which obviously, for sporulating bacteria, can indeed interconvert (“reversibly”). However, another level or meaning implies a mechanistic reversibility, i.e. the path from A to B is simply traversed in the opposite direction when B reverts or interconverts to A. Not only is this not what we mean but (also for thermodynamic reasons) it is certainly not what is done (sporulation and germination in B. subtilis are definitely quite separate processes, as indicated by the referee, and one is not at all the reverse of the other). We have added clarificatory comments accordingly. (One might also have added, but we have not in the ms as it would distract, that similar issues apply to the ‘reversibility’ of enzymes and of biochemical pathways (gluconeogenesis is not mechanistically a reversal of glycolysis, even if the “start” and “end” states are the same molecules.)

- "The operational definition of dormancy given deliberately leaves open the possibility of metabolic activity and seems only to require that the cell so defined should not divide; this did not allow me to recognise which operational tests might be applied to enumerate or detect dormant cells. Subsequently the detection of molecular signals indicative of bacterial presence in samples from which they were not isolated in culture is taken as evidence of dormancy. In the first case do we accept any non-dividing cell as dormant and in the second I can (and will) offer multiple alternate explanations other than dormancy. Moreover, returning briefly to the conflation between dormancy and persisters, the recent work of John McKinney and colleagues shows that antibiotic exposed persisting cells are not necessarily non-dividing cells in the mycobacterial system he studied."

The hallmark of the dormant macrostate, stated in quotation marks in the second paragraph of the ‘dormancy’ section, is indeed that the cells in question do not immediately grow when attempts to culture them under “suitable” conditions (that normally admit their growth), are often (but not necessarily) of low metabolic activity, but are not operationally dead since they can be resuscitated. On this basis we think that this should allow the referee or anyone else to determine the operational tests. It follows that we do not accept ‘any’ non-dividing cell as dormant since only resuscitable cells can – post hoc – be considered dormant, and certainly a non-dividing cell it may be irreversibly injured or operationally dead. However, the presence of molecular signals (e.g. 16S) in samples from which nothing (or many fewer colonies or OTUs) may be recovered by culture is certainly an indication of the possibility of resuscitation, and hence dormancy. The referee is entirely correct that we had missed John McKinney’s recent and very relevant work, and we mention it accordingly.

- "Alternative interpretations of the presence of bacterial 16SrDNA sequences in blood when culture fails to detect the organisms from which they derive, include the presence of dead, injured or moribund cells. If they are shown to be repeatedly present then they must either be able to persist in the face of clearance mechanisms or be supplied at a rate equal to their clearance; both seem equally plausible to the dormancy explanation to me. Moreover, why the first three explanations offered for “Not-yet-cultured” should apply to environmental bacteriology but not to clinical samples escapes me."

The referee is entirely correct with regard to the last sentence, and the whole point (or at least a major theme) of our review is precisely that what is well established in environmental microbiology has had much less impact in clinical microbiology (referee 2 makes this exact point, even more explicitly). We agree that in a steady state such cells must be supplied at a
rate equal to that of their clearance, and that the fact that clearance is lower than probably expected implies a significant ability to evade the innate and adaptive immune systems. We also take it that for common organisms (not very slow growers such as certain mycobacteria) the former rates must be much lower than those typically attainable in laboratory cultures, else we would have classical sepsis. We have added a few comments on these issues accordingly, in the section entitled ‘Generalised failure of classical techniques to detect dormant bacteria in clinical microbiology’.

- "I am led to the conclusion that the authors have chosen to label evidence for discrepancies between culture and nucleic acid detection of bacteria in blood to give their hypotheses a simple headline. I have no problem with the proposal that human blood and tissues classically considered sterile in the absence of overt symptoms of infection are frequently exposed to bacteria and bacterial products that in many cases contribute to serious chronic disease. However, I consider the burden of available evidence currently provides many potential explanations within the field of microbiomics/metagenomics in contrast to the dormancy hypothesis offered here. Further, I feel this broad application of dormancy to bacterial phenotypes which, even in the case of Rpf dependency, have not been shown to result from a programme of gene expression that could be considered as differentiation, diminishes the value of the term. Indeed there remains no direct proof that dormancy of Mycobacterium tuberculosis underpins what we call latent tuberculosis infection and it is not essential to the observed clinical or pathological pattern, notwithstanding the widespread acceptance of this view by most researches, including me.

I am not fundamentally opposed to the ideas presented by Kell and colleagues but I do not think they are assisted by lack of attention to the contradictions I have identified above."

All of the above is entirely fair, and we do not disagree. We hope that the changes we have now made to the ms to weaken the ostensible claims (and misplaced synonymies) now meet the referee’s approval. For instance we have stressed that while the presence of suitable molecular sequences (e.g. 16S) implies that it is worth seeking to resuscitate the organisms from which it came, an absence would imply that it is not. A success in resuscitating organisms from a sample that initially appeared sterile would from our operational definition imply that those ones were indeed dormant, and we’d like to think that this had now been clarified.

- "Finally I come to the iron dysregulation hypothesis and its pro-inflammatory consequences. It is beyond my expertise to comment on the plausibility of the inorganic chemistry deployed here or to review the evidence relating to more than a fraction of the conditions listed. The importance of the struggle between pathogens and host for access to iron is beyond question. When I entered the medical field of infectious disease it was fully recognised that depriving bacteria from iron was a potential therapeutic angle and indeed iron chelation was studied. Desferioxamine, a widely used agent in iron overload, was investigated and found to effectively deliver iron to the pathogen and the approach was set aside. More recently this agent has been identified as a major risk factor in serious fungal infection and guidance specifically recommends its avoidance. Newer agents seem not to suffer from this problem and the approach deserves renewed attention. However, I would not underestimate the ability of pathogens to outwit our pharmaceutical industry in the battle to sequester iron. While there are reasons beyond the host–pathogen tug-of-war for iron to consider chelation as a therapeutic option, the potential for adverse effects is significant and I think the
The point about desferrioxamine is well made (and we mention it, with citations), but the molecule is of course in fact a natural prokaryotic siderophore, from *Streptomyces pilosus*. We have replaced the term ‘shocking’ with something more suitable.

- “Focussing briefly on the specific diseases cited and their relation to bacterial exposure in one form or another, I find that evidence cited frequently rests on what can be considered “fringe” hypotheses that have little currency in their respective fields. This is not to discourage their continued pursuit but it does weaken the strength of the authors’ argument when investigation of the supporting literature frequently leads to papers that are given little credence in the specialist field. Of course “cave Helicobacter” must remain on the table. But there, an accidental technical breakthrough led to an avalanche of convincing laboratory and clinical data.”

It is probably a philosophical distraction to rehearse how often in science something outside the mainstream is blocked for many years by ‘vested interests’. However, we may as well mention Peyton Rous, whose discovery of a viral cause of certain cancers was sidelined for decades (he received a Nobel prize when he was 87, 40 years after first being nominated [https://en.wikipedia.org/wiki/Francis_Peyton_Rous](https://en.wikipedia.org/wiki/Francis_Peyton_Rous)). Closer to (prokaryotic) home, Barry Marshall has edited a book (Marshall BJ (ed.): *Helicobacter pioneers: firsthand accounts from the scientists who discovered helicobacters*. Melbourne: Blackwell, 2002.) whose invited contributors had all long recognised a bacterial cause of ulcers and treated their patients accordingly, on the simple grounds that the antibiotics worked! Of course Marshall and Warren (and the wider world) knew nothing of this at the time of their discovery of *H. pylori*. Under these circumstances (as here) we rely on the overall weight of evidence (as much as its place of publication) to support our views. In Philosophy of Science circles this bolstering of a view via overlapping circles of self-consistent reasoning and data is referred to as ‘coherence’. Accordingly, in this sense, we have tried to make this a coherent story, and rehearse this point in the concluding section.

- “In summary Kell, Potgieter and Pretorius have produced an interesting read which bring many important ideas to our attention. I am not convinced of the breadth of conditions to which they argue their ideas are applicable and I await with interest, demonstration of of how they may be practically pursued and some selected definitive proofs that iron-driven inflammatory disease is as important as they claim.”

We have no further comments at this stage. Many thanks again for a very thoughtful review.

**Competing Interests:** No competing interests were disclosed.