RESEARCH ARTICLE

Using Akaike’s information theoretic criterion in population analysis: a simulation study [version 1; referees: 2 approved with reservations, 1 not approved]

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Abstract
Akaike’s information-theoretic criterion for model discrimination (AIC) is often stated to “overfit”, i.e., it selects models with a higher dimension than the dimension of the model that generated the data. However, when no fixed-dimensional correct model exists, for example for pharmacokinetic data, AIC, or its bias-corrected version (AICc) might be the selection criterion of choice if the objective is to minimize prediction error. The present simulation study was designed to assess the behavior of AICc when applying it to the analysis of population data, for various degrees of interindividual variability. The simulation study showed that, at least in a relatively simple mixed effects modeling context, minimal mean AICc corresponded to best predictive performance even in the presence of large interindividual variability.
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How to cite this article: Olofsen E and Dahan A. Using Akaike’s information theoretic criterion in population analysis: a simulation study [version 1; referees: 2 approved with reservations, 1 not approved] F1000Research 2013, 2:71 (doi: 10.12688/f1000research.2-71.v1)

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Grant information: This work was funded by institutional resources.
The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: No competing interests were disclosed.

Introduction

Population data consist of one or more measurements in two or more individuals. Such data can be characterized by mixed-effects models, where the mixed effects consist of fixed and random effects. Fixed effects are, for example, the times at which the measurements are obtained, and covariates such as demographic characteristics of the individuals. When mixed-effects models are fitted to population data, the question arises how many of those effects should be incorporated in the model. This is the so-called problem of variable selection.1

One strategy is to observe the change in goodness-of-fit by adding one more parameter and testing the significance of that change. In the maximum likelihood approach, the objective function value (OFV), being the minus two logarithm of the likelihood function, is minimized. To attain a $p$-value of e.g., 0.05 or less, the decrease in OFV, when adding one parameter, should be 3.84 or more.3

Another strategy is to apply Akaike’s information theoretic criterion (AIC), which can be written as

$$\text{AIC} = \text{OFV} + 2 \cdot D, \quad (1)$$

where $D$ is the number of parameters in the model.2 The model with the lowest value of AIC is considered the best one. In the case of just adding one parameter, the OFV needs to decrease only 2 points or more to be incorporated in the model, so the associated $p$-value > 0.05 seems too high to justify this strategy.4

When additional model parameters are incorporated, the significance of one model parameter might change, but the interpretation of AIC does not. However, when multiple significance tests are performed, the significance level of each individual test should be corrected to a lower value, so a decrease of 2 points for one parameter does again seem to be too low.4

Even if the strategy of using AIC leads to optimal variable selection, the question arises if this is also the case when using mixed-effects models. In theory, the model that is best according to AIC is the one that minimizes prediction error;3 and this is also true for a mixed effects model when predicting data for individuals for which no data have been obtained so far.3

In the literature, simulation studies have assessed the performance of AIC in selecting the model with the lowest prediction error, but to our knowledge these were never done for population data. In this article, we will define a toy pharmacokinetic model and observe the performance of AIC when adding fixed effects to this model, as well as when adding interindividual variability.

Methods

A hypothetical pharmacokinetic model

Consider the following function $y(t)$, an infinite sum of exponentials, and its relationship with a (negative) power of time:

$$y(t) = \int_0^\infty \exp(-\lambda t) \, d\lambda = -\frac{1}{t} \exp(-\lambda t) \bigg|_0^\infty = \frac{1}{t} \quad \text{for} \quad t > 0. \quad (2)$$

Figure 1A shows that this function looks like a typical pharmacokinetic profile after bolus administration. This model is to be regarded as a toy model, because we do not expect it to adequately describe pharmacokinetic data, although variations of power functions of time have been shown to fit pharmacokinetic data well. Here we will use the fact that if we approximate $y(t) = 1/t$ by the following sum of $M$ exponentials with $K$ nonzero coefficients $\alpha$ and $M$ fixed parameters $\lambda$ (as chosen in the next subsections):

$$\hat{y}(t_j; \alpha, \lambda) = \sum_{m=1}^M \alpha_m \exp(-\lambda_j t_j), \quad (3)$$

that with $M$ time instants $t_j$ we would need no less than $K = M$ exponentials to obtain a perfect fit. Moreover, with noisy data, it might be that for $K < M$ an optimal fit is obtained in the sense that the associated prediction error of the model is minimal. Figure 1B shows how eleven (in this case error-free) samples from this function can be approximated by sums of exponentials.

Data simulation

In the following, the time instants $t_j, j = 1, \ldots, M$, centered around 1, were chosen within $[1/t_{\max}, t_{\max}]$ according to

$$t_j = \left(\frac{j}{M + 1 - j}\right)^\gamma, \quad (4)$$

with $\gamma = \log(t_{\max}) / \log(M)$; $t_{\max}$ was set to 100 (see the time axis of Figure 1B for an example with $M = 11$). Simulated data were generated via

$$y(t_j) = \frac{1}{t_j} (1 + \epsilon_j), \quad (5)$$

where $\epsilon_j$ denotes Gaussian measurement noise with variance $\sigma^2$. The $M$ time constants $\lambda$ were fixed according to $\lambda_m = 1/t_{\max}, m = 1, \ldots, M$. In this setting the model eq. (3) can be fitted to simulated data using weighted linear least squares regression, with weight factors $\omega(t_j) = 1/t_j$ (note that no precaution is needed against $\epsilon \leq -1$).
Simulation scripts
23 Data Files
http://dx.doi.org/10.6084/m9.figshare.157246

Population data simulation and modeling
Population data consisting of \( N \) individuals were simulated via

\[
y_i(t_j) = \frac{1}{t_j} \cdot (\exp(\eta_i) + \epsilon_{ij}) \quad \text{with} \quad i = 1, \ldots, N, \tag{6}
\]

where \( \eta_i \) denotes interindividual variability with variance \( \omega^2 \).

The nonlinear mixed effects model for the population data was then written as:

\[
\hat{y}_i(t_j; \alpha, \lambda) = \sum_{m=1}^{M} \alpha_m \exp(-\lambda_m t_j + \eta_i). \tag{7}
\]

Note that with \( N > 1 \), a perfect fit is no longer obtained with \( K = M \), because the \( \epsilon_{ij} \) are generally different for different \( i \) (individuals).

Statistical analysis
Simulation data were generated via eq. (6), with random generators in R. Model fitting was also done in R, with function “lm()” from package “stats”, except for nonlinear mixed-effects model fitting for simulated data with \( \omega^2 > 0 \), which was done in NONMEM version 7.3 (beta version a6.5)\(^4\). Parameters \( \alpha \) (see eq. (7)) were not constrained to be positive, so that it was not possible for parameters to become essentially fixed to zero, reducing the dimensionality of the model. Prediction error \( (\nu^2) \) was calculated with

\[
\nu^2 = \frac{1}{N \cdot M} \sum_{i=1}^{N} \sum_{j=1}^{M} \left( \frac{z_i(t_j) - \hat{y}_i(t_j)}{w(t_j)} \right)^2, \tag{8}
\]

using predictions based on eq. (7) with the random effects \( \eta_i = 0 \), and validation data \( \epsilon(t) \) also generated via eq. (6), but with different realizations of \( \epsilon_i \) and \( \eta_i \). The objective function OFV was also calculated at the estimated parameters using the validation data, denoted OFV, which should on average be approximately equal to Akaike’s criterion (see Supplementary material). OFV was compared with AIC and also with Akaike’s criterion with a correction for small sample sizes (AICc)\(^4\):

\[
AIC_c = \text{OFV} + 2 \cdot D \cdot \left( 1 + \frac{D + 1}{N \cdot M - D - 1} \right) \tag{9}
\]

The above criteria were normalized by dividing them by the number of observations, and averaged over 1000 runs (unless otherwise stated; and runs where NONMEM’s minimization was not successful were excluded). For plotting purposes, 95% confidence intervals or confidence regions for means were determined using R’s packages “gplots” and “car”, under the assumption that averages over 1000 variables are normally distributed.

Selection of parameter values
Simulation parameters \( M \) and \( \sigma^2 \) are expected to determine the number of exponentials \( K \); if \( M \) increases and/or \( \sigma^2 \) decreases, \( K \) will increase. Without inter-individual variance, so \( \omega^2 = 0 \), the information in the data increases as \( N \) increases, so that \( K \) is also expected to increase. With \( N = 2, M = 11 \) and \( \sigma^2 = 0.5 \), pilot simulations indicated a \( K = 4 \). When \( \omega^2 > 0 \), the prediction error will increase, but it is less easy to predict what its

Figure 1 A: function \( y(t) = 1/t \), and B: approximations obtained by fitting six and three exponentials to the depicted eleven samples. Note the log-lin and log-log scales for panels A and B, respectively. Time has arbitrary units.
effect will be on $K$. For $\omega^2$ values of 0, 0.1, and 0.5 were selected – values that are encountered in practice. Because there is only one random effect in the mixed effects model, the relatively low number of individuals $N = 5$ was selected.

For a certain choice of $M$, there are $2^M - 1$ possible combinations of $\lambda$s to choose for the terms $\exp(-\lambda_m t)$ in the sum of exponentials (excluding the case of zero exponentials). Because accurate evaluation of all models at different parameter values is not feasible with respect to computer time, the set of possible combinations was reduced to one with evenly spaced $\lambda$s. Table 1 gives an example for the case $M = 11$.

**Results**

Figure 2 shows the averaged prediction error versus number of exponentials for all possible choices of $\lambda$, with $N = 2$, $M = 11$, $\sigma^2 = 0.5$, and $\omega^2 = 0$. From the figure it is clear that prediction error may indeed increase if the number of exponentials selected is too large. The bigger solid circles correspond to the models chosen in Table 1; in general the evenly spaced selection of exponents resulted in models with the smallest prediction error.

Figure 3 shows simulation results using the model set defined in Table 1, starting from $K = 4$, with parameters $N = 5$, $M = 11$, $\sigma^2 = 0.5$, and $\omega^2 = 0$. The model with $K = 6$ exponentials had both minimal mean OFV, and minimal mean AIC, (and minimal prediction error $\nu^2$ (not shown)). With $N = 5$, $M = 11$, there are still visible differences between AIC, and AICc; although AIC would in this case also select the optimal model, AIC appears to favor more complex models. Note that the sizes of the confidence intervals and confidence regions can be made arbitrarily small by choosing the number of runs higher than the selected number of 1000 (at the expense of computer time).

Figure 4 shows simulation results with $\omega^2 = 0.1$; mixed-effects analysis was used to fit the population data. The main difference with the results of data with $\omega^2 = 0$ is the overall increase in OFV, and AIC. The optimal number of exponentials remained $K = 6$.

Figure 5 shows simulation results with $\omega^2$ set at the higher value of 0.5. The main differences with the results of data with $\omega^2 = 0.1$ are again the overall increase in OFV, AIC, and prediction error, and also in the variability in the prediction error. The optimal number of exponentials remained $K = 6$, although AIC begins to favor the models with larger $K$ (a simulation with $N$ increased to 7, both OFV, and AIC, favored larger models; data not shown).
Table 1  Selecting $K = 1, ..., M = 11$ evenly spaced rate constants from $\lambda$: 0 and 1 denote $\alpha_m$ to be fixed to zero, and a free parameter to be estimated, respectively (see eq. (7)).

<table>
<thead>
<tr>
<th>$K$</th>
<th>$m$ : 1 2 3 4 5 6 7 8 9 10 11</th>
</tr>
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<tbody>
<tr>
<td>1</td>
<td>0 0 0 0 0 0 1 0 0 0 0</td>
</tr>
<tr>
<td>2</td>
<td>1 0 0 0 0 0 0 0 0 0 1</td>
</tr>
<tr>
<td>3</td>
<td>1 0 0 0 0 1 0 0 0 0 0</td>
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<td>4</td>
<td>1 0 0 1 0 0 0 0 1 0 0</td>
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<td>5</td>
<td>1 0 1 0 1 0 1 0 1 0 0</td>
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<tr>
<td>6</td>
<td>1 1 0 1 0 1 0 1 1 0 1</td>
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<td>7</td>
<td>1 1 0 1 1 0 1 1 0 1 1</td>
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<td>8</td>
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<td>9</td>
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<td>10</td>
<td>1 1 0 1 1 1 1 1 0 1 1</td>
</tr>
<tr>
<td>11</td>
<td>1 1 0 1 1 1 1 1 0 1 1</td>
</tr>
</tbody>
</table>

Figure 3  Mean OFV, as a function of minus two log likelihood (-2LL), the number of exponentials, AIC and AIC$_c$, (top four panels), and AIC and AIC$_c$ as a function of the number of exponentials (lower two panels), averaged over 1000 runs, $N = 5$, $M = 11$, $\sigma^2 = 0.5$, $\omega^2 = 0$. The dashed lines represent the theoretical values for an infinite amount of data (see Supplementary material). Error bars and ellipses denote 95% confidence intervals and confidence regions, respectively. Each solid line in the middle panels denotes the line of identity.
Discussion

With the objective of creating a simulation context resembling pharmacokinetic analysis where concentration data are approximated by a sum of exponentials, the toy model \( y(t) = \frac{1}{t} \) was chosen. In this setting, reality - the reality of the toy model – is always underfitted. When mixed effects models were fitted to simulated data, mean AICc was approximately equal to the validation criterion mean OFVv, and their minima coincided. With large interindividual variability, mean expected prediction error \( (\nu^2, \text{see eq. (8)}, \text{with random effects fixed to zero}), \) was less discriminative between models, so that it became less suitable as a validation criterion.

Akaike’s versus the conditional Akaike information criterion

Vaida and Blanchard proposed a conditional Akaike information criterion to be used in model selection for the “cluster focus”. It is important to stress that the cluster focus as they defined is the situation where data are to be predicted of a cluster that was also used to build the predictive model. In that case, the random effects have been estimated, and then the question arises how many parameters that required. In our case, a cluster is the data from an individual; AIC was used in the situation of predicting population data consisting of individual data that were not used to build the model. This would seem to be the most common situation in clinical practice. Furthermore, AIC for the population focus is asymptotically equivalent with leave-one-individual-out cross-validation; AIC for the individual focus with leave-one-observation-out cross-validation.

Akaike’s versus the Bayesian information criterion

We chose to perform simulations using the model given by eq. (2) because approximating data with a sum of exponentials is daily practice in pharmacokinetic analysis where data are obtained from “infinitely complex” systems, and we cannot hope to find the “correct” model. The Bayesian information criterion (BIC) is consistent in the sense that it selects the correct model, given an infinite amount of data. The reason that AIC can be used in “real-life” problems is that as the amount of data goes to infinity, the complexity, or dimension, of the model that should be applied should also go infinity. Burnham and Anderson show that it is possible to choose the prior probability distribution for BIC in such a way that it incorporates the knowledge that more complex models should be favored if the amount of data increases, and so that the BIC “reduces” to AIC. In the situation that the correct model set belongs to the set of evaluated models, a selection criterion that both finds the correct model and minimizes prediction error would be preferable - but Yang concluded that this may not be possible.
In pharmacokinetic analysis, it may not really be appropriate to test (using a hypothesis test assuming a $X^2$ distribution for the objective function) whether an added exponential is statistically significant\textsuperscript{12}. Here the hypothesis $H_0$: the data originate from a $K$-exponential model (and $H_0$: the data originate from a higher dimensional model) is almost certain to be false. Furthermore, when taking a low $p$-value, it is also almost certain that the model selected has worse predictive properties. If a model is to be applied in clinical practice, for example for drug administration in a patient never studied before, the model should be as predictive as possible. However, it may be sensible to test whether a certain fixed effect has both a clinically and statistically significant effect, if it is costly to reach a false conclusion, for example in case of increased risks for patients, or in the field of drug development.

**Model selection criterion AIC and predictive performance**

Intuitively, predicting data for an individual that cannot be “individualized” seems problematic because the data are predicted using a random effect $\eta$ set to zero, instead of the value fitting for that individual. However, AIC is related to the expected model output; and for individual data not used in building the predictive model, the expected model of output is obtained with mixed effects set to zero, although nonlinearities may bias expectation - but this is also true for nonlinear models without mixed effects.

Furthermore, it should be noted that minimizing AIC has a more general interpretation, namely optimally capturing the information contained in the data\textsuperscript{4}. Independent or future population data $x$ are not just predicted by $\hat{y}$; also the distributions of the expected random effects $\varepsilon$ and $\eta$ are characterized by $\sigma^2$ and $\omega^2$. That is why OFV is the criterion to be used to assess the predictive performance of a model.

**Regression weights as functions of the model output**

The simulated data were analyzed using weighted (non)linear regression, see eq. (6), where measurement noise was weighted according to the exact function value. In practice, when the weights are unknown, a choice must be made to weight the data according to the measurements or to the model output, depending on which is likely to be the most accurate. To match the latter case, simulated data should be generated (cf. eq. (6)) via

$$y_i(t_j) = \frac{1}{t_j} \cdot \exp(\eta_i) \cdot (1 + \epsilon_{ij}).$$

(10)

The likelihood function and AIC are both still well-defined if the model output $\hat{y}(t) \neq 0$. Prediction errors are to be calculated with

$$\nu^2 = \frac{1}{N \cdot M} \sum_{i=1}^{N} \sum_{j=1}^{M} \left( \hat{z}_i(t_j) - \hat{y}_i(t_j) \right)^2 \frac{1}{\hat{y}_i(t_j)},$$

(11)

where $\hat{y}$ possibly becomes arbitrarily close to zero for less than optimal models, and $\nu^2$ may be based on long-tailed distributed numbers. To be able to compare prediction errors from different models, the weight factors could be chosen identical for all $K$ to the model output of the largest model – see the Supplementary material for further analysis.

**Limitations of the study**

We recognize the following limitations of our study:

- The model contained only one random effect, and therefore the number of random effect (co)variances was fixed to one. While the number of (co)variance parameters should be counted as ordinary parameters\textsuperscript{5}, at least in well behaved situations\textsuperscript{13}, we did not investigate the process of optimizing this part of a random effects model.

- The nonlinearity in the mixed-effects model was simply due to a multiplicative factor $\exp(\eta)$ in the model output. Usually, random effects in pharmacokinetic models have more complex influence on the model output. However, the lognormal nature of $\exp(\eta)$ is a characteristic property of both our toy model and general pharmacokinetic models.

- The characteristics of the exponentials incorporated in the regression models were evenly spaced, and the values of the rate constants $\lambda$ were fixed. We expect that with more freedom in the specification of the set of models, prediction errors with overfitted models may be worse. However, the agreement between AIC, and prediction error should persist.

- We did not evaluate all possible models within their definition, but only those listed in Table 1, and it makes sense to limit the model set to avoid overfitting the data\textsuperscript{4,11}. We did not address how to optimally select the rate constants $\lambda$. Stepwise selection methods have their disadvantages\textsuperscript{12}. With stepwise forward selection, AIC may even perform worse than AIC\textsuperscript{14}.

- We did not evaluate the process of covariate selection. However, the set of exponentials may be viewed as a number of (somewhat correlated) predictors. It is therefore expected that the present findings also hold for other types of covariates.

**Conclusion**

In conclusion, the present simulation study demonstrated that in the presence of inter-individual variability in a relatively simple mixed effects modeling context, minimum mean AIC coincided with best predictive performance.

**Author contributions**

EO performed the numerical analyses, and EO and AD contributed to the interpretation of the results and the
Supplementary material

In the following, we summarize theory on the maximum likelihood approach and AIC relevant to this paper. Suppose the model for measured data \(y_j, j = 1, \ldots, M\) is given by (cf. eq. (5), eq. (6), and eq. (10))

\[
y_j = \hat{y}_j + w_j \cdot \epsilon_j,
\]

where \(\hat{y}_j\) is the model output, \(w_j\) are weight factors, and \(\epsilon_j\) are independent normally distributed with mean zero and variance \(\sigma^2\). The likelihood function \(L\) for this data set is then given by

\[
L(y; \theta) = \prod_{j=1}^{M} \frac{1}{w_j \sigma \sqrt{2\pi}} \exp \left[ -\frac{1}{2} \left( \frac{y_j - \hat{y}_j}{w_j \sigma} \right)^2 \right],
\]

where the set of parameters \(\theta\) contains \(\sigma^2\) and those needed to calculate \(\hat{y}\). The objective function value (OFV) is defined as minus two times the natural logarithm of the likelihood:

\[
\text{OFV} = -2 \log(L(y; \theta)) = \sum_{j=1}^{M} \log(w_j^2) + M \log(\sigma^2)
\]

\[
+ M \log(2\pi) + \frac{1}{\sigma^2} \sum_{j=1}^{M} \left( \frac{y_j - \hat{y}_j}{w_j} \right)^2.
\]

(14)

Note that in writing “OFV”, the data and parameters it depends on have been omitted. Now maximum likelihood is obtained when OFV is minimal; constant terms such as \(M \log(2\pi)\) may then be discarded (for example, in NONMEM’s calculation of the objective function). The minimum is attained for certain values of parameters of \(\hat{y}\), and for the parameter value of \(\sigma^2\), when the derivative of \(\text{OFV}\) with respect to that parameter is zero:

\[
\frac{\partial \text{OFV}}{\partial \sigma^2} = \frac{M}{\sigma^2} - \frac{1}{\sigma^4} \sum_{j=1}^{M} \left( \frac{y_j - \hat{y}_j}{w_j} \right)^2 = 0,
\]

(15)

so the maximum likelihood estimator of \(\sigma^2\) is

\[
\hat{\sigma}^2 = \frac{1}{M} \sum_{j=1}^{M} \left( \frac{y_j - \hat{y}_j}{w_j} \right)^2.
\]

(16)

By substituting this estimate in eq. (14), we obtain

\[
\text{OFV} = \sum_{j=1}^{M} \log(w_j^2) + M \log(\hat{\sigma}^2) + M \log(2\pi) + M.
\]

(17)

By substituting this result in eq. (1), we have

\[
\text{AIC} = \sum_{j=1}^{M} \log(w_j^2) + M \log(\hat{\sigma}^2) + M \log(2\pi) + M + 2D.
\]

(18)

The term \(2D\) arises from the fact that in minimizing the Kullback-Leibler information, i.e., a measure of the distance between reality and the best approximating model, expectations have to be taken over a data space leading to estimates of parameters \(\theta\) (and hence \(\hat{y}\), and possibly \(\omega\) (see below)) and over a second independent data space \(y^d\). So AIC as defined above should on average be approximately equal the value of OFV eq. (14), with estimated values for the parameters and validation data \(z_j\), denoted \(\text{OFV}_v\).
\[
OFV_v = \sum_{j=1}^{M} \log(w_j^2) + M \log(\hat{\sigma}^2) + M \log(2\pi) + \frac{1}{\hat{\sigma}^2} \sum_{j=1}^{M} \left( \frac{z_j - \hat{y}_j}{w_j} \right)^2.
\] (19)

So when OFV and AIC are both minimized, the latter term – the sum of squared weighted prediction errors – should also be minimal. For the plots in this paper, the measures OFV, OFV′, AIC, and AIC′, were normalized by dividing them by the number of data samples. With an infinite amount of data, and \( \hat{\sigma}^2 = \sigma^2 \), the normalized criteria should attain the value of \( \log(\sigma^2) + \log(2\pi) + 1 \).

Note that if the weights \( w \) are taken as the model output of the best model - but since that is unknown at this point - to the output of the largest model.

For population data, the likelihood function is the product across individual marginal likelihoods where the random effects \( \eta \) contained in eq. (13), when \( \hat{y} \) is given by eq. (6), have been integrated out. Usually, these integrals need to be numerically approximated, e.g., as is done here, by NONMEM. So the context of AIC is then also the one where the \( \eta \)s have been integrated out (but with the parameters at their estimated values), which is to be done when all data are acquired. So while the characteristics of the set of (validation) data are optimally captured, this context is different from the case where prediction errors are calculated with the random effects set to zero instead of being integrated out. In that case, the above AIC and OFV criteria do not match, as the components of the likelihood in eq. (13) are no longer independent (they can only be independent if the true values of \( \eta \) for the individuals are also zero). Note however, that from the higher perspective of optimally characterizing a future set of population data, this is a less important case. Finally, it should be noted that the parameter estimates may not be consistent (i.e., do not converge to their true values when the amount of data goes to infinity if the \( \hat{y} \) do not properly account for heteroscedasticity). In the derivation of AIC, it is only required that the likelihood function is maximized; consistency is not required.

References

The title should indeed specify that this work focuses on pharmacokinetics (PK). However I must add that the model function considered is unusual enough that it seems difficult to extend their conclusions to a real PK study analysis.

The abstract is too general and more details should be provided on the simulation study (model function, number of samples, number of subjects, number of random effects) and the results (differences between selection on OFV, AIC and AICc, impact of increasing the random effect variance).

The whole methodology is very well described. But one aspect is missing, as underlined by the other reviewer: the (very direct here) link with the best sum of exponential model and the information in the design. I was not much surprised that K=6 (or 5) exponential got the best AIC when you have 11 evenly spaced samples and the candidate models all had evenly spaced rate constants. Also, why not investigate the performance of BIC (with log(N) and log(NxM))?

Finally, the conclusions are balanced in the sense that the authors have rightly identified the limit of their exercise which is the generalization of their results to a real PK data analysis: only one random effect, no covariance parameters, only slope parameters, etc...

I have read this submission. I believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

**Competing Interests:** No competing interests were disclosed.
modeling. AIC can be an excellent metric for selecting from among a very limited number of models. If used in a stepwise process it can result in all the severe problems that stepwise variable selection has. The authors need to be much more careful about multiplicity and model uncertainty. This needs to be carefully discussed, and the authors would add to the literature if they can derive the maximum number of models that can be compared with AIC before the method breaks down.

I have read this submission. I believe that I have an appropriate level of expertise to state that I do not consider it to be of an acceptable scientific standard, for reasons outlined above.

**Competing Interests:** No competing interests were disclosed.